



Introduction to Flow Cytometry

-- BD FACSCanto II™

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BDBiosciences

Outline

- Basic Concept of Flow Cytometry
- FACSCanto II System Introduction
- Application Examples

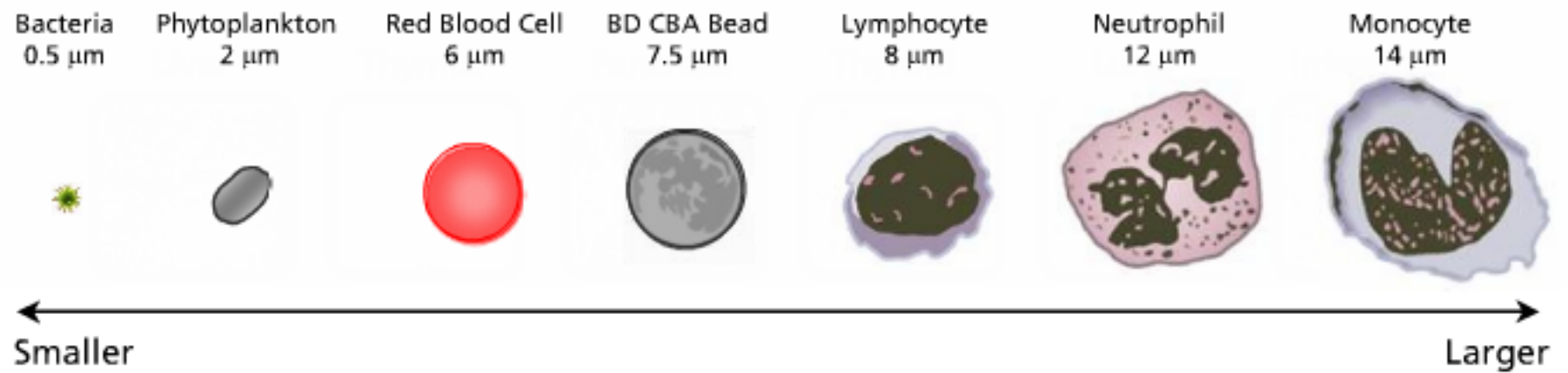
What is Flow Cytometry?

- Flow = Fluid
- Cyto = Cell
- Metry = Measurement

- A variety of measurements are made on cells, cell organelles, and other objects **suspended in a liquid** and flowing at rates of **several thousands per second** through a flow chamber.

Particle Size

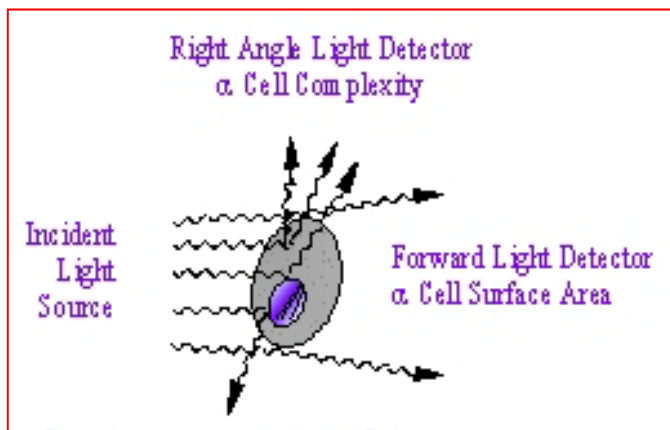
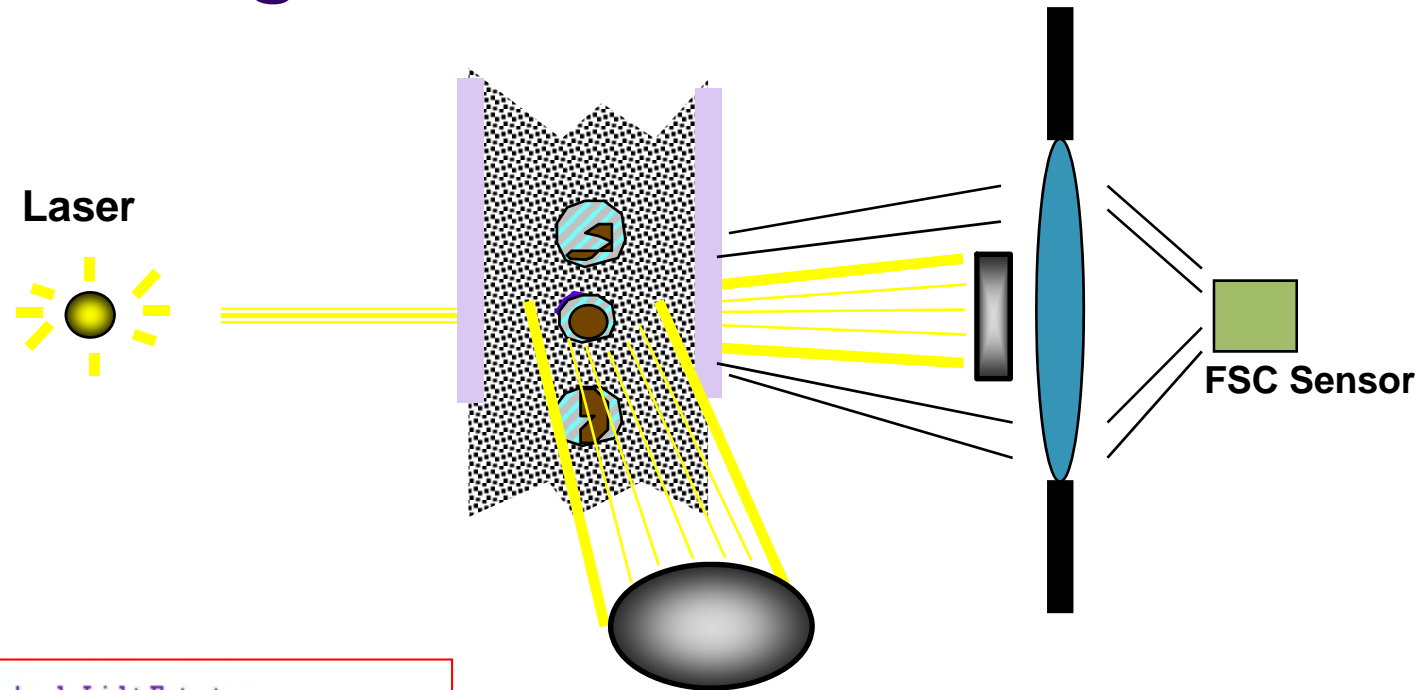
- Detection range: 0.5~50 μ m



What Can a Flow Cytometer Tell Us About a Cell?

- Its relative size (Forward Scatter—FSC)
- Its relative granularity or internal complexity (Side Scatter—SSC)
- Its relative fluorescence intensity

Scatter Light



SSC Sensor

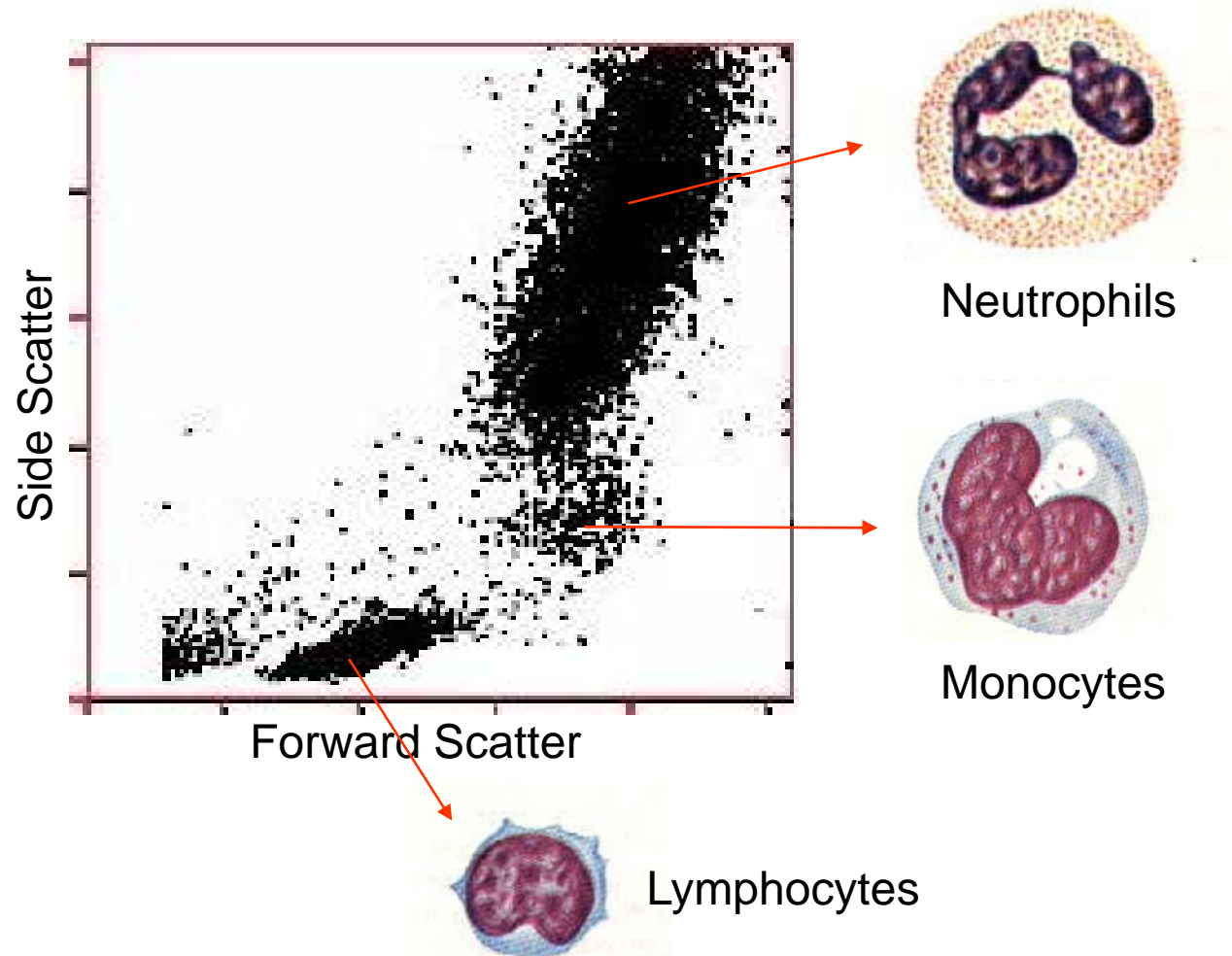
Forward Scatter—diffracted light

- Related to cell surface area
- Detected along axis of incident light in the forward direction

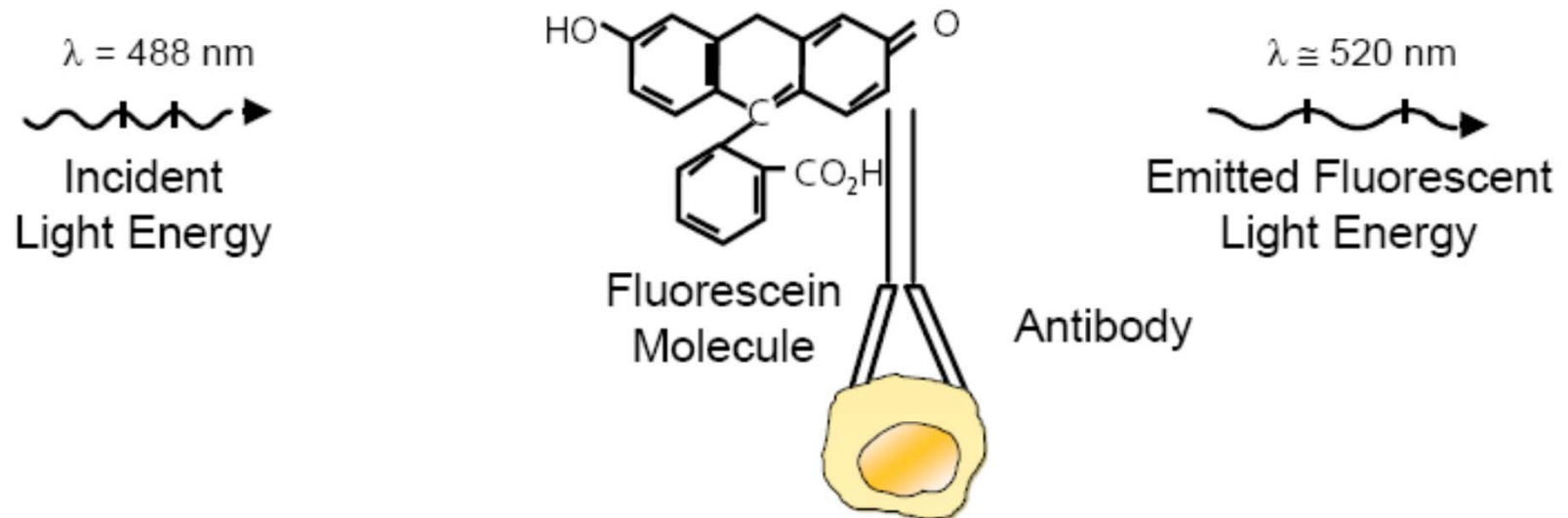
Side Scatter—reflected and refracted light

- Related to cell granularity and complexity
- Detected at 90° to the laser beam

Lysed Whole Blood



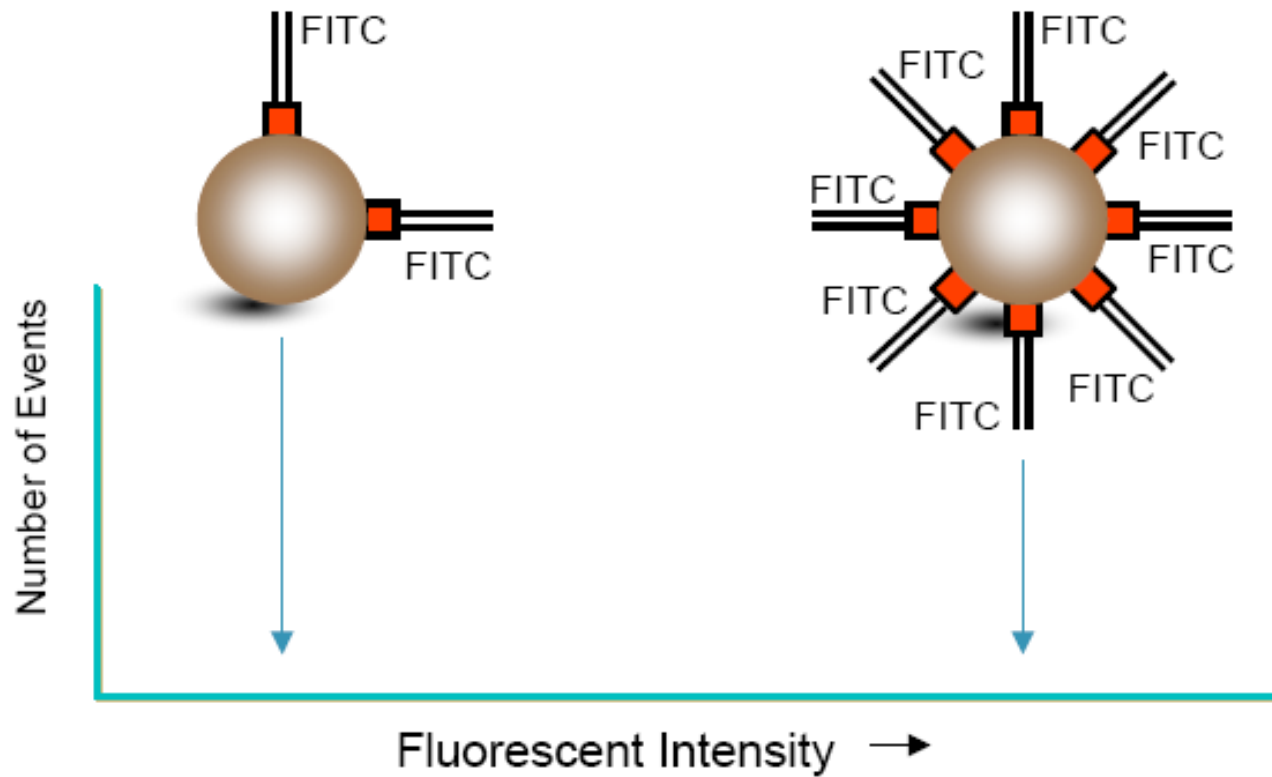
Fluorescence Light



- The fluorochrome absorbs energy from the laser.
- The fluorochrome releases the absorbed energy by:
 - vibration and heat dissipation.
 - emission of photons of a longer wavelength.

Fluorescence

Emitted fluorescence intensity proportional to binding sites



BD FACSCanto II™



Subsystems

Fluidics

To introduce and focus the cells for interrogation.

Optics

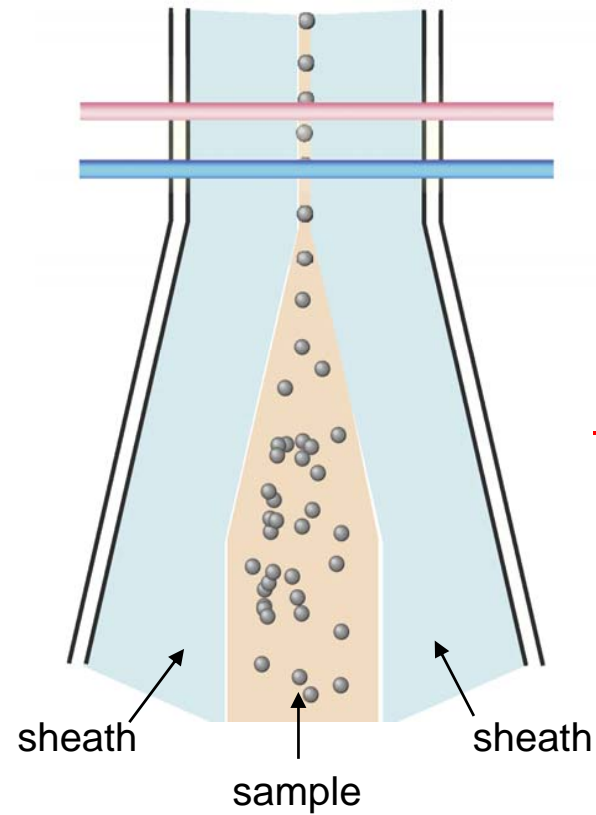
To generate and collect the light signals.

Electronics

To convert the optical signals to proportional digital signals, process the signals, and communicate with the computer.

