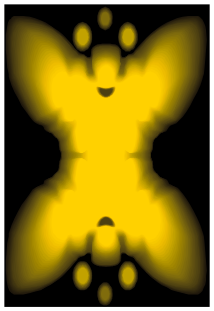
Confocal Microscopy & Time-lapse Video Recording IVIS Spectrum

錢宗良 (x88193)
臺灣大學醫學院
解剖學暨細胞生物學研究所

Why confocal microscopy ?

- **Optical sectioning:**
  Specimen is monitored slice by slice (3D-resolution)
  Each slice produces a sharp image by confocal optics



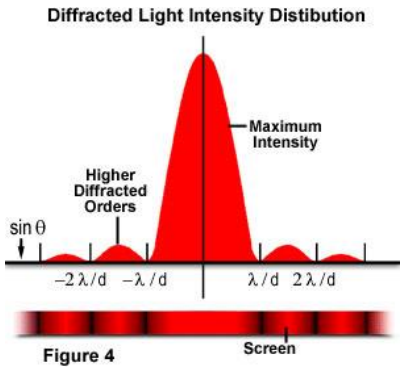
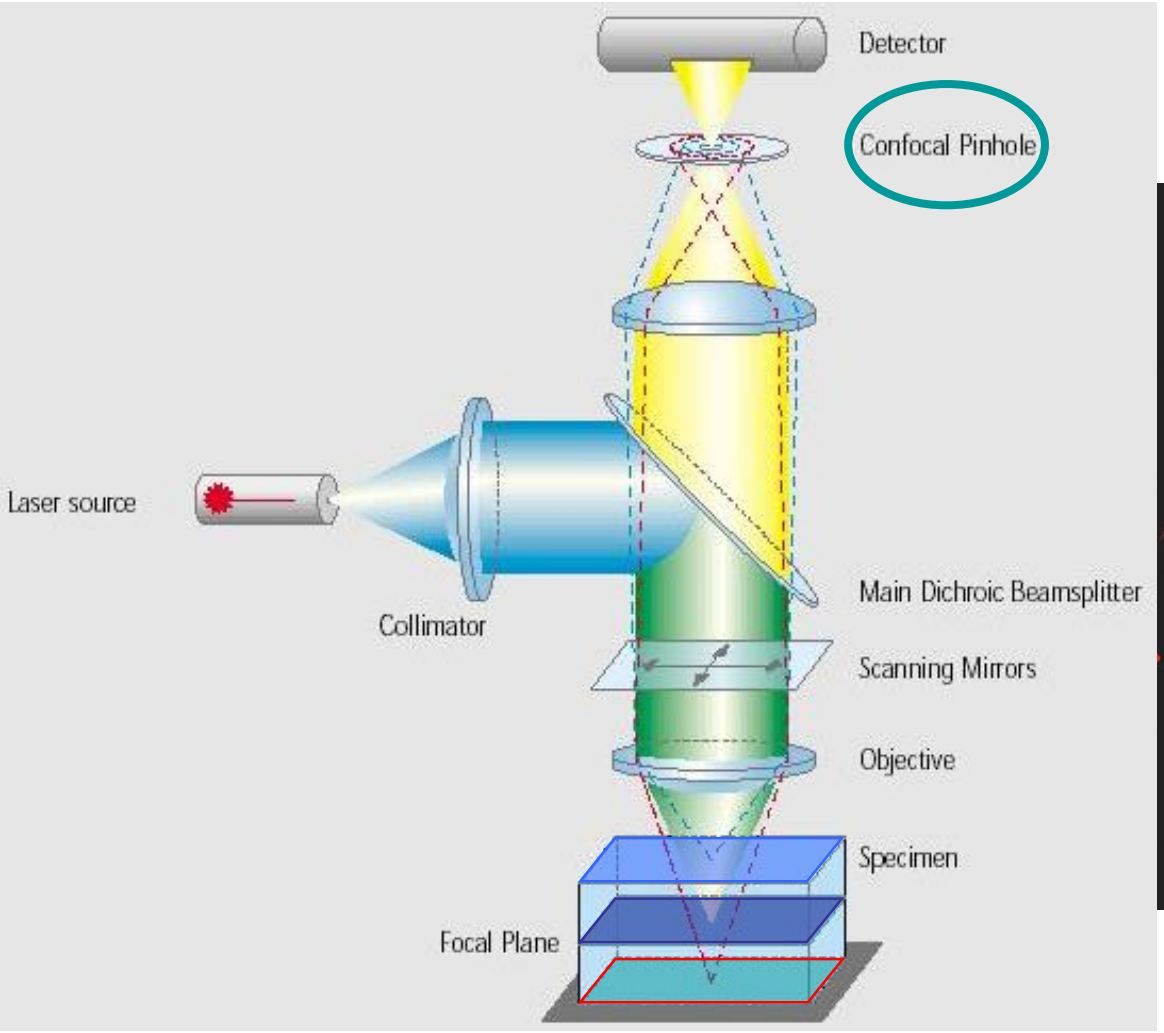


