# SameSpots 2D DIGE – the limitations of current 2D image analysis

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A Nonlinear Dynamics Group presentation



- Progenesis SameSpots V3.0 (later V3.1)
- Prodigy SameSpots V1.0
- TotalLab TL100 and TL120 V2008
- TotalLab 21CFR part 11 module
- Coming soon! Progenesis LC-MS



### SameSpots Approach

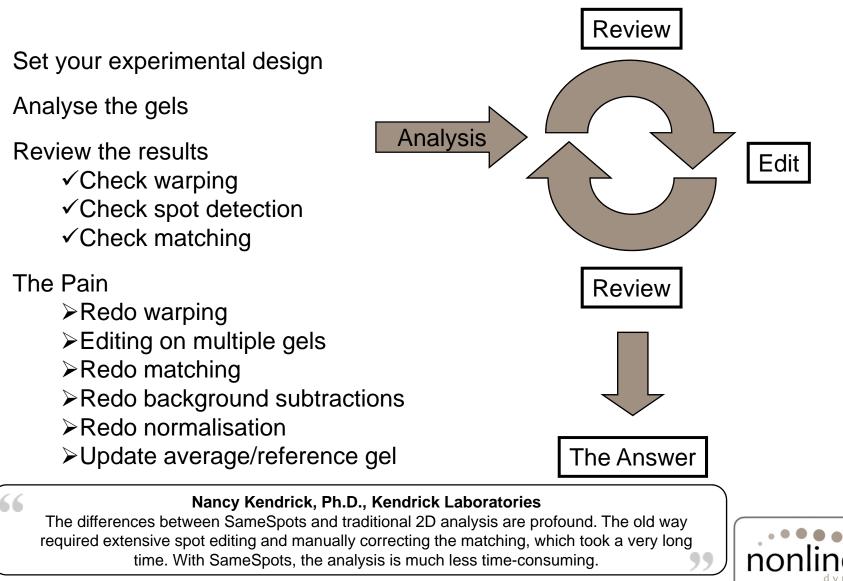
- <u>Simple</u>, highly automated linear Workflow
- Near <u>pixel-level</u> alignment <u>before</u> spot detection.
- Common spot map applied to <u>all</u> <u>gels</u> simultaneously
- <u>100%</u> matching of all spots across every gel in experiment
- <u>No</u> missing values
- Editing on <u>one gel</u> is applied to all others
- Common spot boundary <u>minimises</u> quantification variance.

# **Traditional Approach**

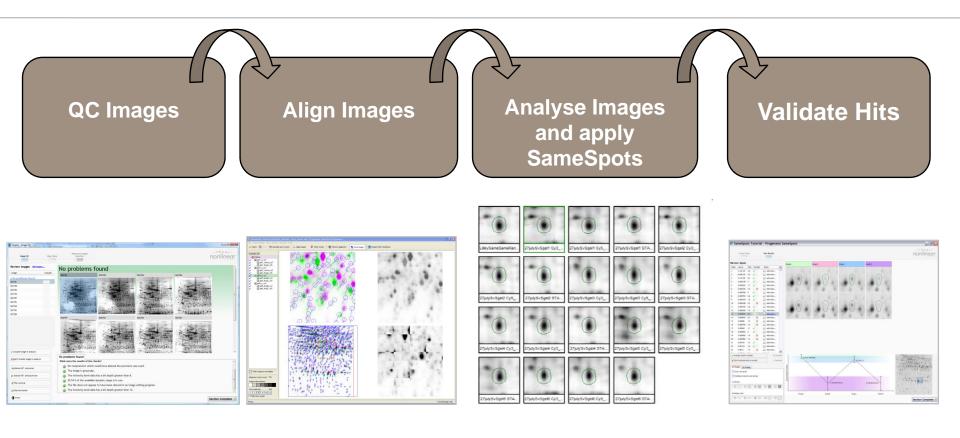
- <u>Complex</u>, manual workflow
- <u>Spot-level</u> alignment by "warping" <u>after</u> spot detection
- Spots detected <u>independently</u> on each gel
- Typically many spot mis-matches even after editing.
- <u>Many</u> missing values typical
- Editing of spots on <u>every gel</u> necessary
- Different spot boundary on every gel – <u>large</u> quantification variance.