Sample Flow

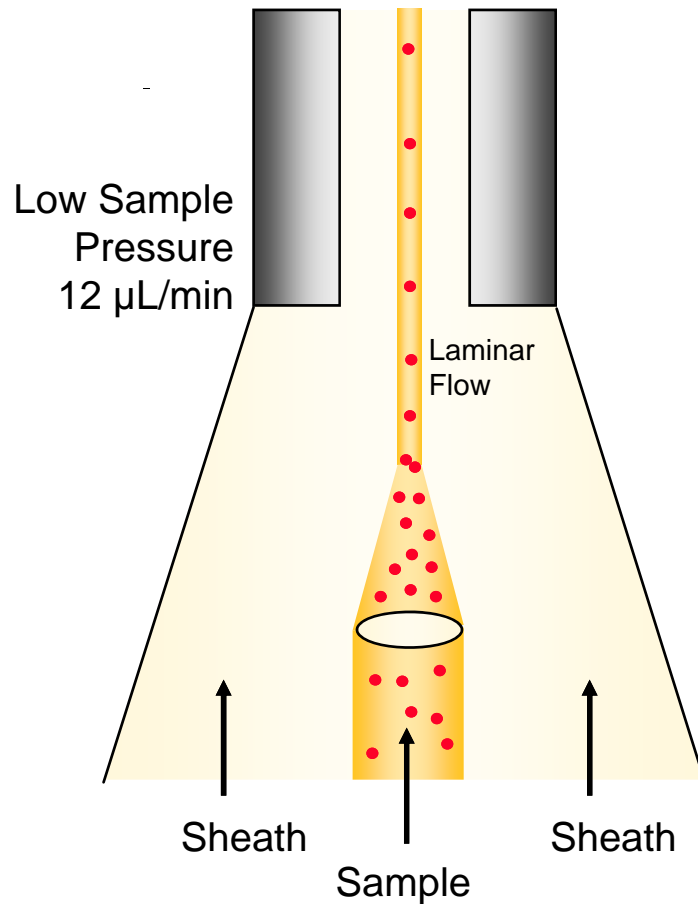
Excitation
Lasers



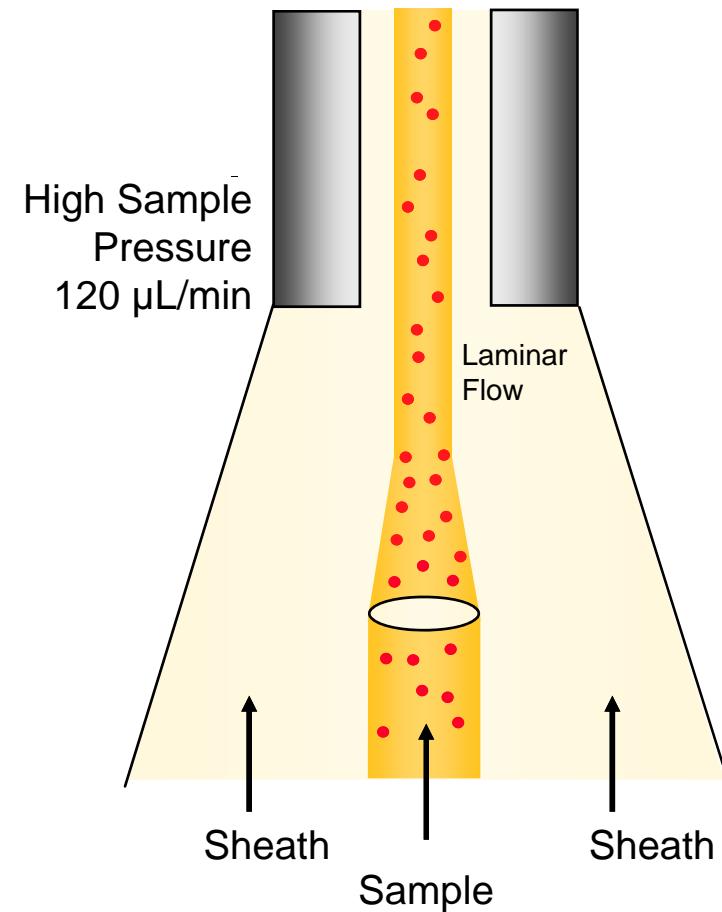
Hydrodynamic Focusing

Sample Differential

Low Differential Pressure



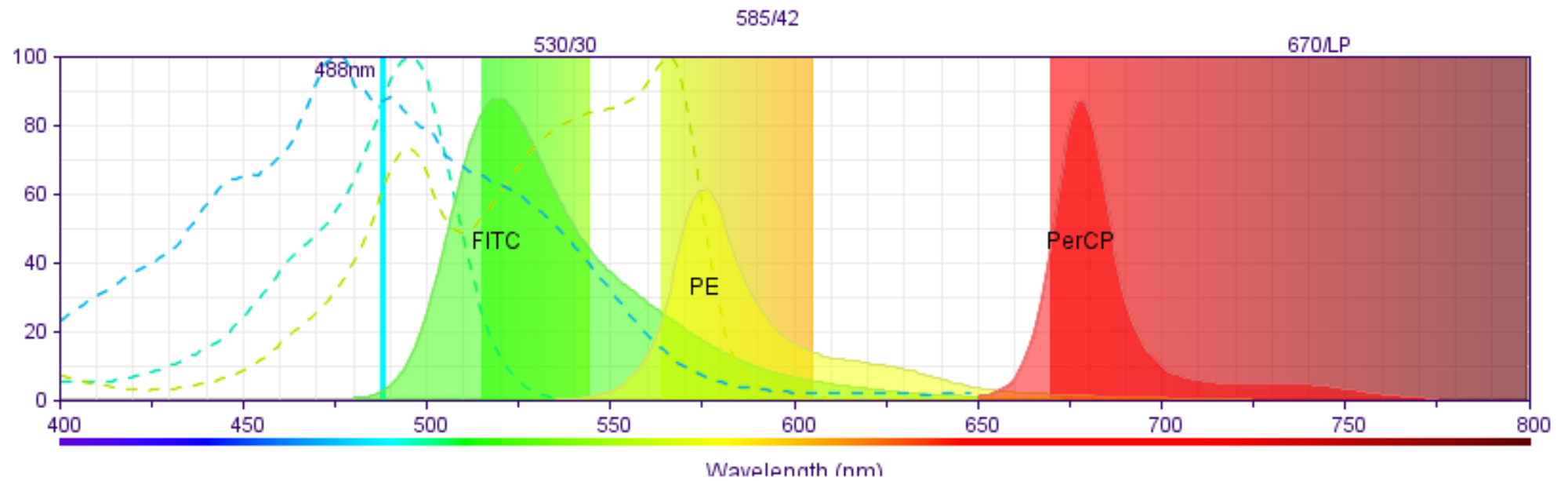
High Differential Pressure



Optics

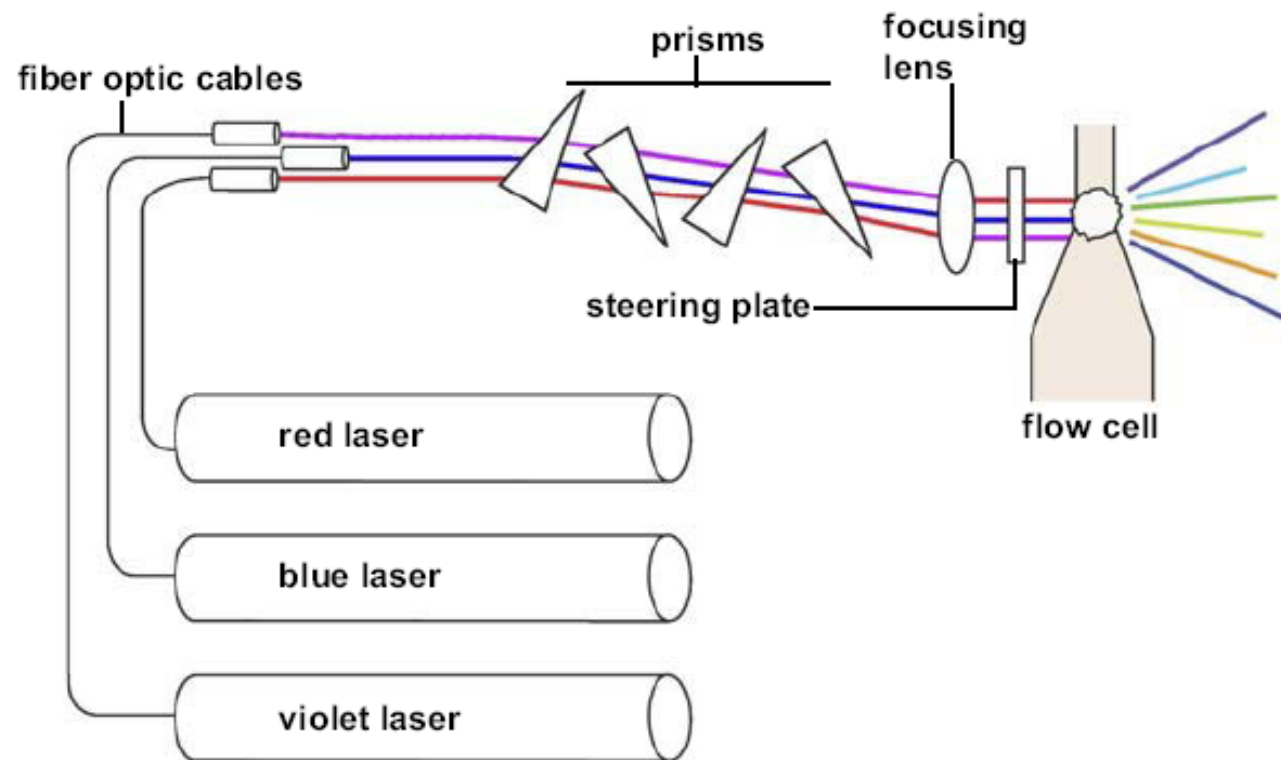
- Excitation optics
 - Lasers
 - Lenses to shape and focus the laser beam
- Collection optics
 - A collection lens to collect light emitted from the article-laser beam interaction
 - A system of optical mirrors and filters to route specified wavelengths of emitted light to designated optical detectors

Fluorochrome Spectra

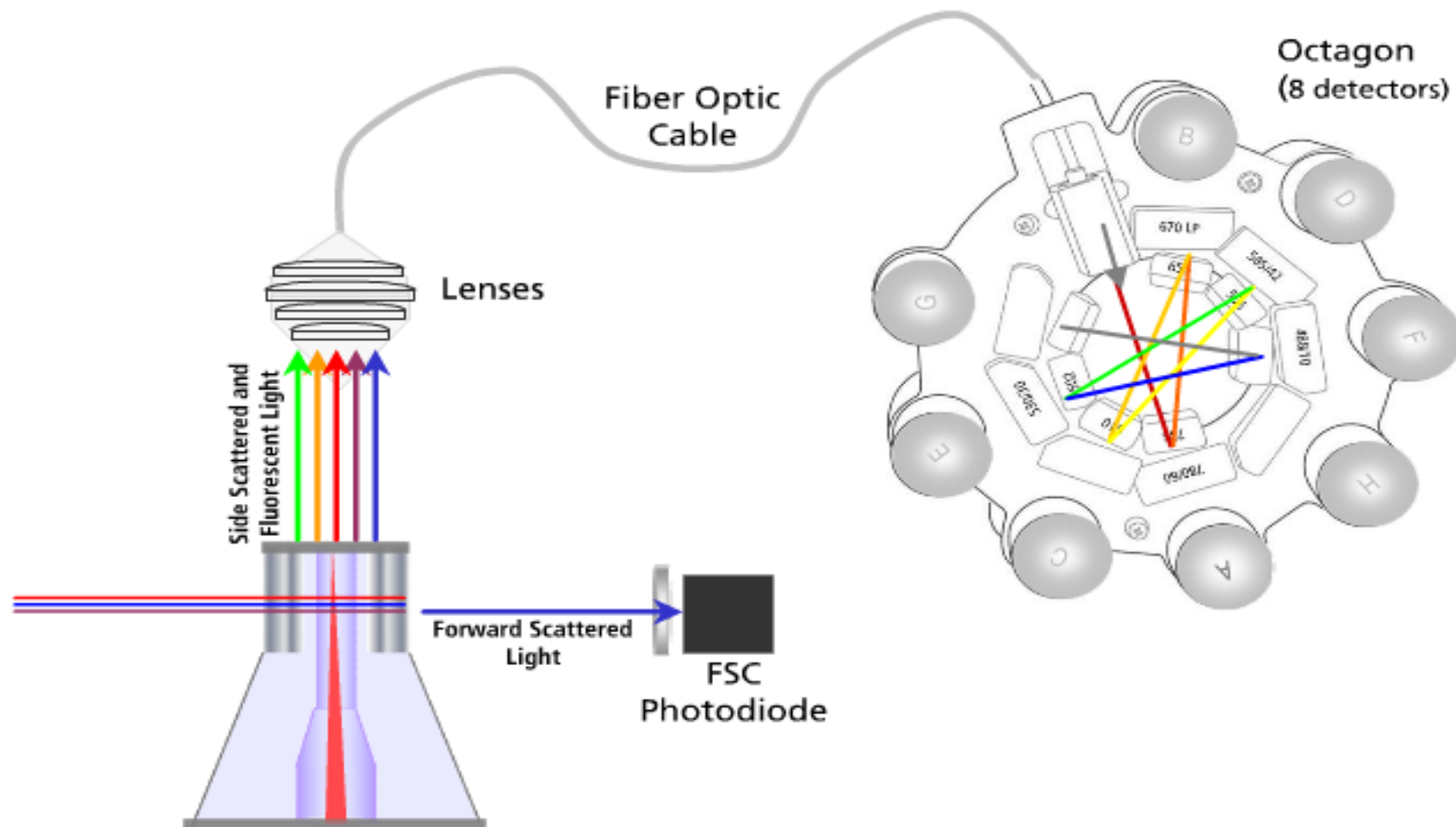


Excitation Optics

- Spatially separated laser beams lower the possibility of fluorescence spillover



Collection Optics



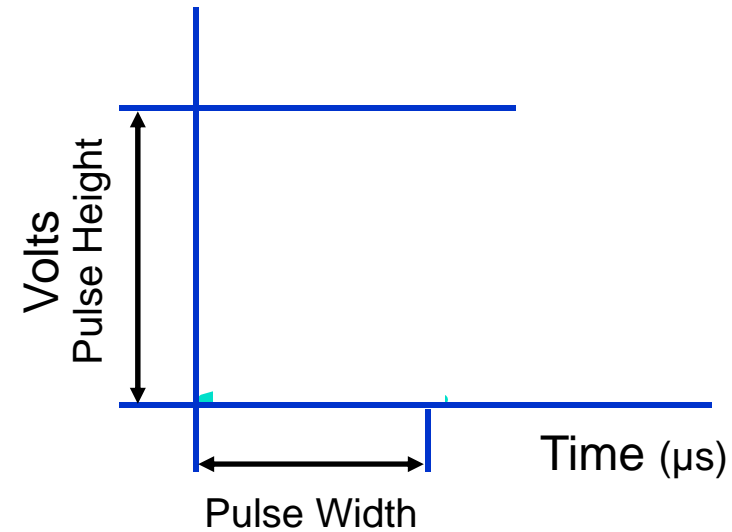
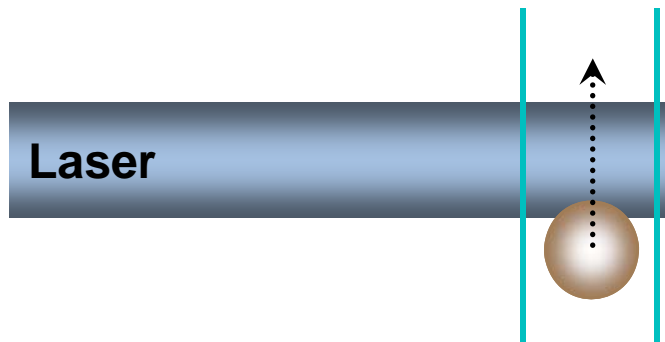
Optics-- Configuration

Lasers	Primary Fluorochrome	PMT	Dichroic Mirror	Bandpass Filter	Other Fluorochrome
488 nm (blue)	Side Scatter	F	none	488/10	
	FITC	E	502LP	530/30	GFP
	PE	D	556LP	585/42	PI
	—	C	610LP	blank optical holder	
	PerCP or PerCP-Cy5.5	B	655LP	670LP	PI, PE-Cy5.5, 7-AAD
	PE-Cy7	A	735LP	780/60	
633 nm (red)	APC	C	none	660/20	Alexa Fluor® 633
	—	B	685LP	blank optical holder	
	APC-Cy7	A	735LP	780/60	
407 nm (violet)	Pacific Blue™	B	none	450/50	DAPI, Hoechst Dye
	AmCyan	A	502 LP	510/50	Cascade Blue®

Electronics

- PMTs and preamps convert photons to voltage pulses.
- Analog-to-digital converters translate analog signals to proportional digital signals.
- Compute area and height for each pulse.
- Perform compensation and calculate ratios and width.
- An embedded computer interfaces with the computer workstation for data transfer.