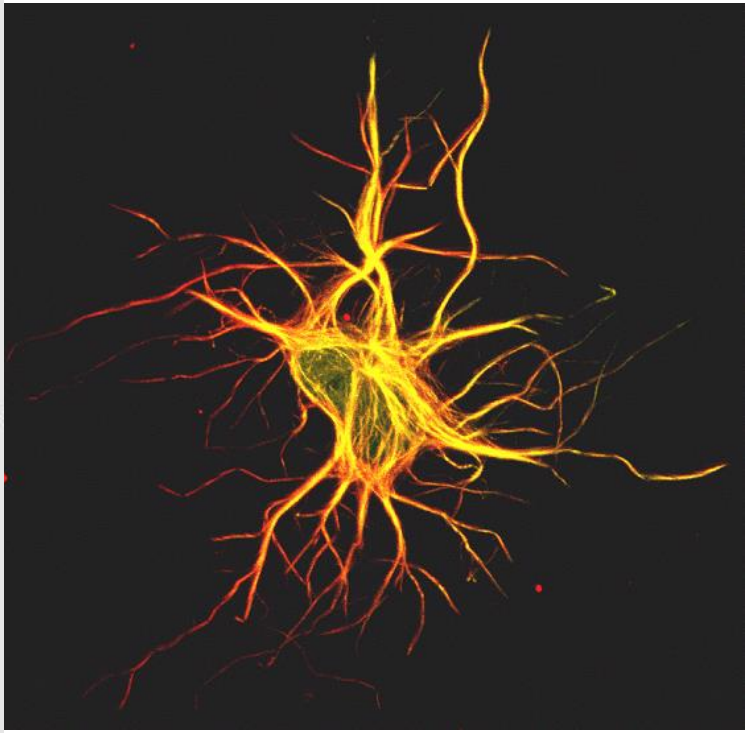
Figure 4

- **Improved resolution power (PSF) :**
  lateral resolution improved
  Real axial resolution power
- **Improved contrast:**
  Rasterizing the specimen, stray light due to scattering is suppressed
- **Multi-dimentional acquisition with digital image processing**