## **Traditional Workflow**



#### **Progenesis SameSpots Workflow**



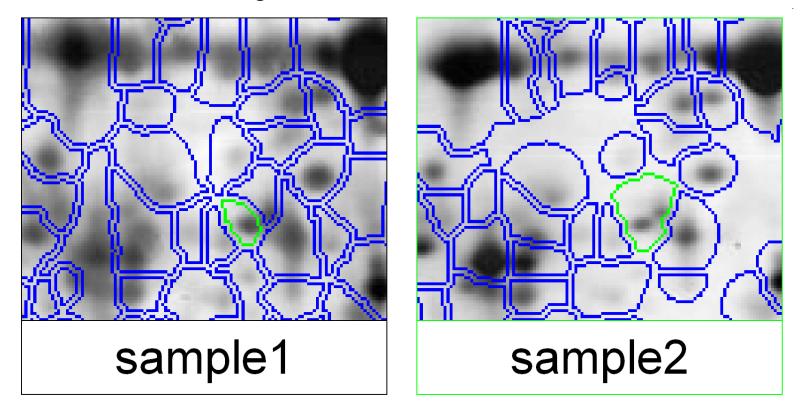
QC review of your images allows identification of problems before you start your analysis. DIGE or single stained gels are aligned before detection to correct for positional variation. Images are analysed and identical spot outlines are transferred to all gels.

Statistics are performed automatically. Simply work down the list and validate hits.



### Common Problems with 2D Image Analysis

Detection and Editing



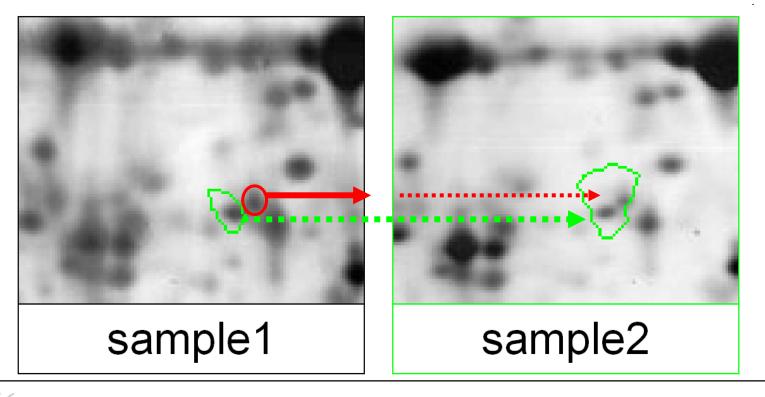
#### Dr. Jean Louis Dacheux, INRA Nouzilly, France

I am a fan of the new SameSpots approach which allows me to easily realign my gels prior to analysis. SameSpots also reduces my hands on time while increasing the quality and the level of confidence in the results produced. I am very excited to be pioneering this technology.

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### Common Problems with 2D Image Analysis

- Detection and Editing
- Matching inaccuracies



#### Dr. Peter Ashton, University of York, UK

The changes we have noted in our analysis have been very encouraging. The ability to perfectly align images dramatically improves matching performance. This enables highly consistent and reliable results to be obtained from all our 2D data sets, even those that previous image analysis packages were unable to accurately quantify.

## Common Problems with 2D Image Analysis

- Detection and Editing
- Matching inaccuracies

#### Statistics – "Missing Values"

Control 2a	Control 2b	Control 2c	Treatment 2d	Treatment 2e	Treatment 2f	
Norm. Vol.	Norm. Vol.	Norm. Vol.	Norm. Vol.	Norm. Vol.	Norm. Vol.	t-test (p)
0.018	0.03	0.008	0.026	-	0.008	0.425
0.021	0.026	0.007	0.036	0.007	0.014	0.273
0.012	-	0.003	0.011	0.015	0.016	0.673
0.018	0.026	-	0.035	0.023	-	0.0627
0.049	0.037	0.032	0.009	0.006	0.014	0.0017
0.004	0.002	-	0.12	0.058	0.13	0.032

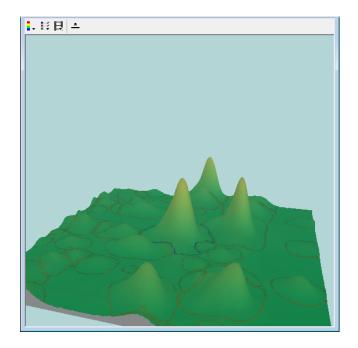
David S. Gibson, PhD, Arthritis Research Group, Queen's University Belfast

With SameSpots we are able to highlight significant changes between patient subgroups with a higher degree of confidence, as previously overlooked missing values are now retained within the data set.

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### What are the goals of 2D expression studies?

- Find the biologically significant expression changes
  - Reliably
  - Objectively
  - Quickly
  - Statistically relevant



Professor Mark S. Baker, CEO, Australian Proteome Analysis Facility Ltd (APAF) SameSpots delivers an alternative starting point for 2D image analysis - offering significant improvements in image matching. The benefit of reliably matching spots across all gel images from the start means you can quickly focus your research efforts on investigating the few spots showing statistically valid changes, which are indicative of potentially interesting expression profiling changes.