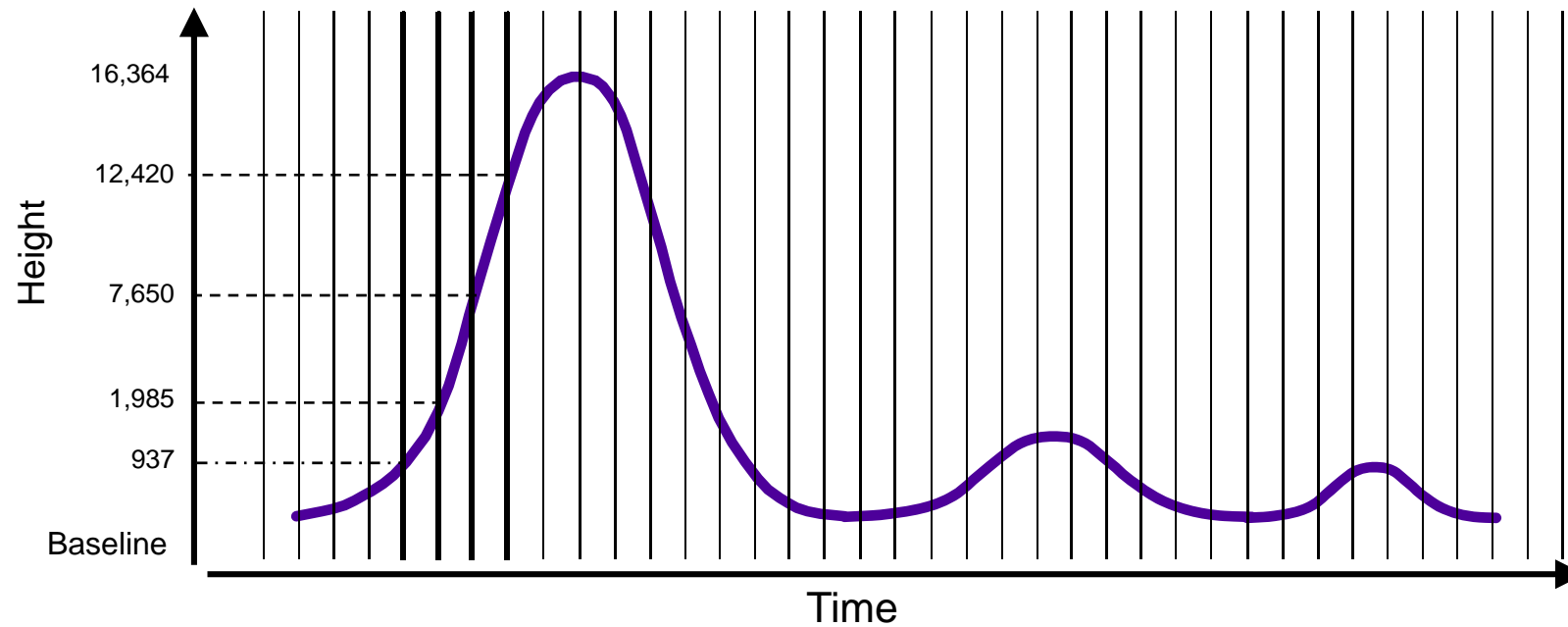
Creation of a Voltage Pulse



Cytometer - FACSCanto (V0041)

Laser	Compensation	Ratio
Status	Parameters	Threshold
Parameter	Voltage
• FSC	407	<input type="checkbox"/> <input checked="" type="checkbox"/> <input type="checkbox"/> <input type="checkbox"/>
• SSC	432	<input type="checkbox"/> <input checked="" type="checkbox"/> <input type="checkbox"/> <input type="checkbox"/>
• FITC	530	<input checked="" type="checkbox"/> <input checked="" type="checkbox"/> <input type="checkbox"/> <input type="checkbox"/>
• PE	473	<input checked="" type="checkbox"/> <input checked="" type="checkbox"/> <input type="checkbox"/> <input type="checkbox"/>
• PerCP-Cy5-5	637	<input checked="" type="checkbox"/> <input checked="" type="checkbox"/> <input type="checkbox"/> <input type="checkbox"/>
• PE-Cy7	778	<input checked="" type="checkbox"/> <input checked="" type="checkbox"/> <input type="checkbox"/> <input type="checkbox"/>
• APC	613	<input checked="" type="checkbox"/> <input checked="" type="checkbox"/> <input type="checkbox"/> <input type="checkbox"/>
• APC-Cy7	641	<input checked="" type="checkbox"/> <input checked="" type="checkbox"/> <input type="checkbox"/> <input type="checkbox"/>

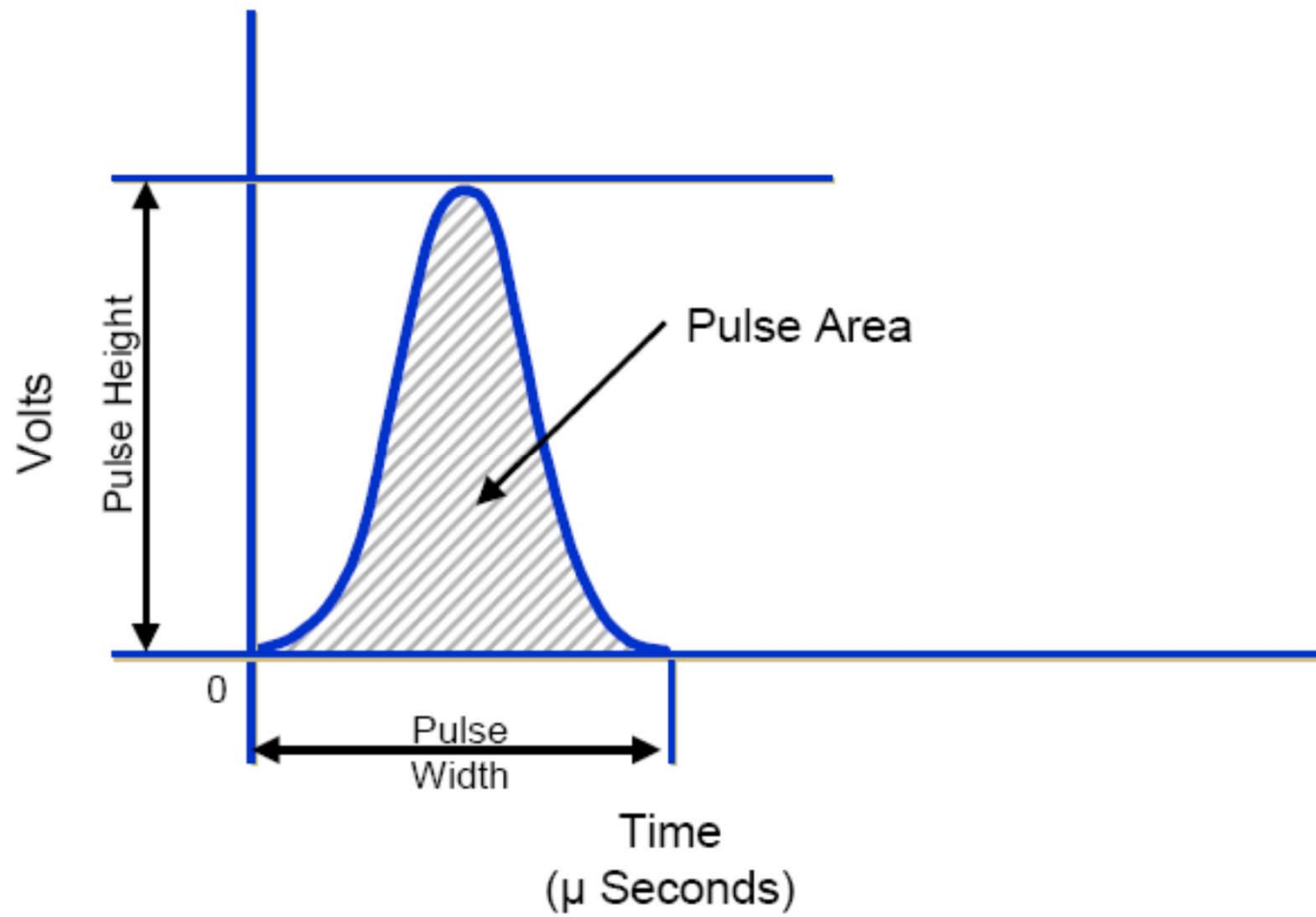
Analog-to-Digital Converter



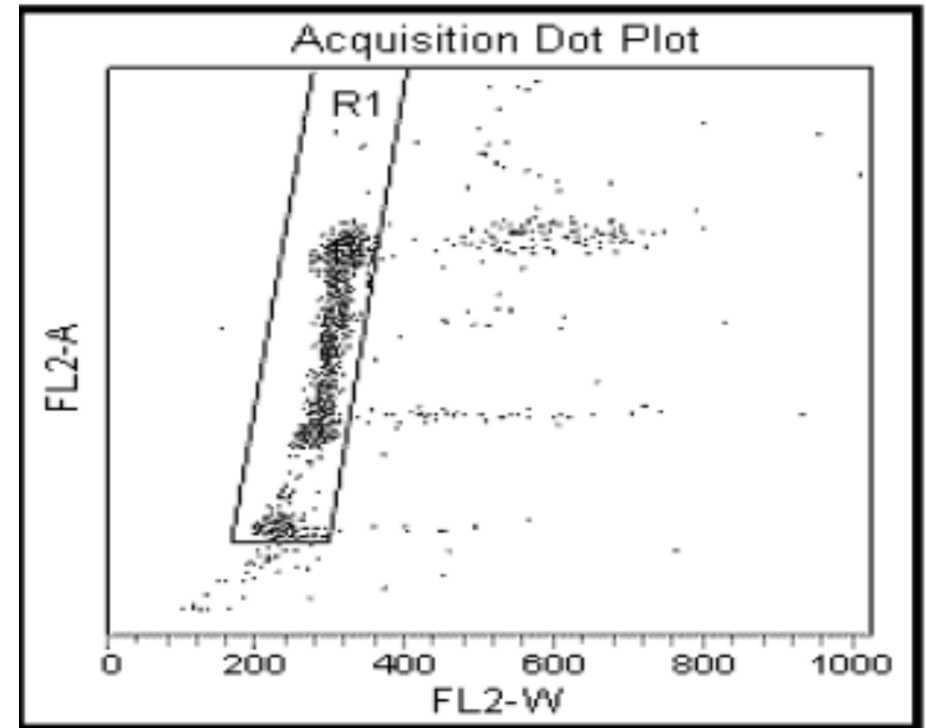
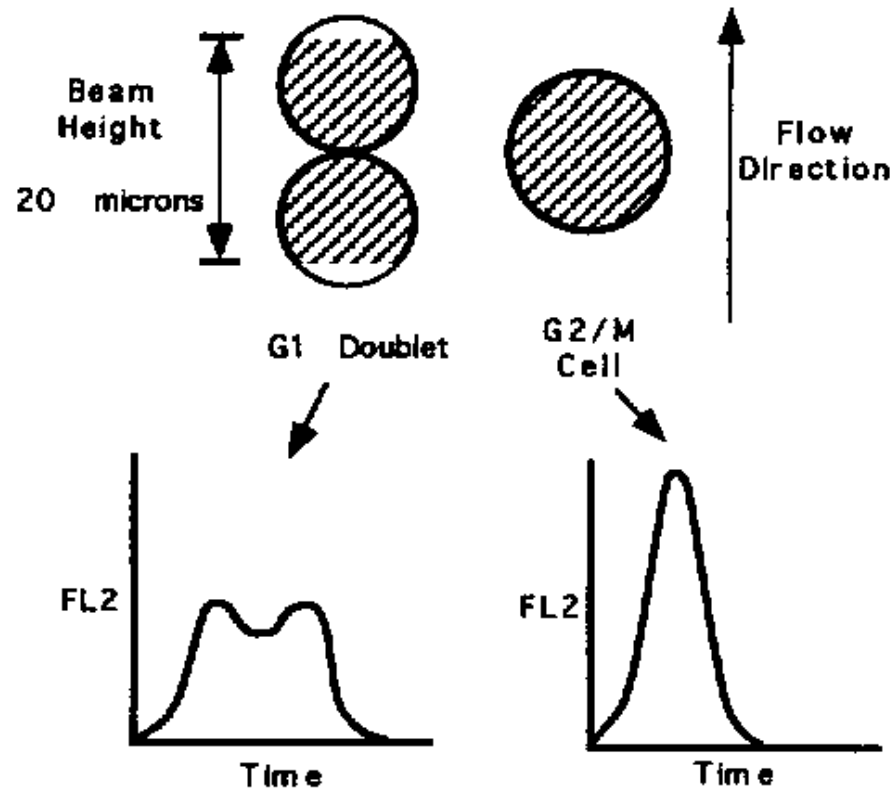
318	367	319	375	423	432	937	1985	7650	12420	15300	13256	5791	2471	842	433	331	311	308	376	349	414	823	1373	903	514	338	418	307	317	353	313	703	403	378	308	406	405	303
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Digitized values

Quantification of a Voltage Pulse



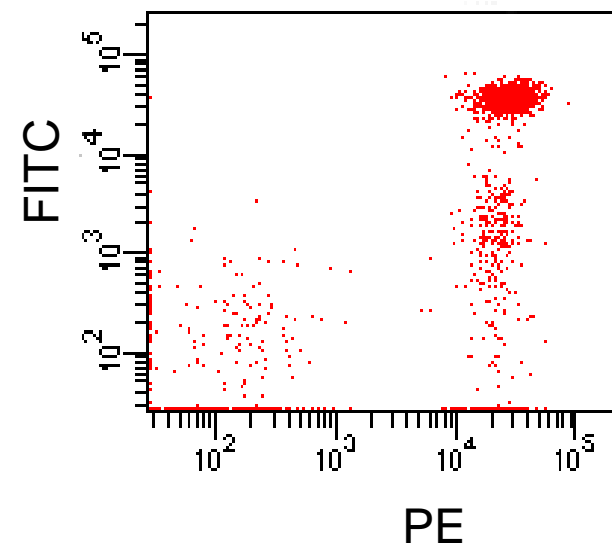
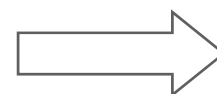
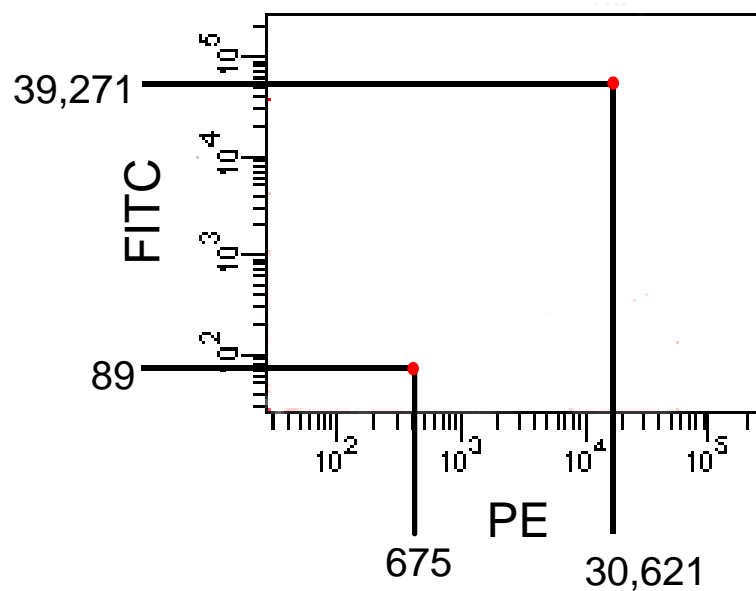
Doublet Discrimination



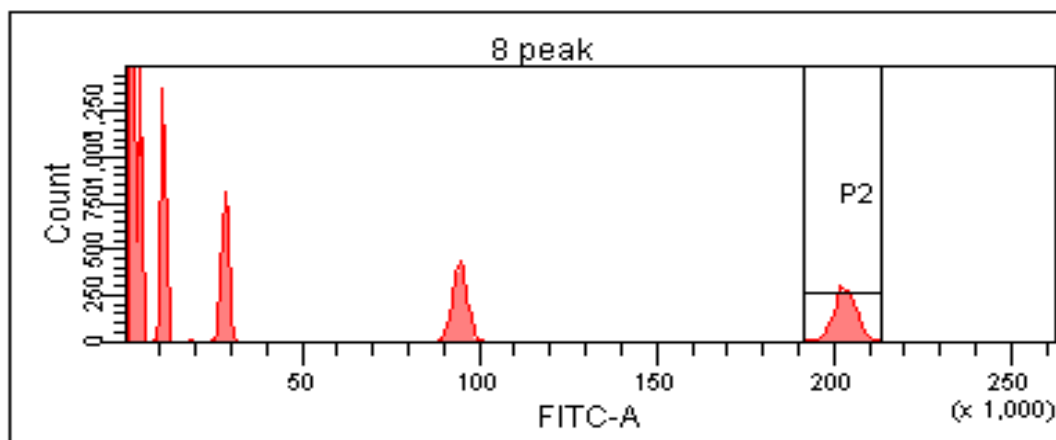
Data Storage

List-Mode Data

	Time	FSC	SSC	FITC	PE
Event 1	0	60	120	89	675
Event 2	10	160	65	39,271	30,621
Event 3	30	650	160	22,688	6,189

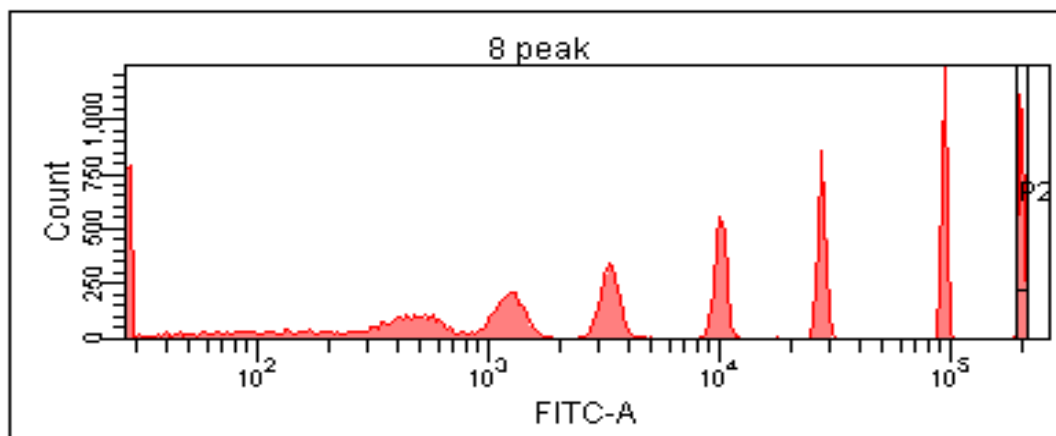


Data Display: Linear vs Log



Tube Name: 8 peak

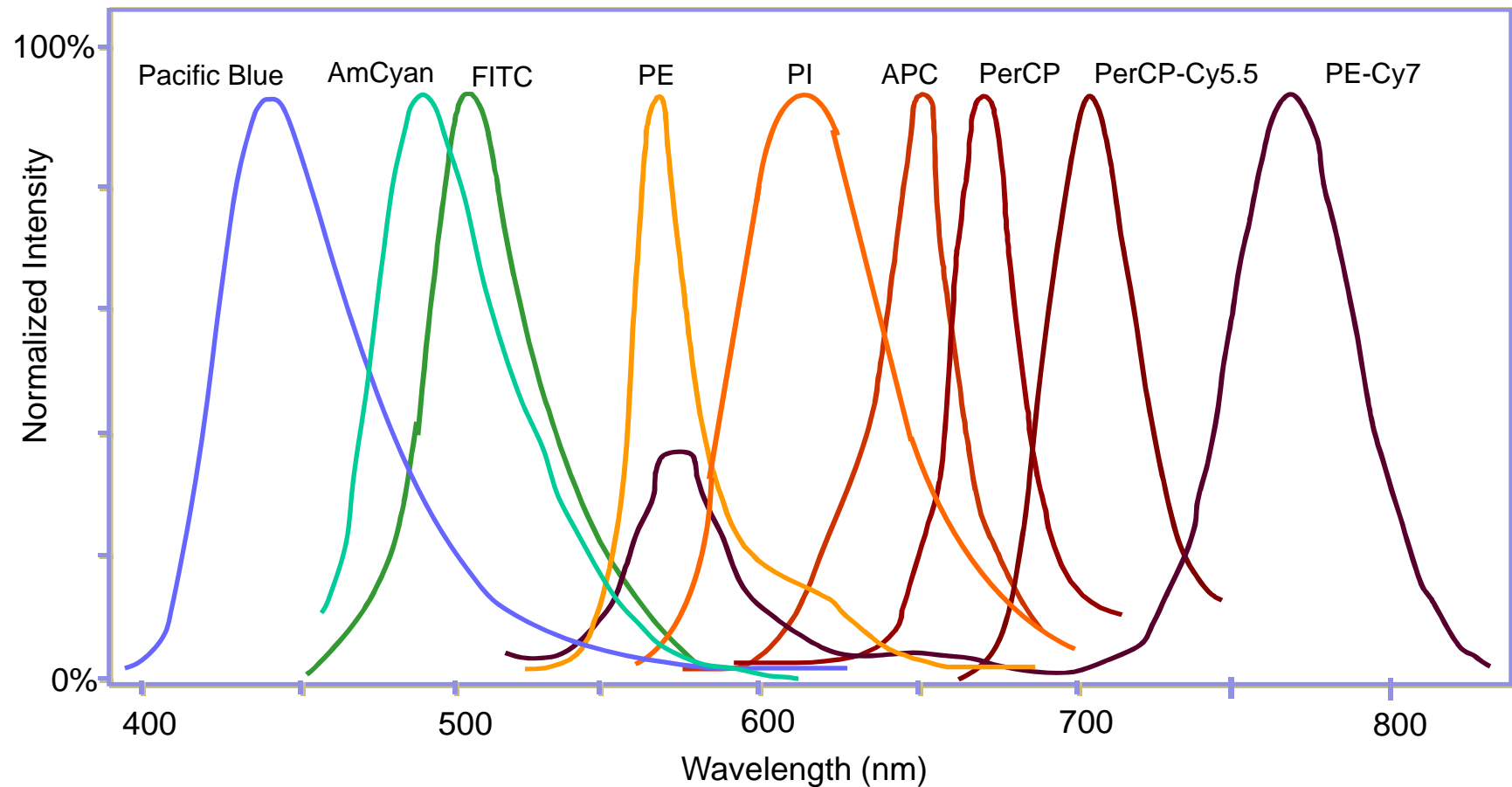
Population	#Events	FITC-A Mean
■ P1	16,589	42,948
☒ P2	2,124	203,334