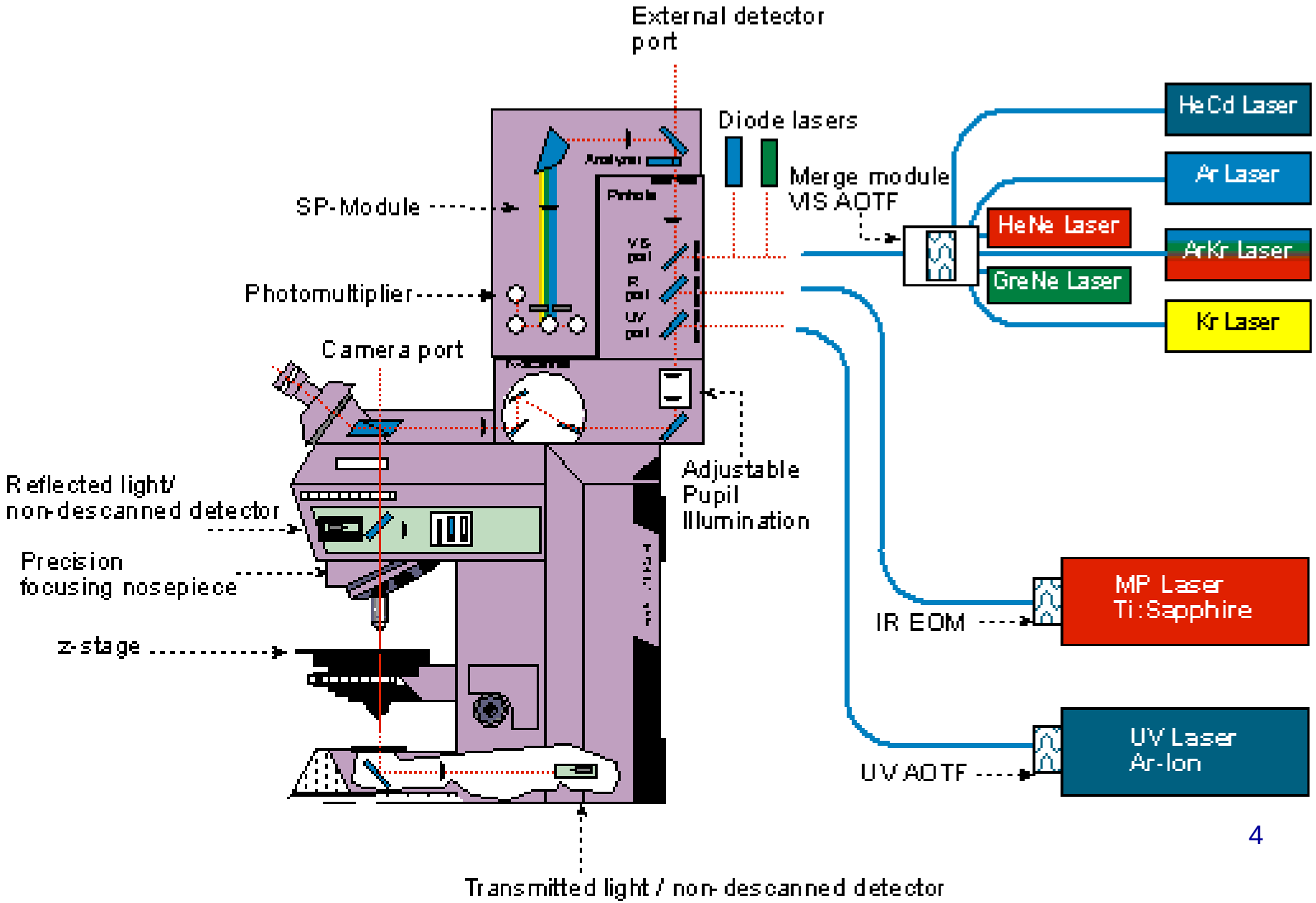
  X-Y-Z-T-I-θ-λ
- **New application, FRAP, FLIM, FRET, Cage, Bio-Mapping .....**