# Progenesis SameSpots V3.0 2D Image software- tutorial experiment



#### So how does SameSpots Technology Solve this Problem?

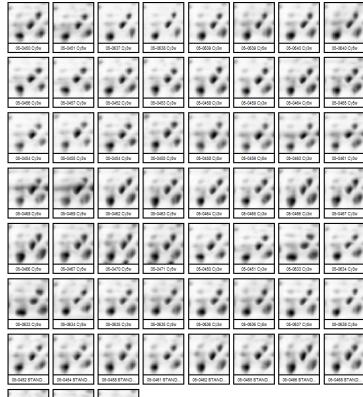


#### Perfect alignment is the KEY to SameSpots

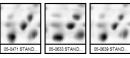
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#### From this...

#### ..to this



**99** 



#### Hans Voshol & Jan van Oostrum, Novartis Institutes for BioMedical Research We thought this level of alignment was impossible!"

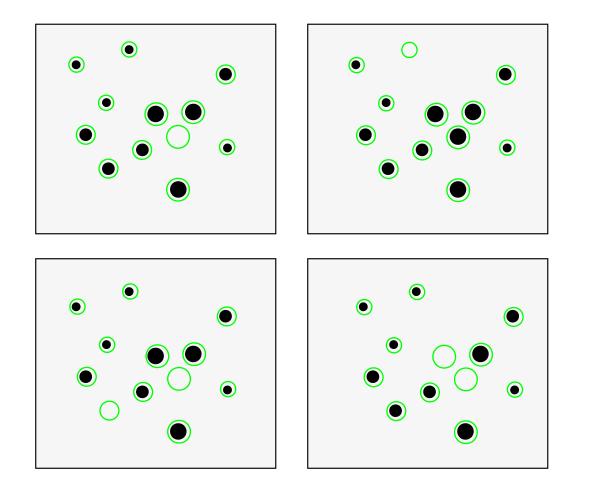


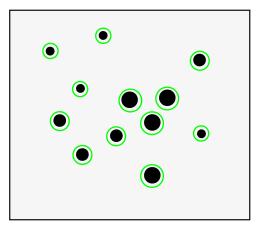
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#### A simplified example of SameSpots detection







### 100% matching, no missing values and a complete dataset

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#### Gel (replicate)

**Doug Hinerfeld, Ph.D., Technical Director, BioMachines Applied Proteomics Group** A well recognized limitation of 2D image analysis is the ability to perform advanced statistical tests on analyzed data. By implementing SameSpots in our analysis workflow, we are achieving complete matching across an experiment, with no missing values, so we can have confidence in any statistical tests we choose to perform.

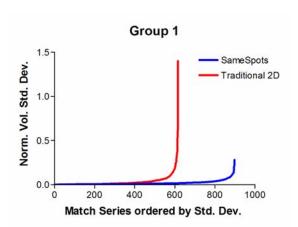


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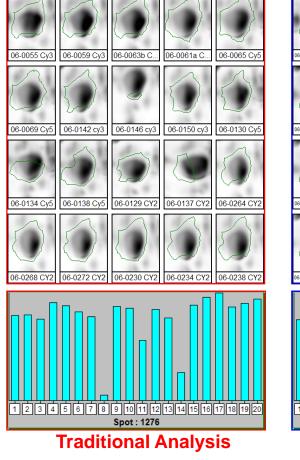
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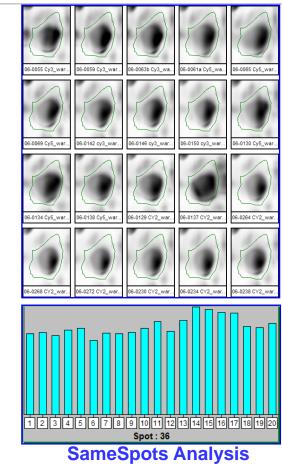
Spot

#### Reduced variability when using SameSpots



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#### Ashutosh Gupta, Rice University

SameSpots is much advanced and ahead of its competitors. It has reduced the analysis time to minutes from hours. Its statistical power is unmatched; with the ability to find same spot on all of the gels on exactly same location, problem of missing values from the gels is gone. SameSpots is definitely better than previous versions and is much advanced than its competitors.



### SameSpots Approach

- <u>Speed</u>: analysis of up to 20 gel set in 2 hours.
- <u>Objectivity</u>: more than 90% correlation of top 100 significant spots.
- <u>Statistical Power:</u> No missing values and low quantification variance = high stat. power
- <u>Simplicity</u>: Linear workflow and high level of automation = simple analysis!

# **Traditional Approach**

- Analysis can take many hours, days, or even weeks!
- Much user dependence due to manual editing and matching stages.
- Missing values and high quantification variance = <u>low</u> statistical power.
- <u>No</u> linear workflow, cycle of reviewing, editing, re-warping, re-matching etc. = complex analysis!