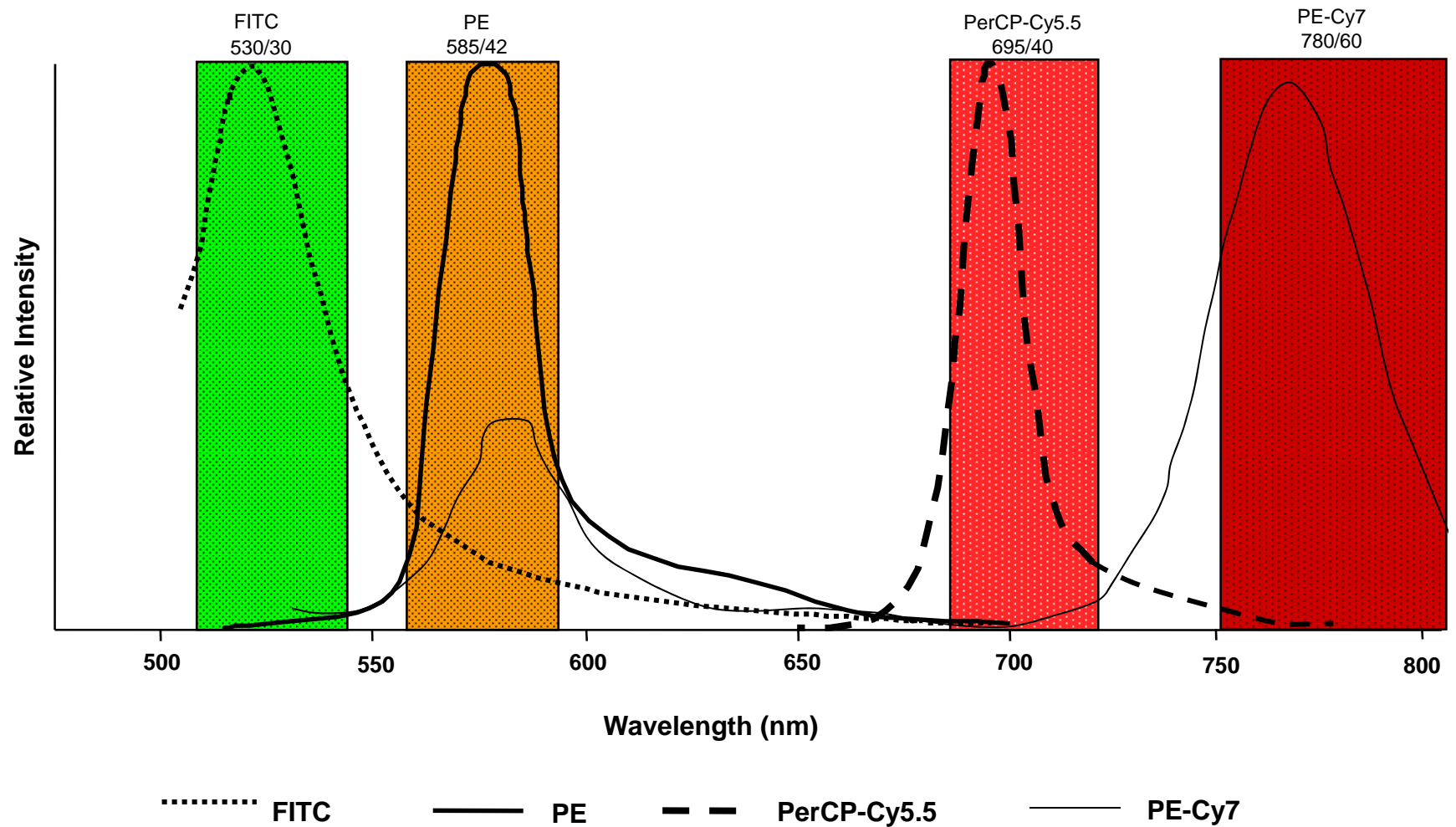
Tube Name: 8 peak

Population	#Events	FITC-A Mean
■ P1	16,589	42,948
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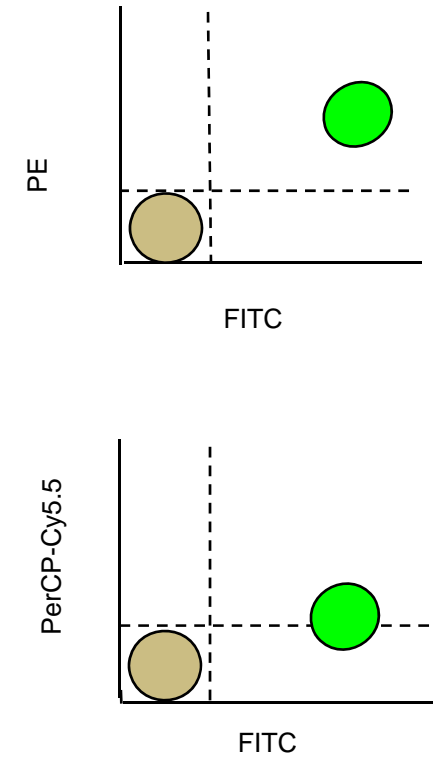
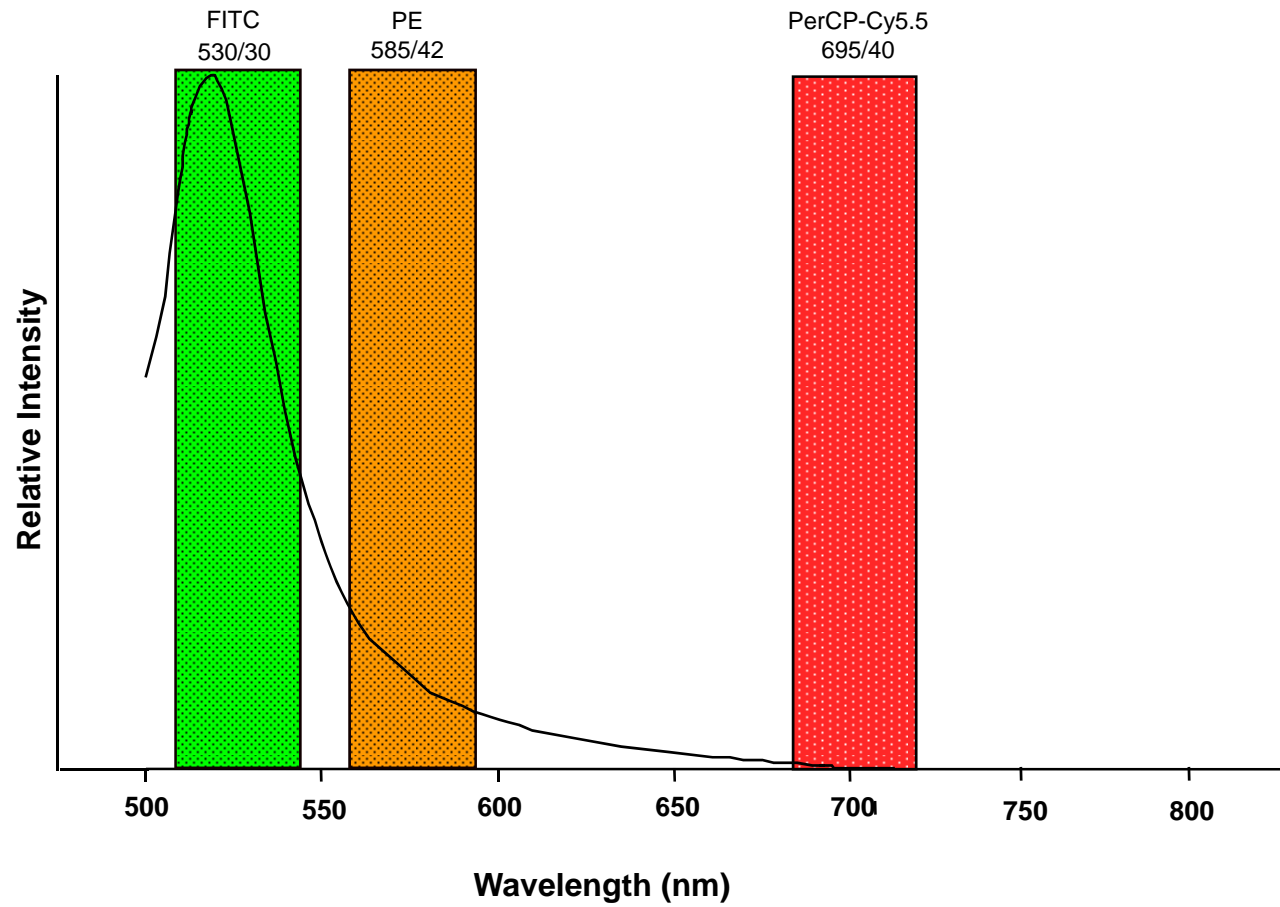
Spectral Overlap- Compensation Theory



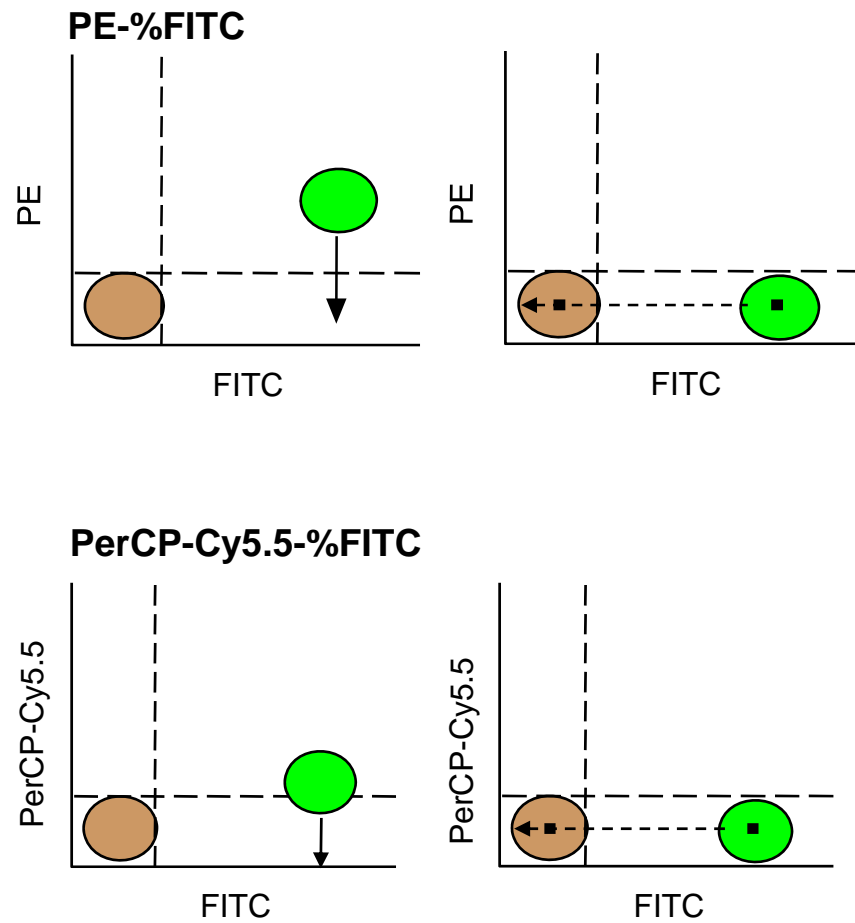
Spillover



FITC Spillover

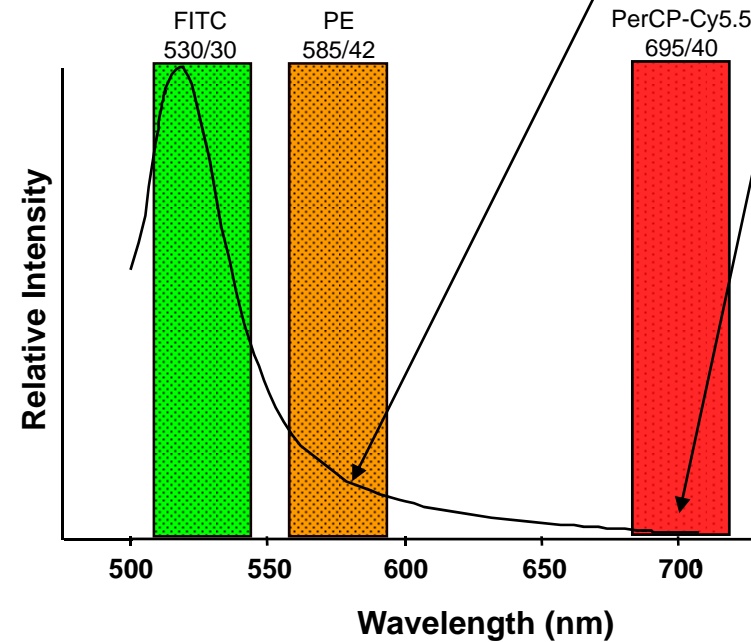


FITC Compensation



To lower cluster, increase value.

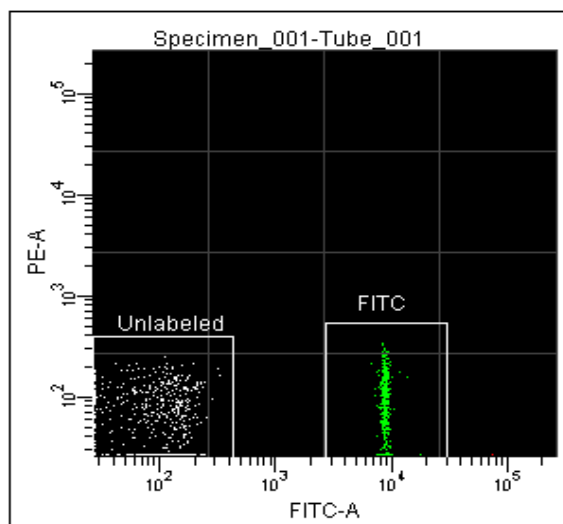
	Fluorochrome	- % Fluorochrome	Spectral Overlap
•	PE	FITC	20.10
•	PerCP-Cy5-5	FITC	0.90



Compensation Examples

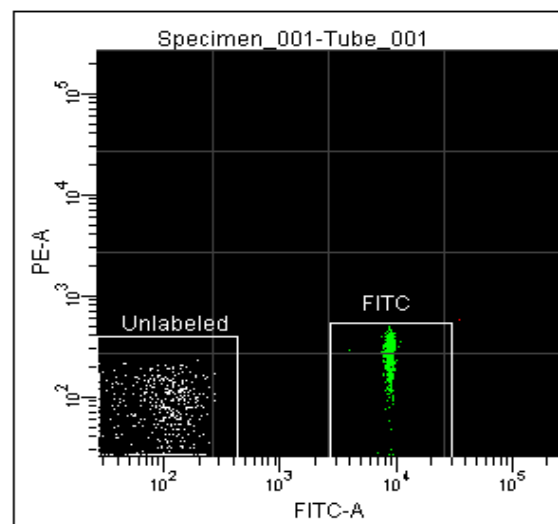
Incorrect Compensation

Correct Compensation



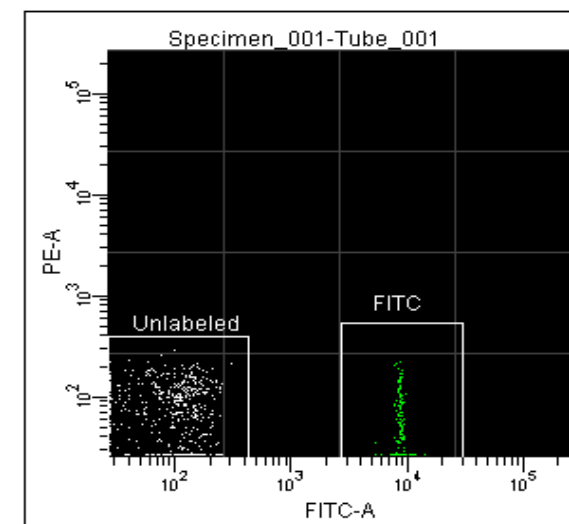
Population	PE-A Mean
Unlabeled	77
FITC	73