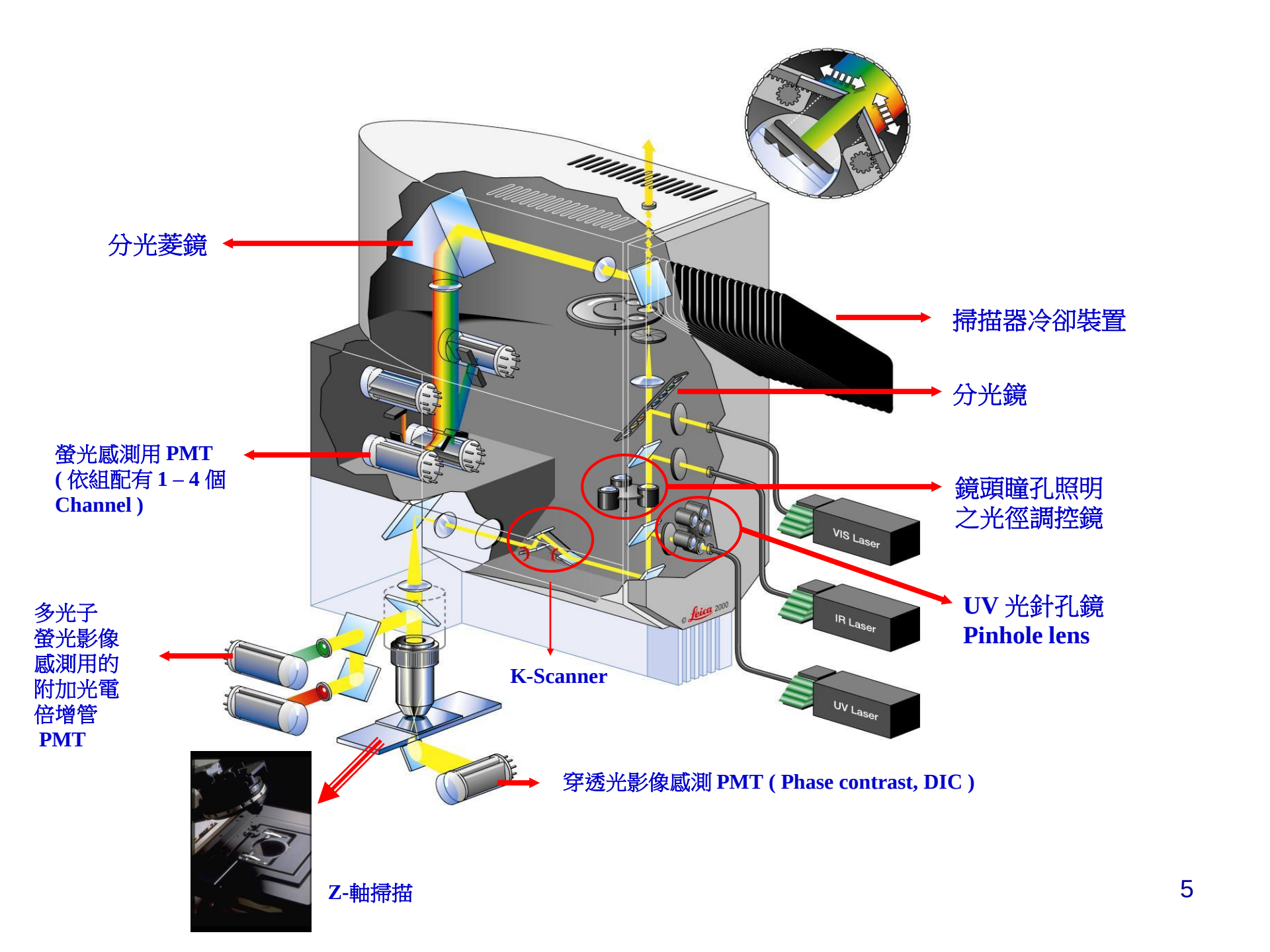
Principle of Confocal



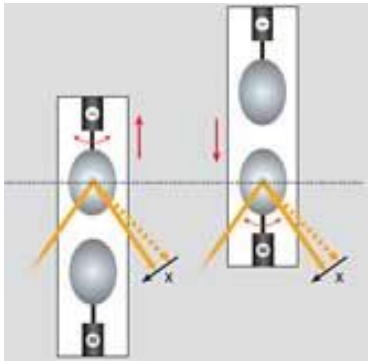


Leica TCS SP2/MP2: System Optics Overview




分光菱鏡
掃描器冷卻裝置
分光鏡
螢光感測用 PMT
(依組配有 1 – 4 個
Channel)
鏡頭瞳孔照明
之光徑調控鏡
VIS Laser
UV 光針孔鏡
Pinhole lens
IR Laser
多光子
螢光影像
感測用的
附加光電
倍增管
PMT
K-Scanner
UV Laser
穿透光影像感測 PMT (Phase contrast, DIC)
Z-軸掃描

Confocal Spectral Microscope Leica TCS SP5



Tandem Scanner 在同一個掃描器, 可切換使用兩組的掃描器, 完全微電腦控制.