Undercompensation



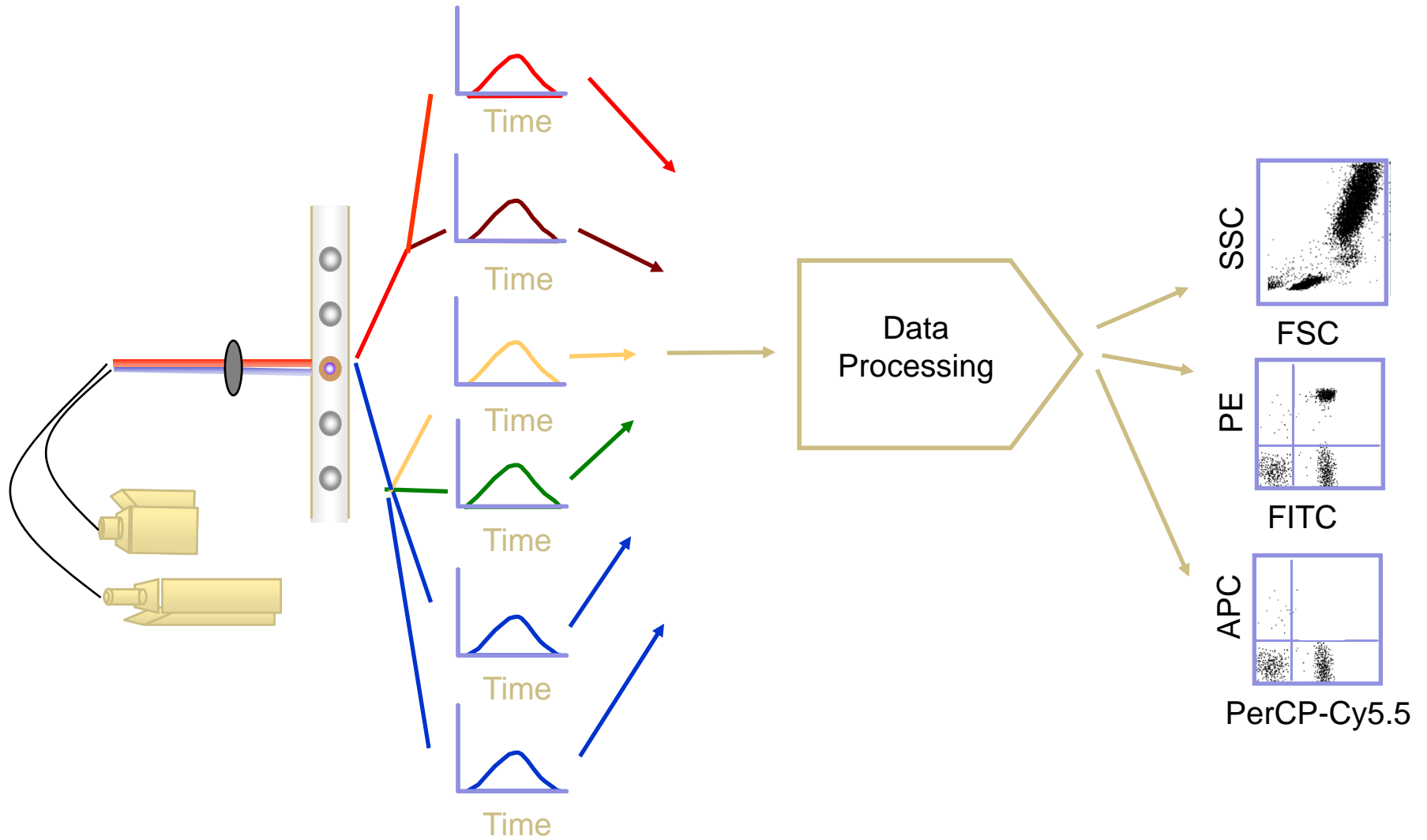
Population	PE-A Mean
Unlabeled	72
FITC	245

Overcompensation



Population	PE-A Mean
Unlabeled	70
FITC	-54

Review



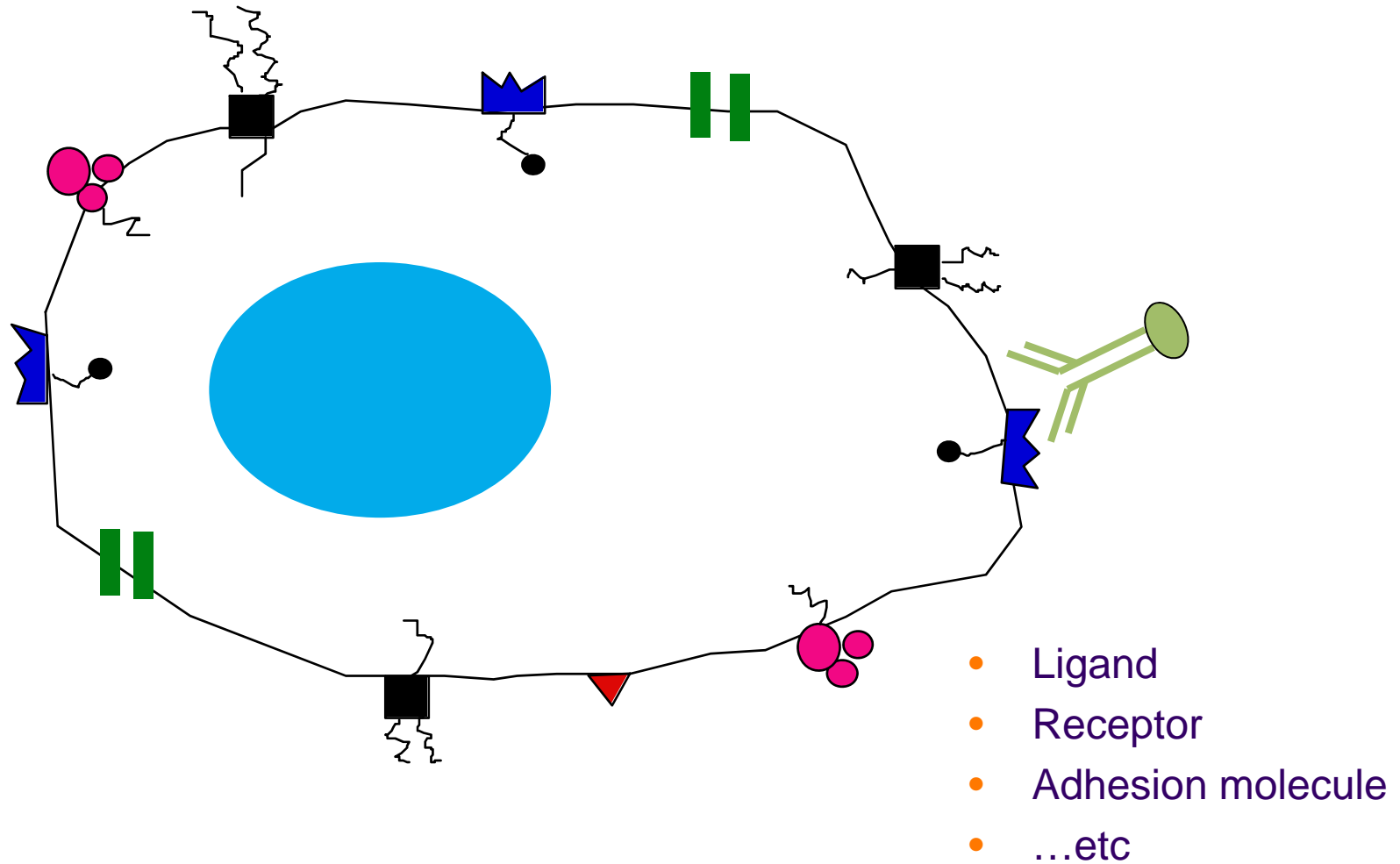


Application Examples

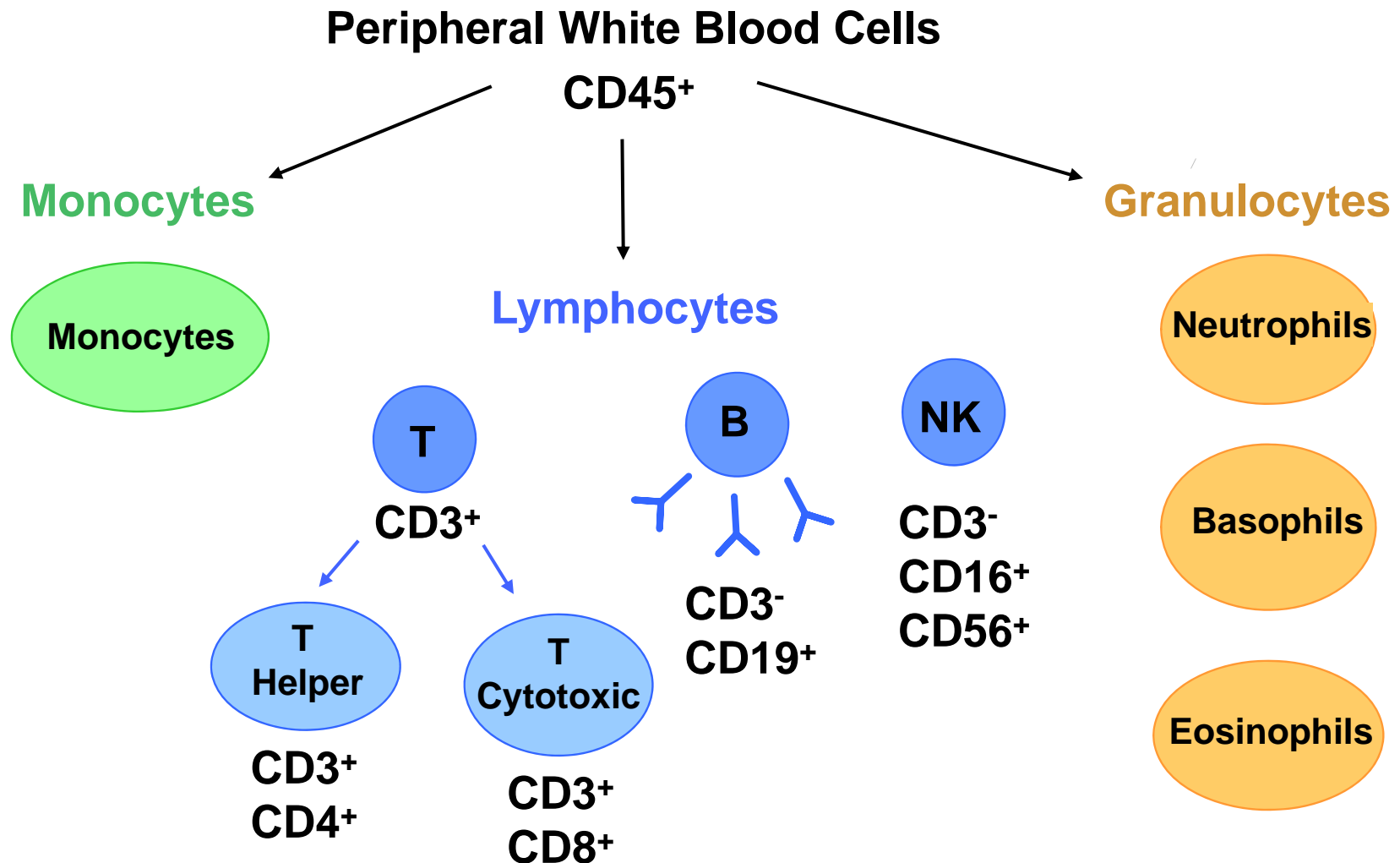
Applications

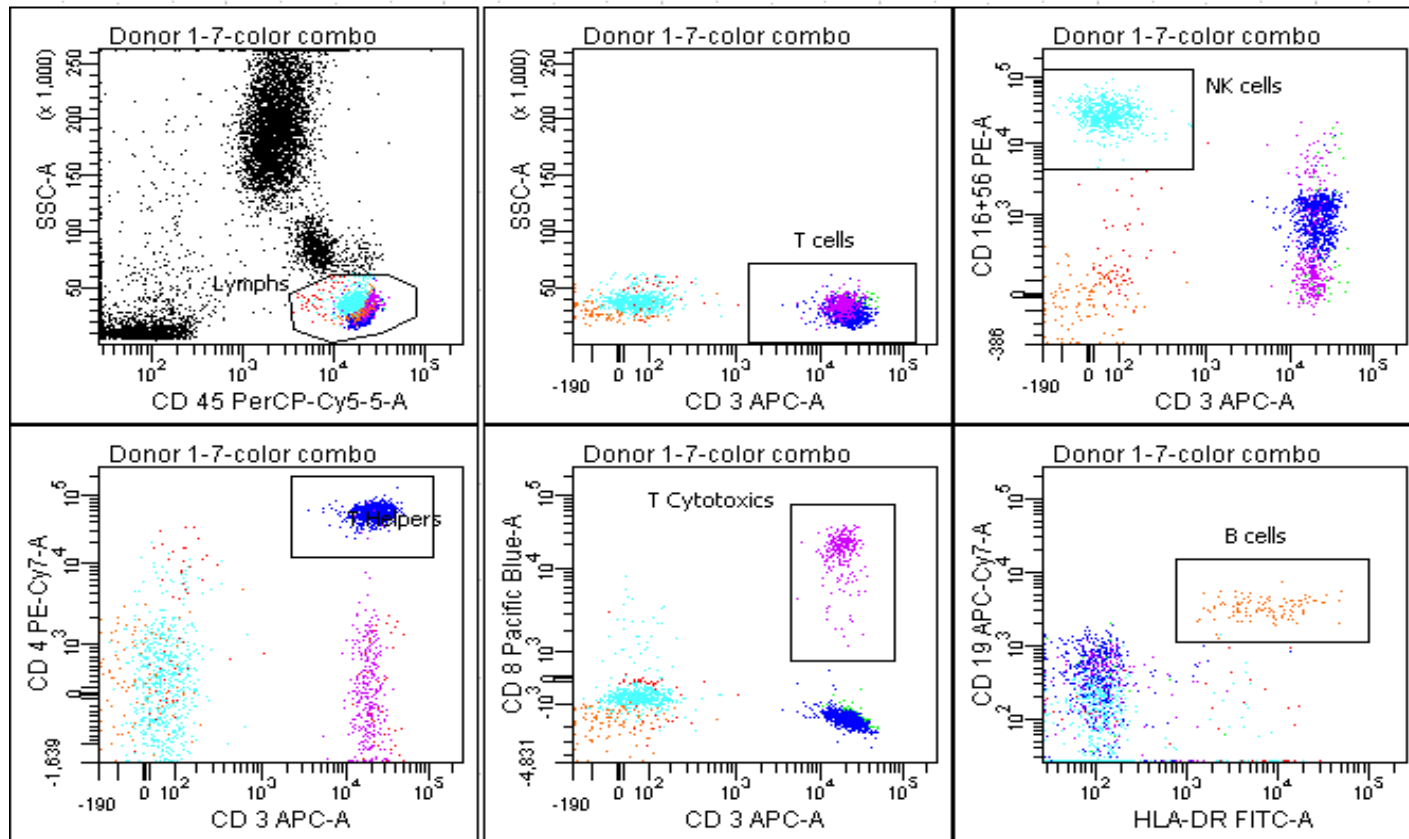
- Phenotype Analysis (Cell Surface Antigens/Markers)
- Intracellular Analysis
 - Eg. Cytokines, Signal Transduction molecules...etc.
- DNA Analysis
 - Eg. Viability, Cell cycle, Apoptosis...etc.
- Cell Function Analysis
 - Eg. Free radicals, Ca²⁺, Reporter genes...etc.
- CBA (Cytometric Bead Array)
- Others

Phenotype Analysis



Lymphocyte Immunophenotyping





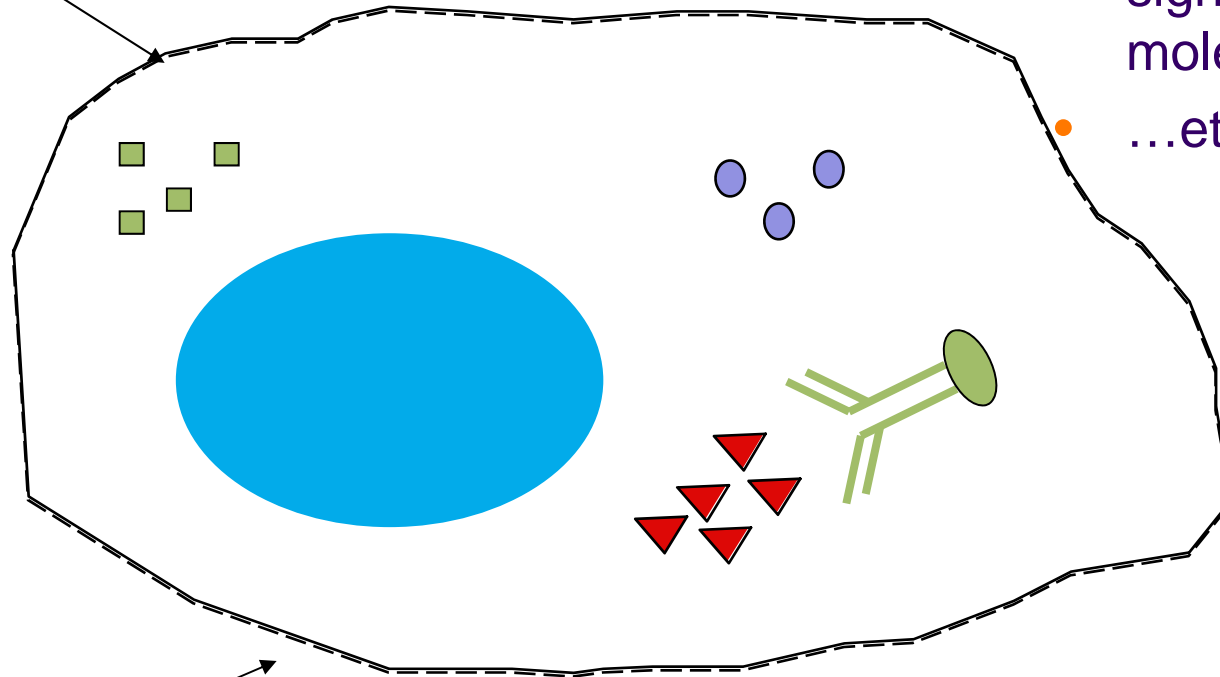
Tube: 7-color combo

Population	#Events	%Parent	%Total
All Events	10,000	####	100.0
Lymphs	1,948	19.5	19.5
T cells	1,195	61.3	12.0
T Helpers	860	72.0	8.6
T Cytotoxics	300	25.1	3.0
B cells	111	5.7	1.1
NK cells	586	30.1	5.9

Intracellular Analysis

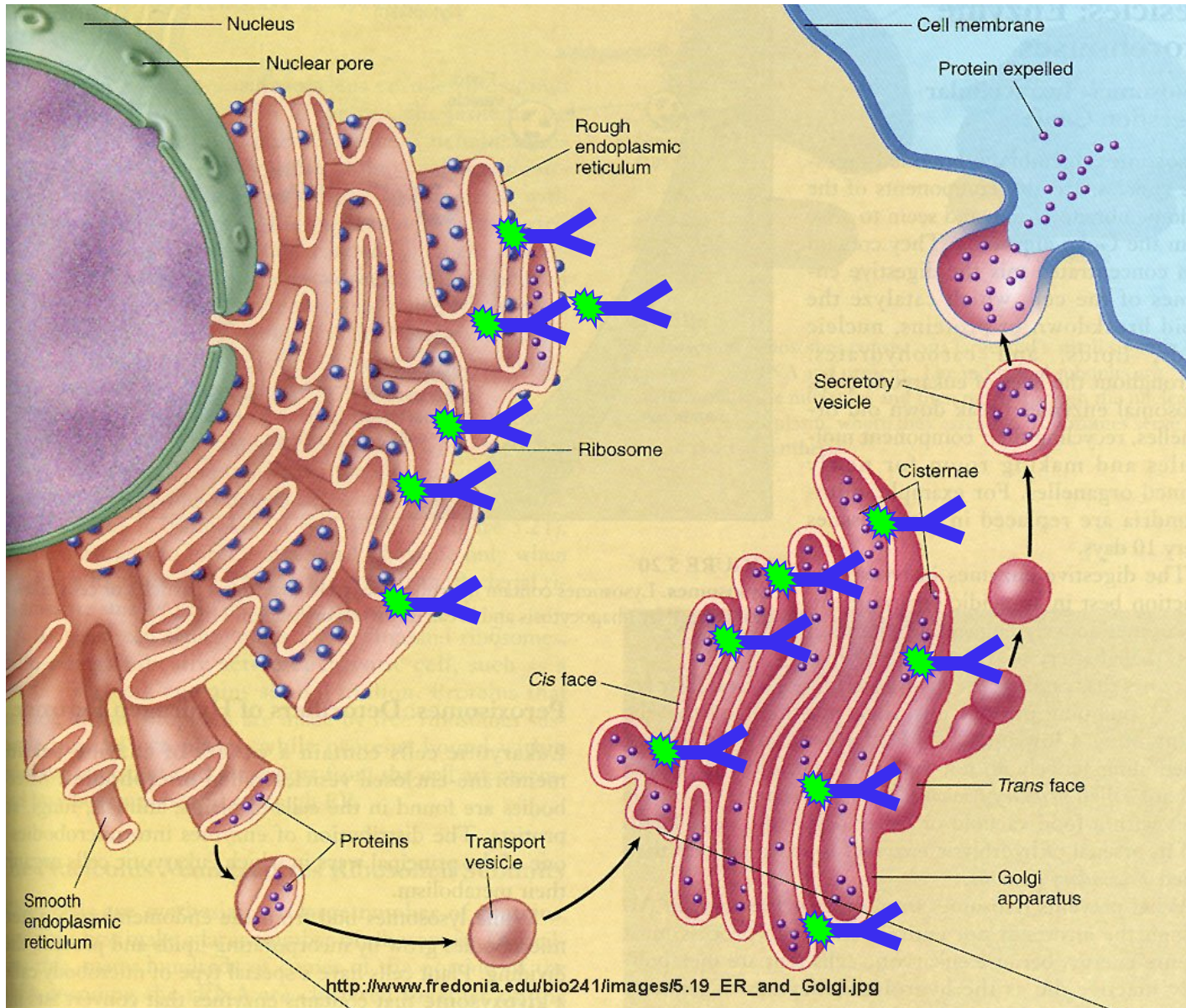
Permeabilizing
solution

Fixation
solution

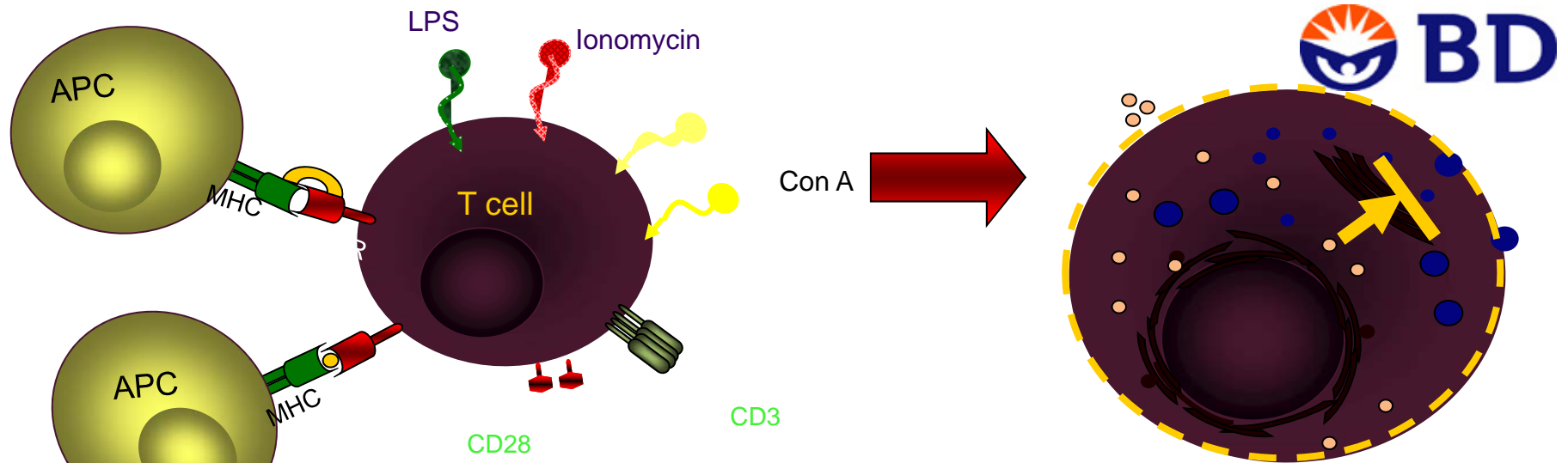


- Cytokine
- Enzyme
- signal transduction molecule
- ...etc.

Cytokine Detection



Picture From www.fredonia.edu



Stimulation

To enhance the accumulation of intracellular cytokines.

Monensin: Cytokines accumulate in the ER
 Brefeldin A: in Golgi complex.

Secretion stop

(Brefeldin A or Monensin)
 Only *in vitro*

To maintain structural integrity.
 Formaldehyde or glutaraldehyde
 Keep the protein structure and doesn't change the
 (accessibility of the) epitopes too much

Intracellular Staining

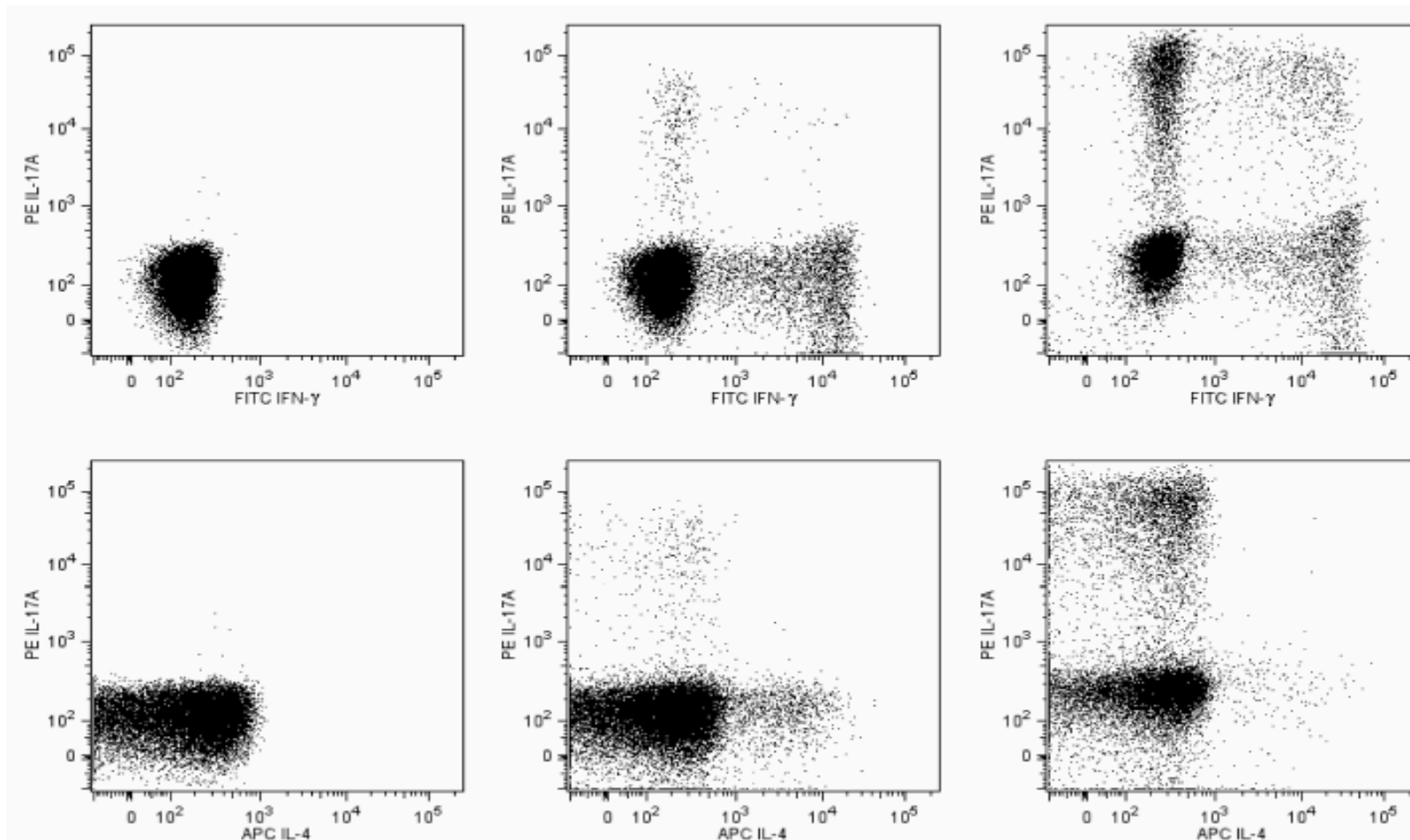
Permeabilisation

Fixation

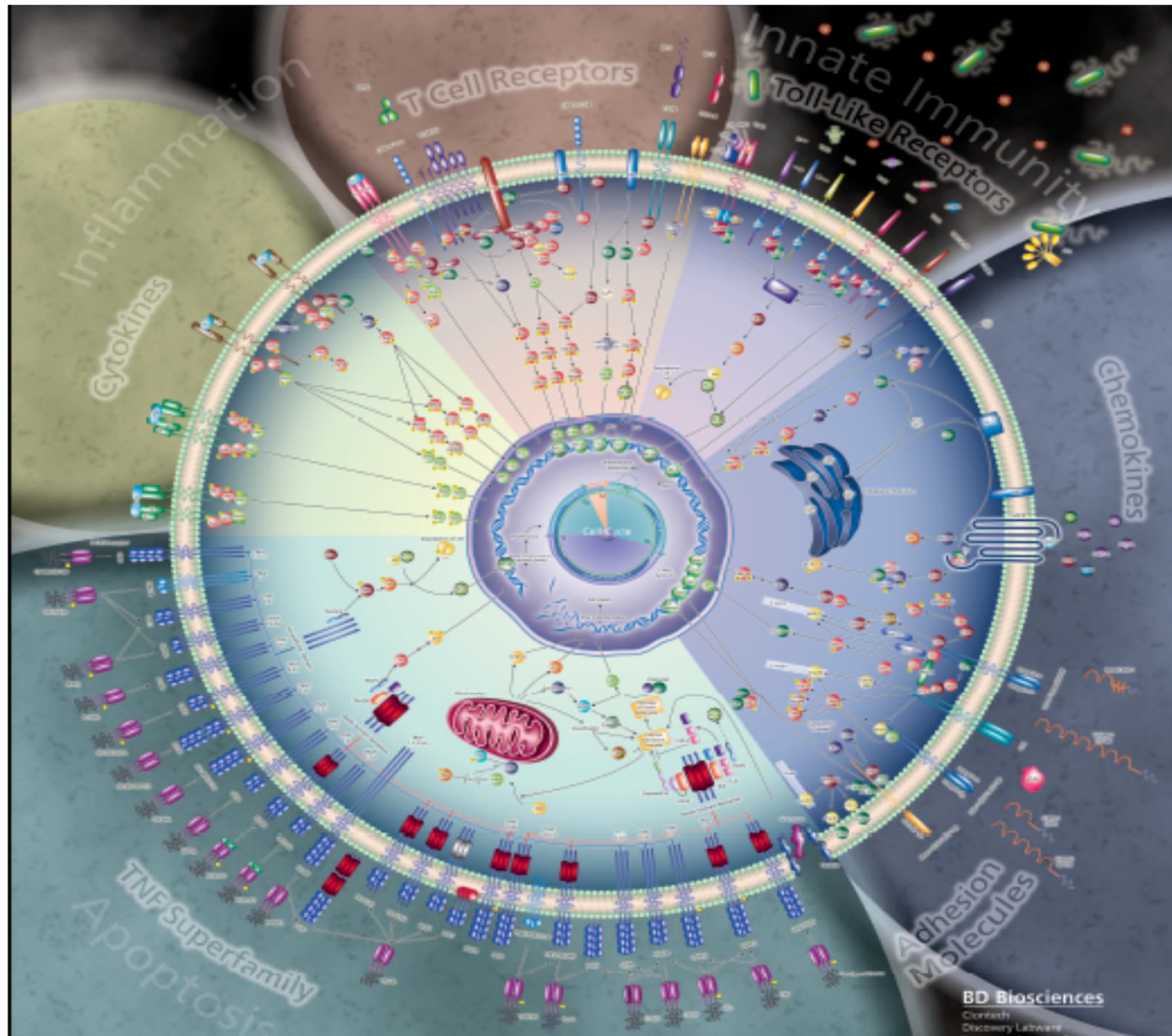
Saponin (permeabilisation buffer).

Combination of Cell Surface and Cytoplasmic Staining

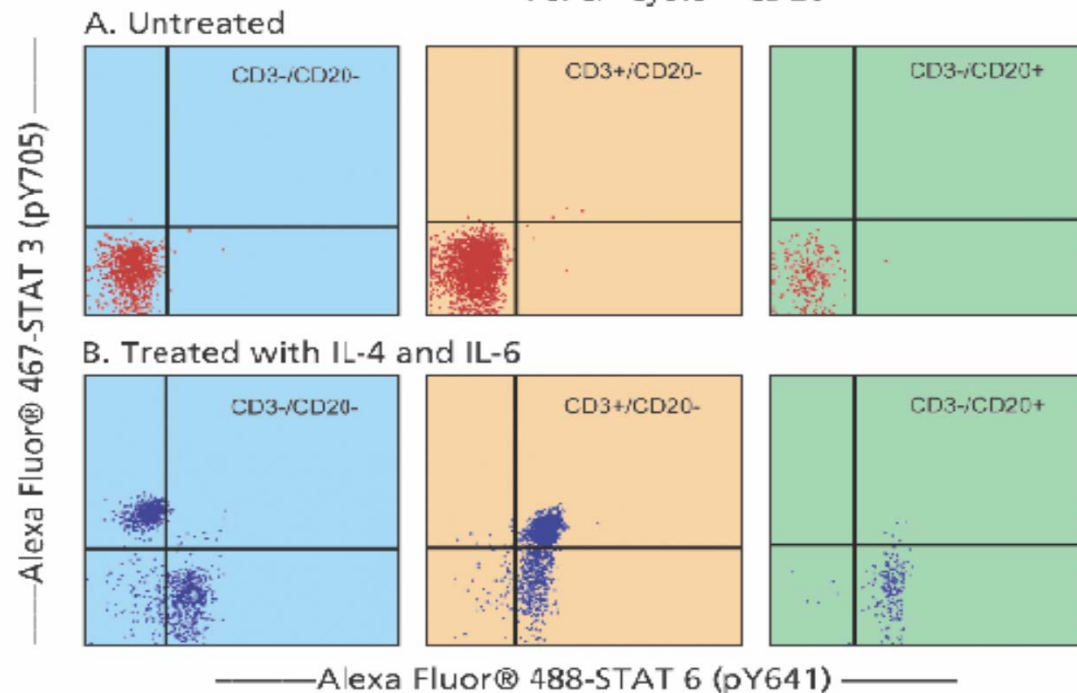
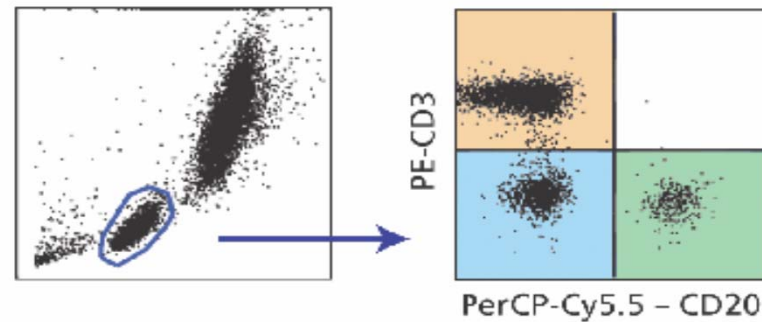
Th1/Th2/Th17 Phenotyping Kit



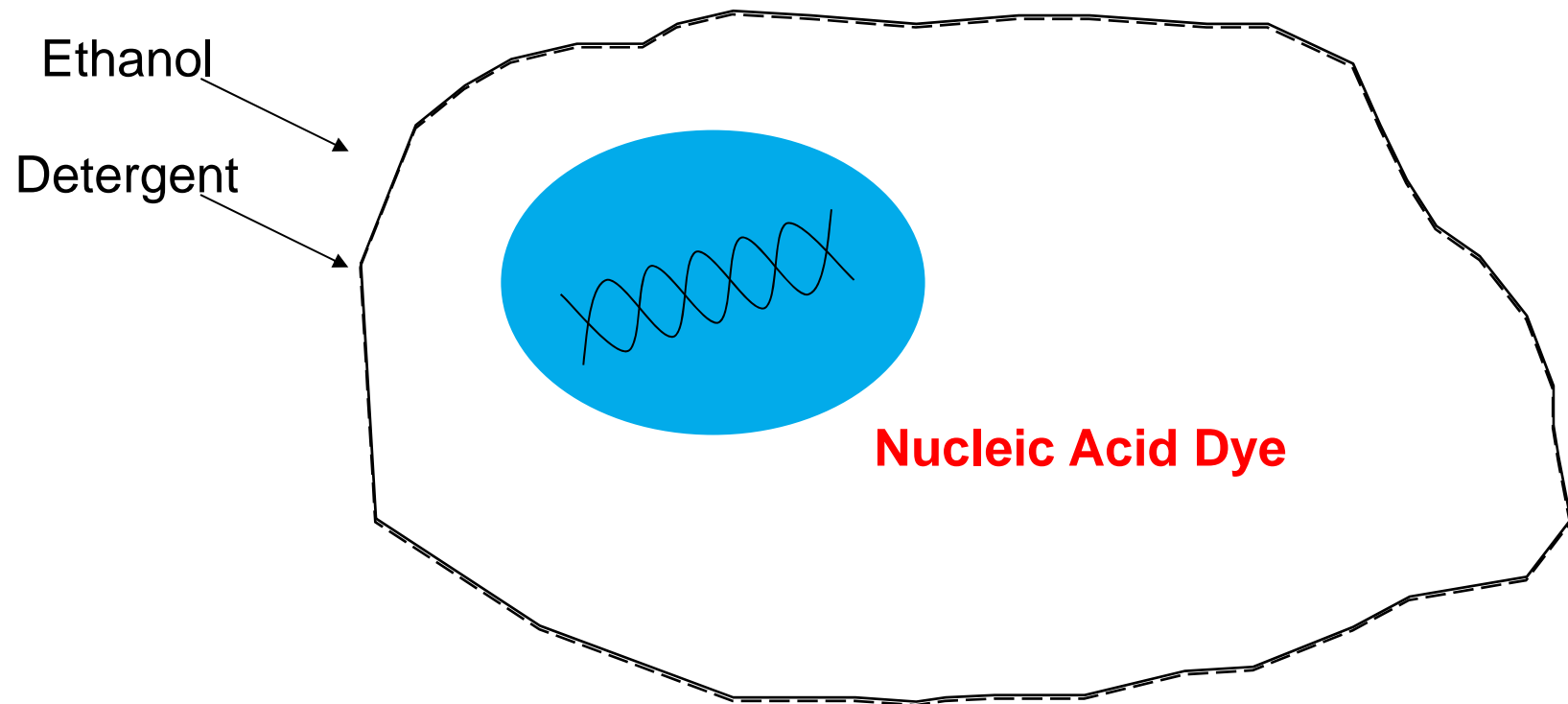
Signal Transduction



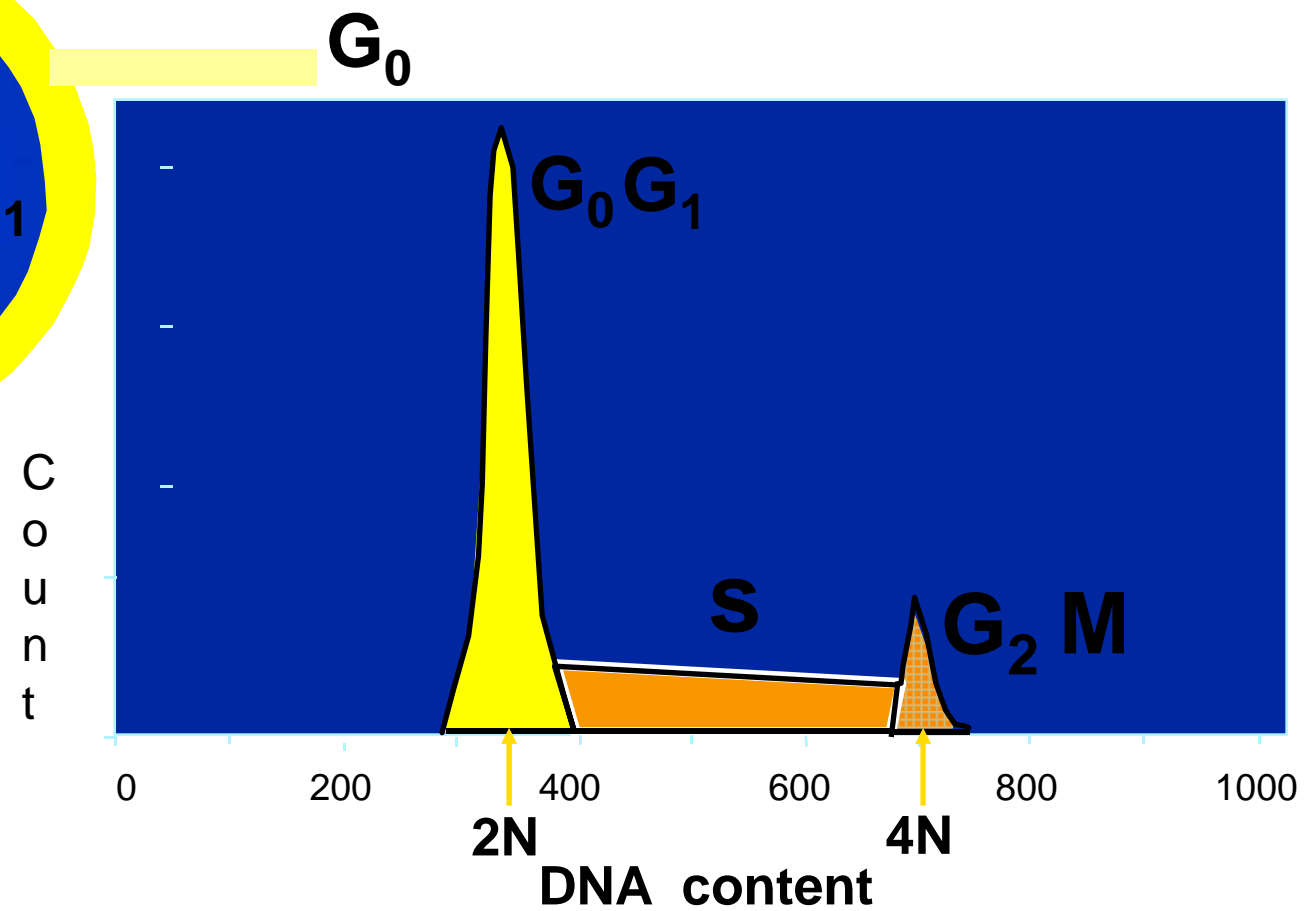
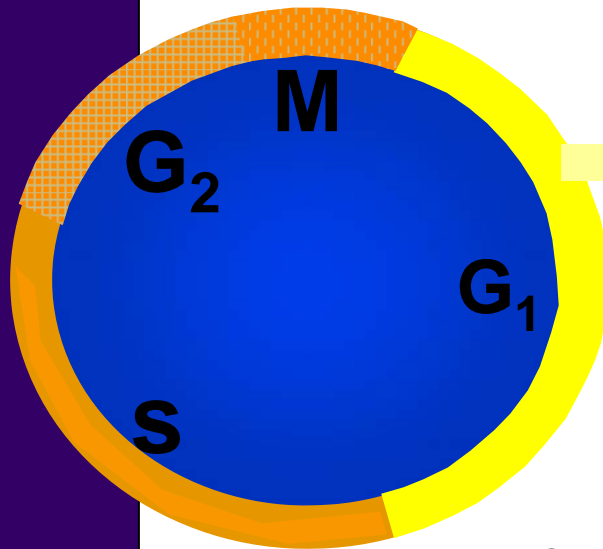
Intracellular Staining in Activated Lysed Whole Blood



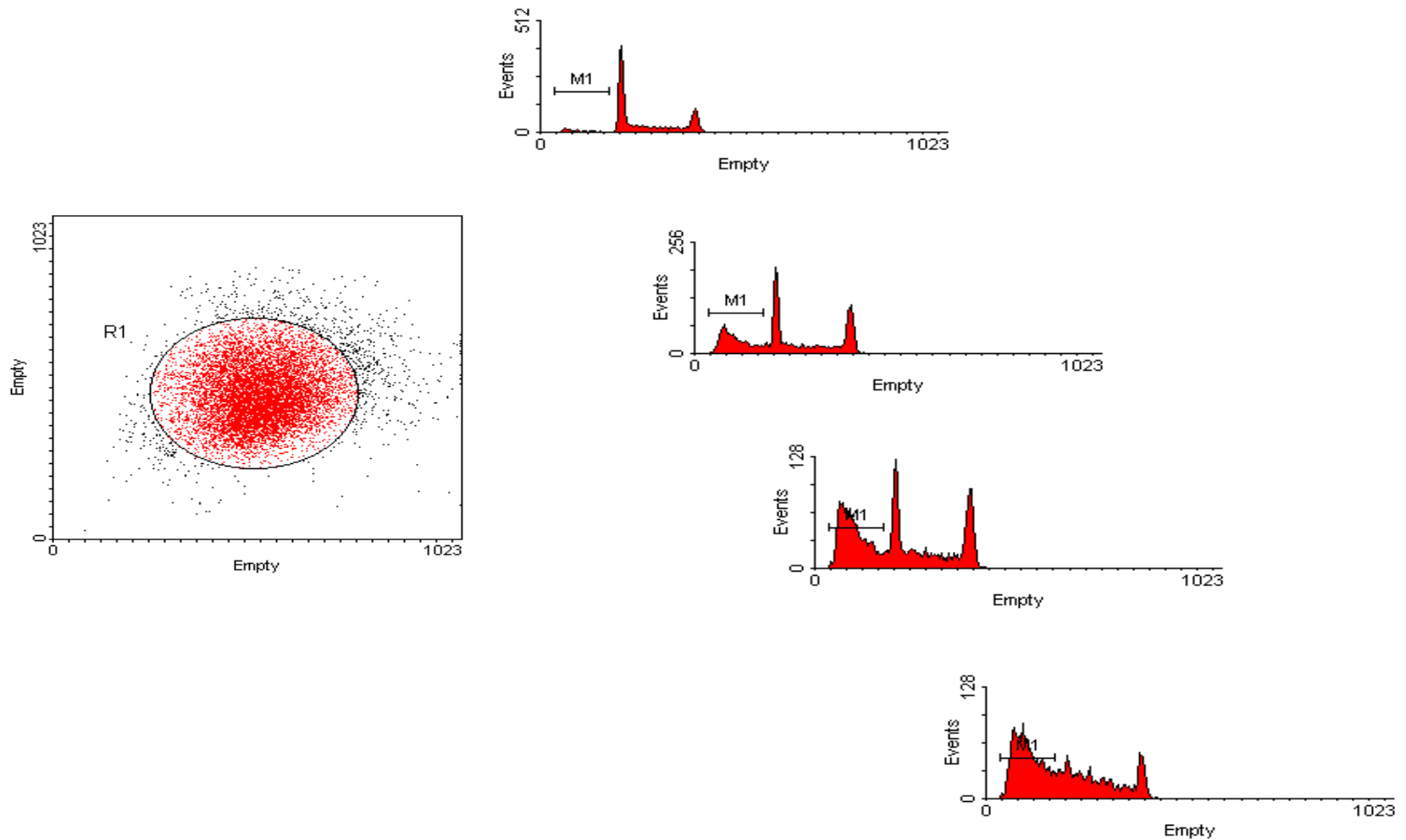
DNA Analysis



Cell Cycle Analysis



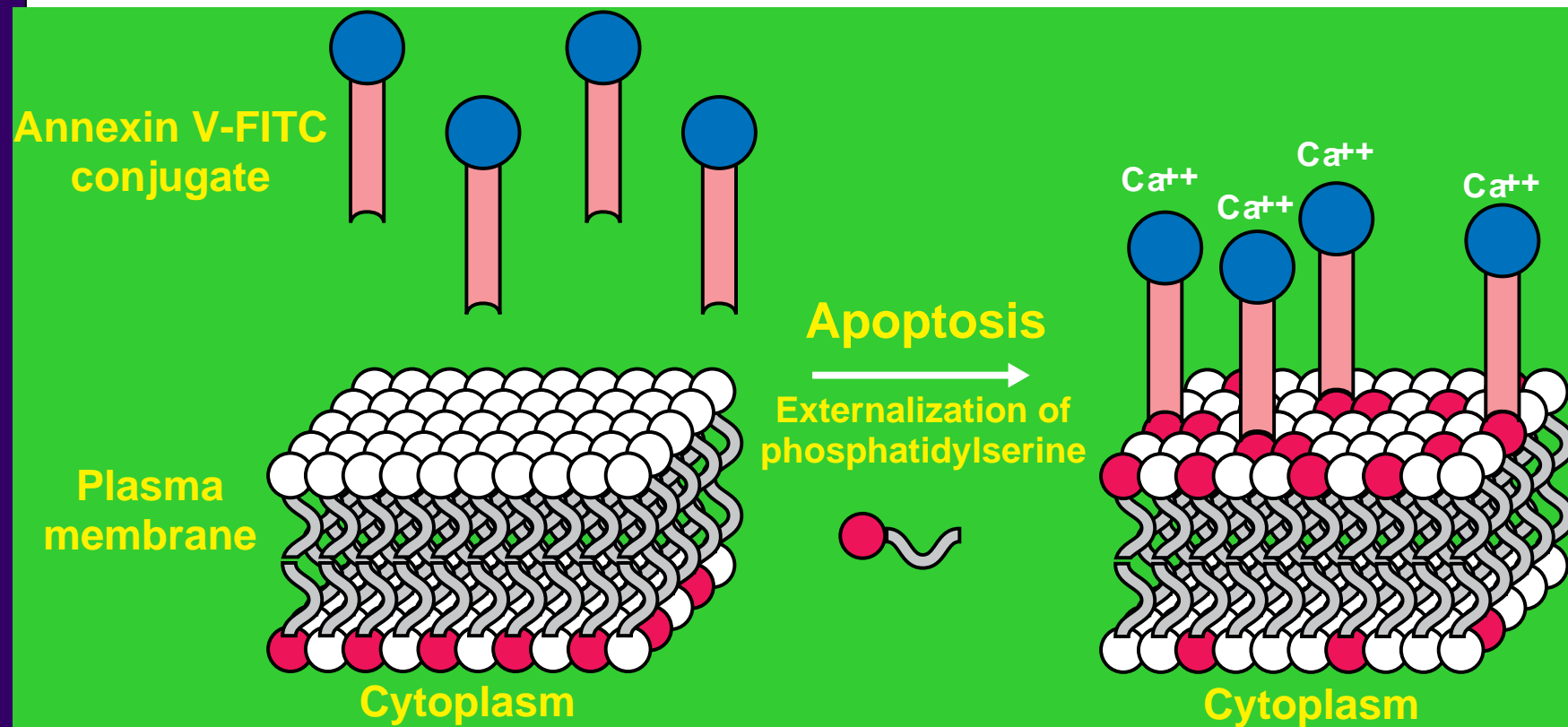
Apoptosis (Sub G1)



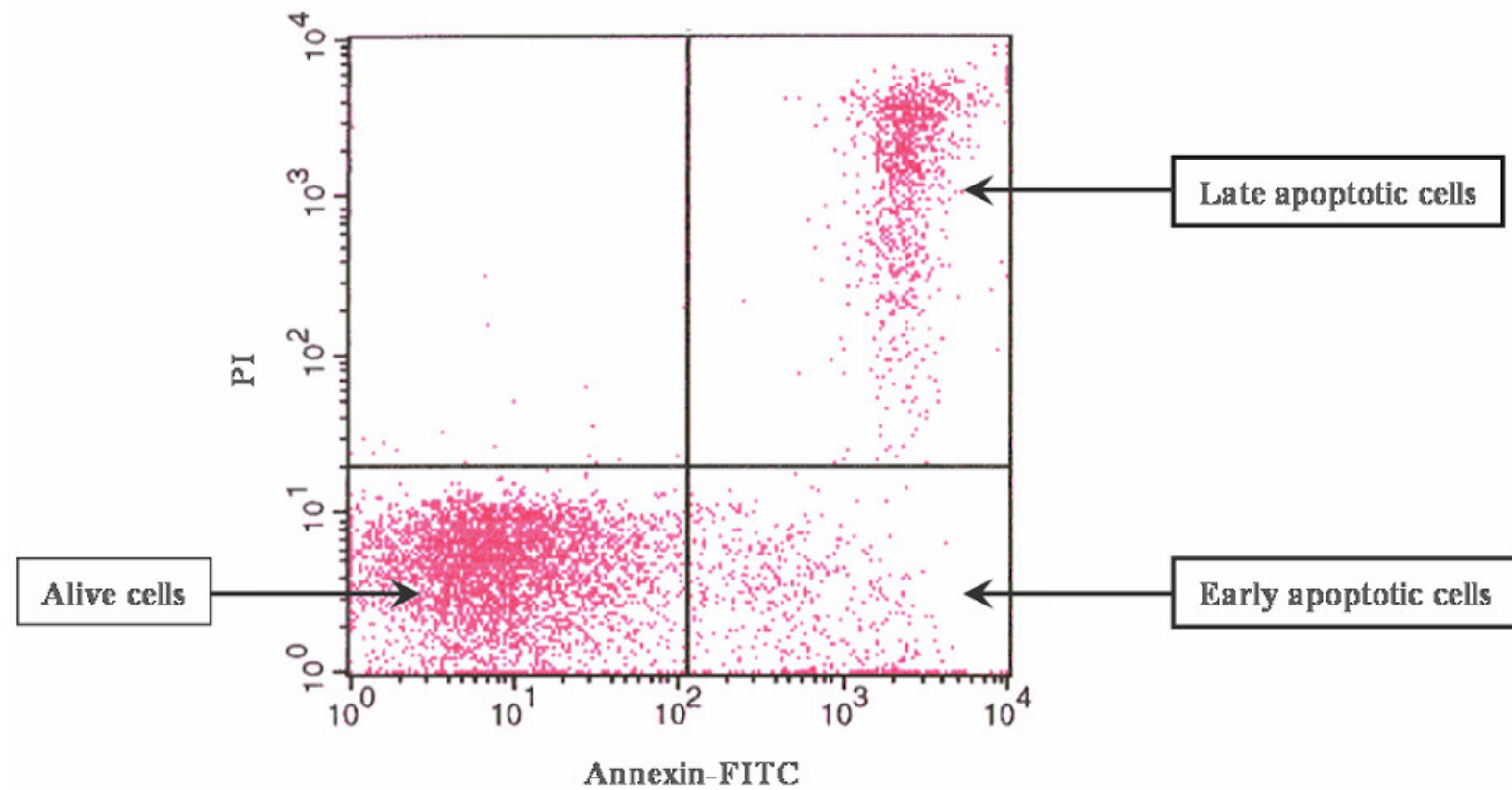
Cell Function Analysis

- Membrane Potential (DiOC6, JC-1)
- Oxidative Metabolism (Free Radicals)
- Intracellular PH Value (Snarf-1)
- Ca⁺⁺ Influx (Fluo-4/Fura Red, Indo-1)
- Phagocytosis
- Cell Proliferation (PI, BrdU, Intracellular Cyclins)
- Apoptosis (Annexin V, active Caspase-3)

Annexin V Assay

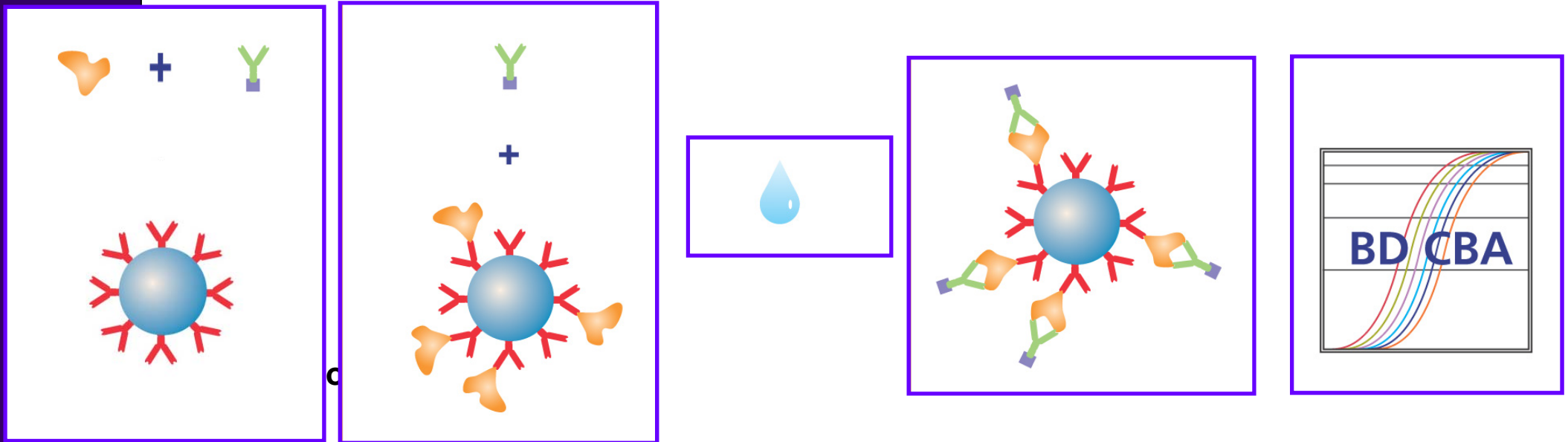


Annexin V/PI Double Staining

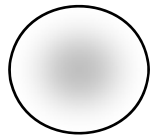
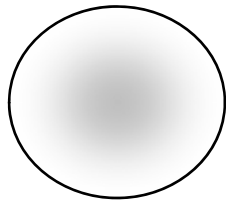


Bordón et al. Radiation Oncology 2009 4:58

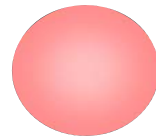
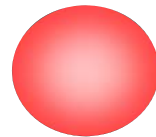
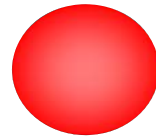
Cytometric Beads Array (CBA)



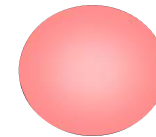
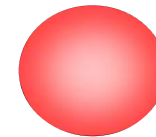
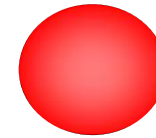
Beads Provide a Flexible Platform



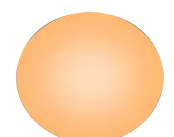
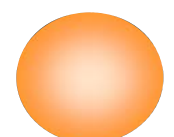
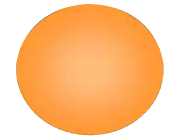
**Multiple
sizes**



**Different
fluorescence
intensities**

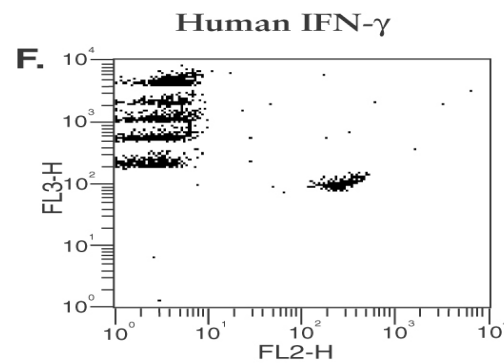
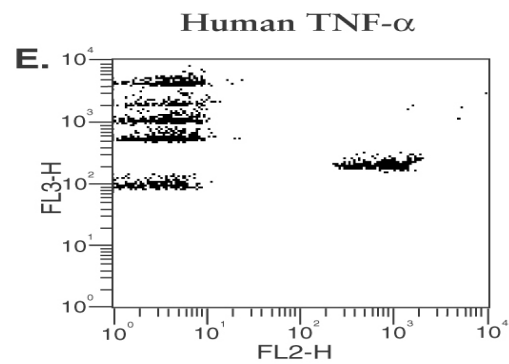
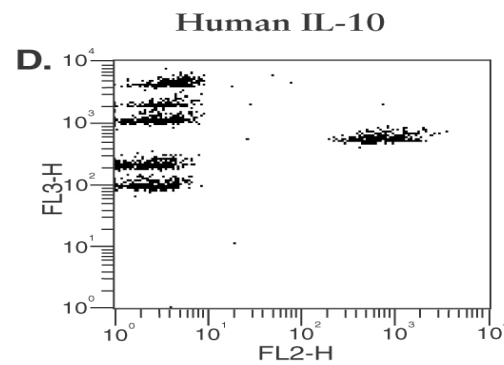
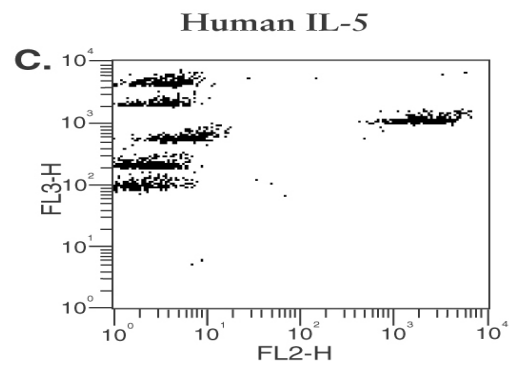
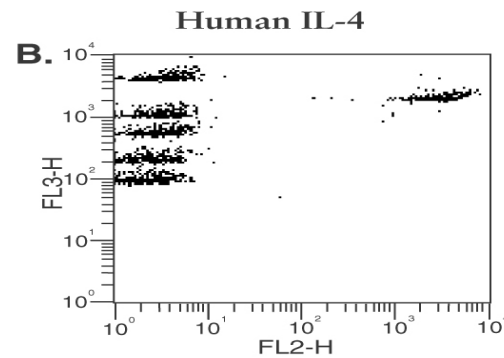
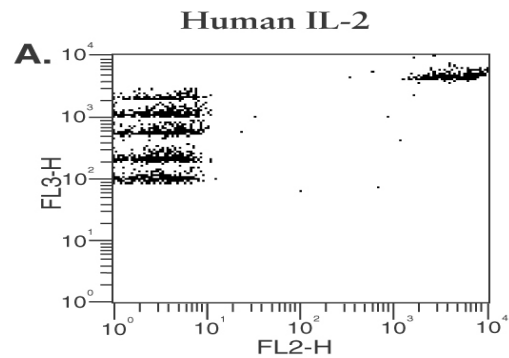


**Different colors with
different intensities**



Advantages of Bead-Based Immunoassays

- Analyze multiple analytes simultaneously
- Reduced sample volume requirements
- Reduced hands-on time by parallel analysis of samples
- Wide dynamic range of fluorescence detection (requires fewer sample dilutions)



Proteins Measured

A. Interleukin (IL)-2

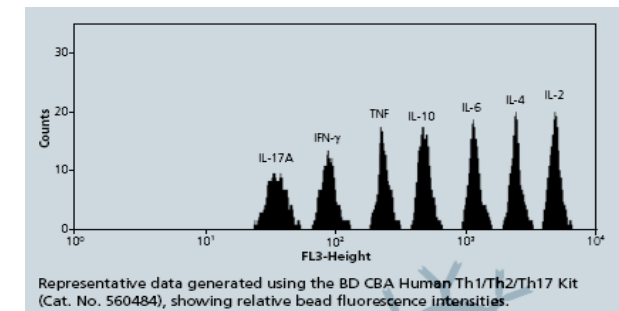
B. IL-4

C. IL-5

D. IL-10

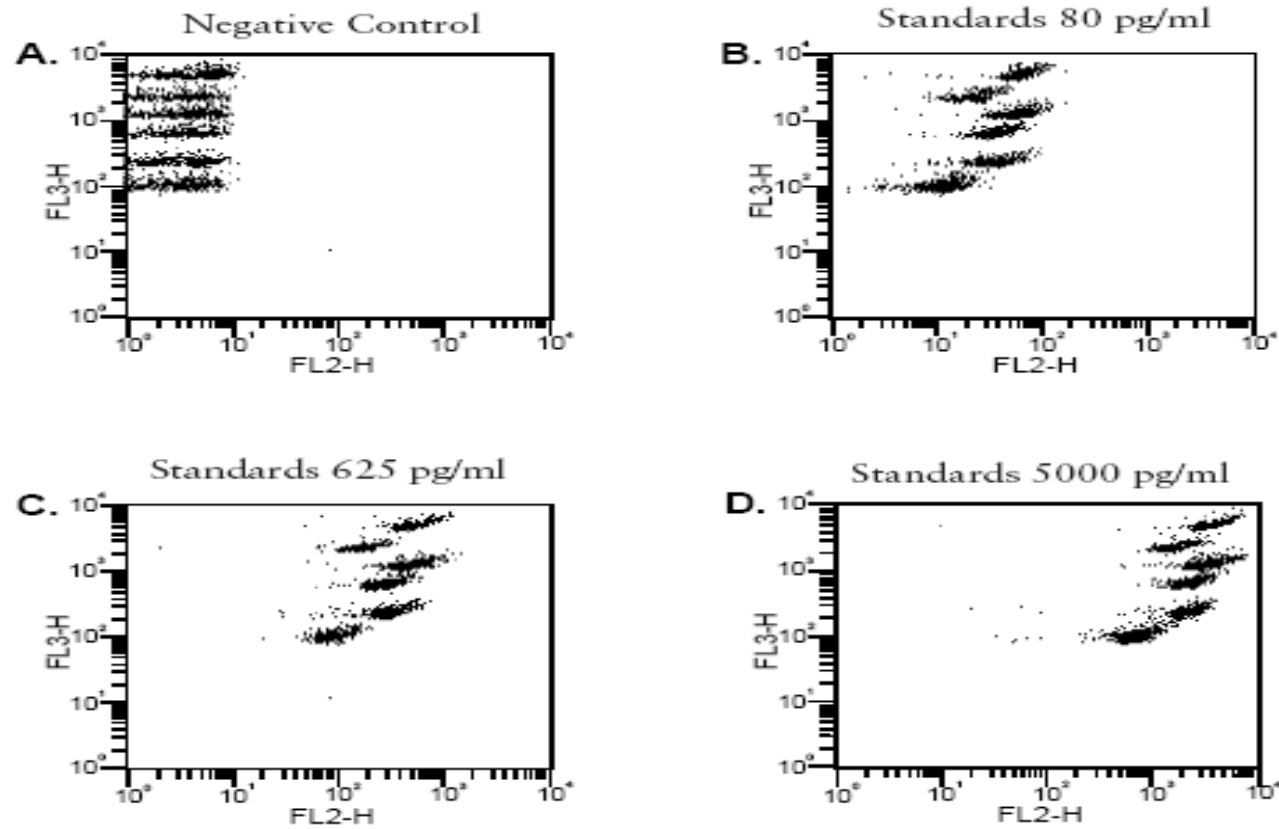
E. Tumor Necrosis Factor- α

F. Interferon- γ

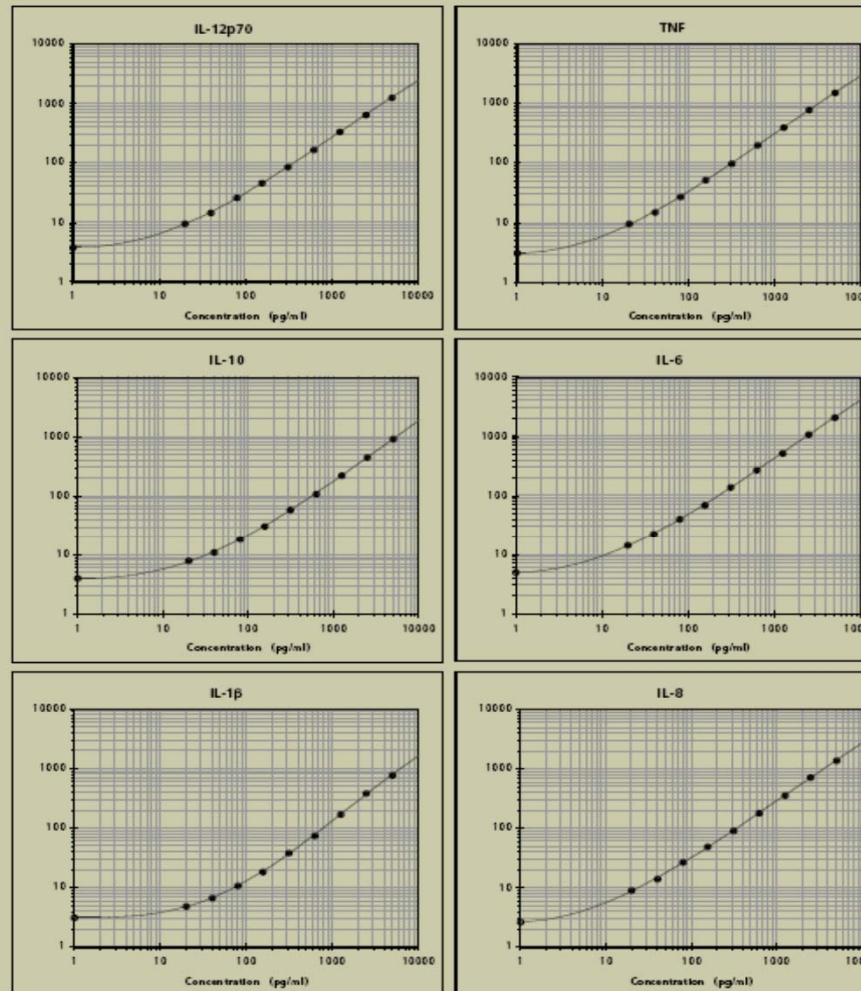
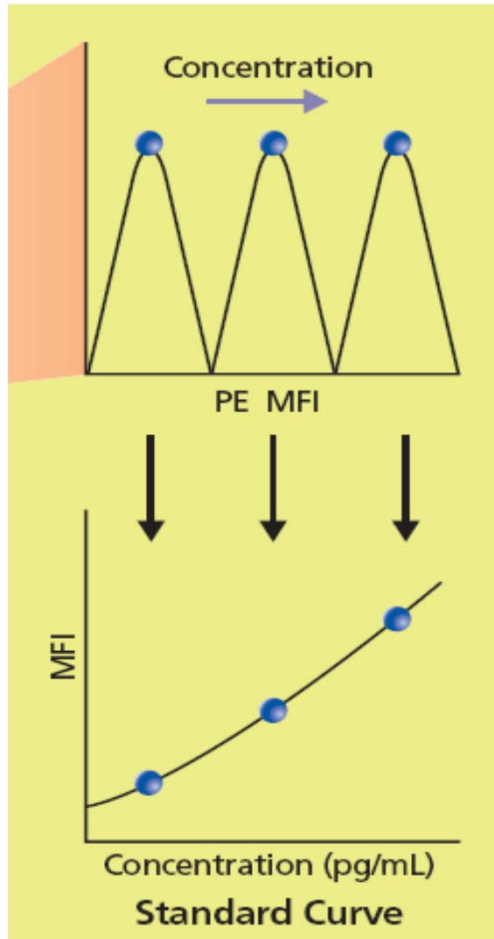


Cytometry Beads Array (CBA)

Typical Data



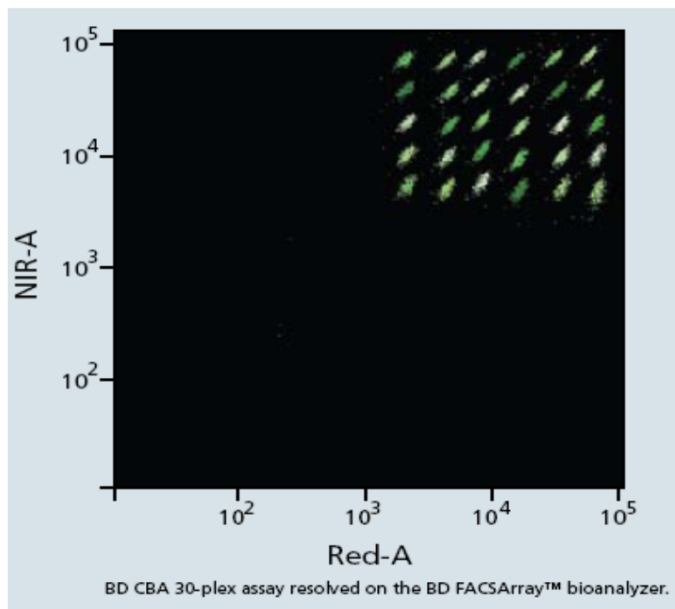
Standard Curves



Representative standard curves generated using the BD CBA Human Inflammatory Cytokines Kit.

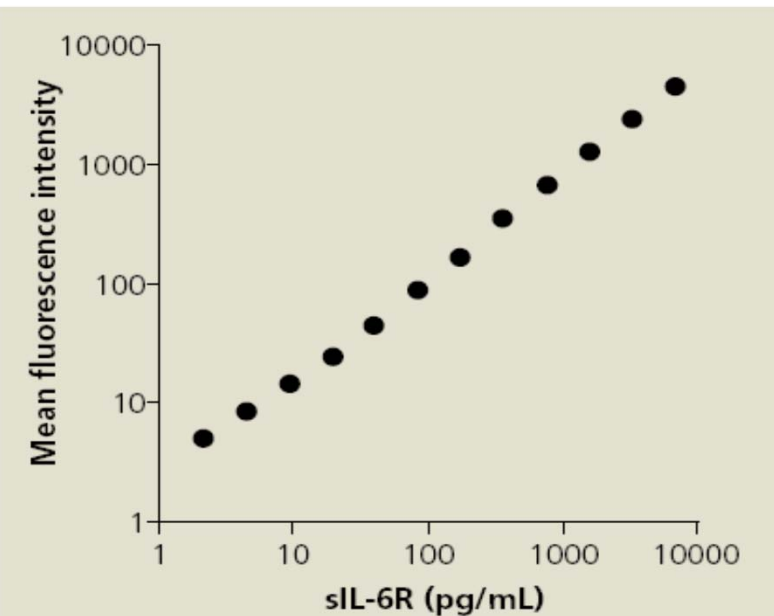
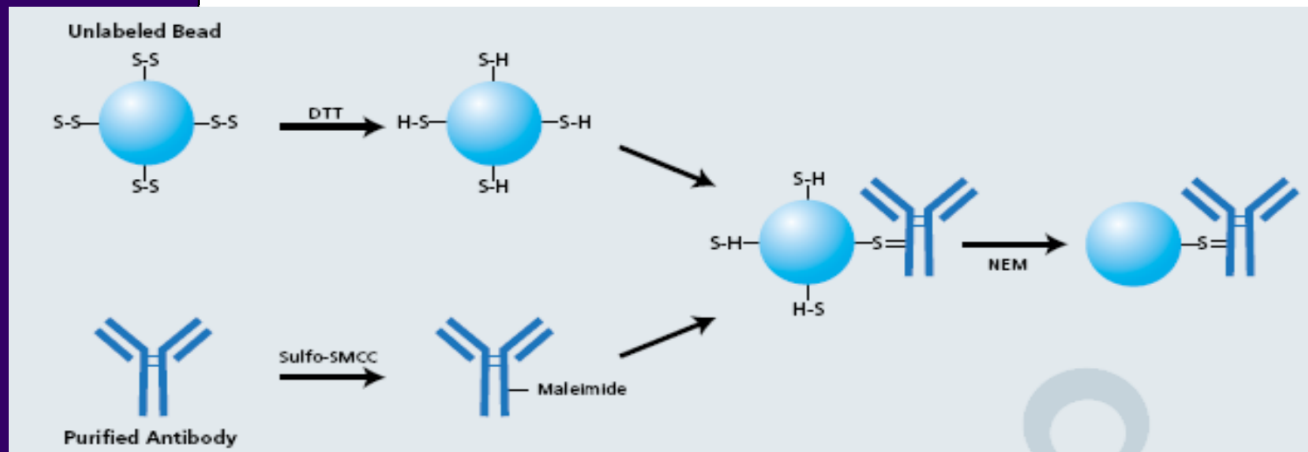
CBA Flex Sets

- Open configuration (Up to 30 plex)
- Clustering based on Red and NIR fluorescence intensity
- Need to be used at dual-laser(488nm blue v.s 633nm red) instrument



CBA Functional Beads

- Can be conjugated with any Ab



Standard curve for a soluble IL-6 receptor assay generated using BD CBA Functional Bead E4 following the conjugation procedure in the BD CBA Functional Bead Conjugation Buffer Set manual.

Data courtesy of Joseph Cannon and Gloria Sloan, Medical College of Georgia.