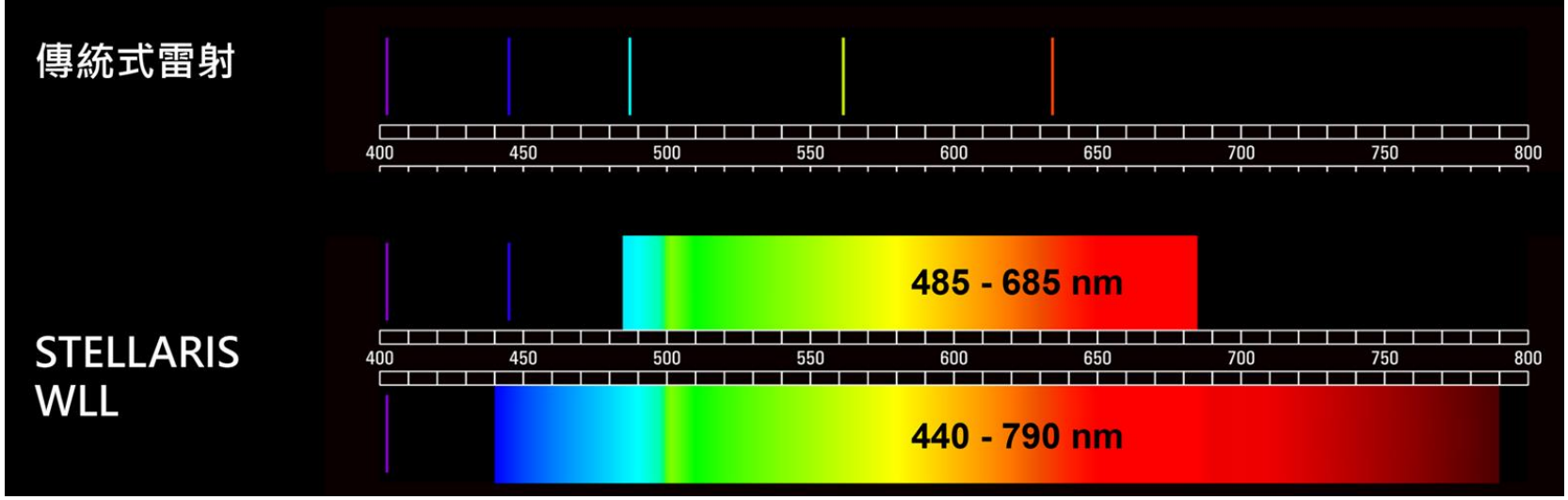
C Scanner 提供超高光學解晰的影像擷取 **Ultra High resolution image acquisition.**

R Scanner 提供超高速的影像擷取 **Ultra-Low Photobleaching image acquisition.**

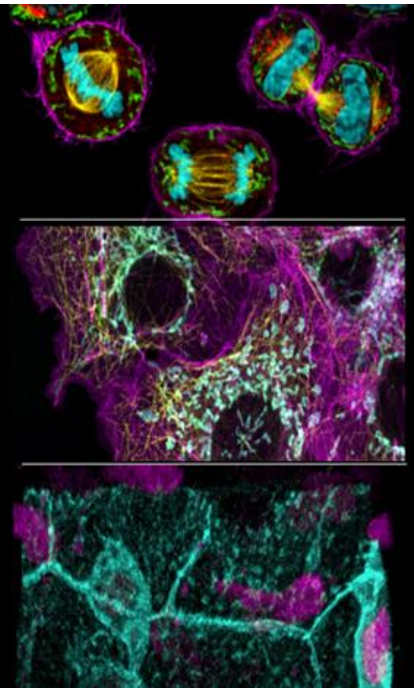
Conventional Scanner (C)		ResonantScanner (R)	
Max. line frequency	2800 Hz	Max. line frequency	16000 Hz
Min. line frequency	1 Hz	Min. line frequency	8000 Hz
Scan speed granulation	1400	Scan speed granulation	1
Max. frame rate 512 x 512	5 Hz	Max. frame rate 512 x 512	25 Hz
Max. frame rate 512 x 16	25 Hz	Max. frame rate 512 x 16	250 Hz
Beam park	Yes	Beam park	No
Max. frame resolution	8192 x 8192 pixels	Max. frame resolution	1024 x 1024 pixels
Scan zoom	1.0x - 32x	Scan zoom	1.7x - 32x
Panning	Yes	Panning	Yes
Field rotation	200° optical	Field rotation	200° optical
Field diameter	21.2 mm	Field diameter	14.8 mm
超高解析掃描 - 多重螢光影像擷像 (Multi-spectral image acquisition)		超高速掃描掃描器 - 多重動態螢光影像擷像 (Multi spectral image acquisition)	

LEICA STELLARIS 光譜式共軛焦顯微鏡系統

LEICA 新世代的全光譜白光雷射 WLL, 脈衝式.



POWER
SEE MORE



加強藍綠光譜代的偵測效益高達 56% 以上

可延伸遠紅光光譜代的偵測達 850 nm, 可增加到3個顏色的偵測與成像

脈衝式白光雷射, 連續光譜範圍 440 - 790 nm

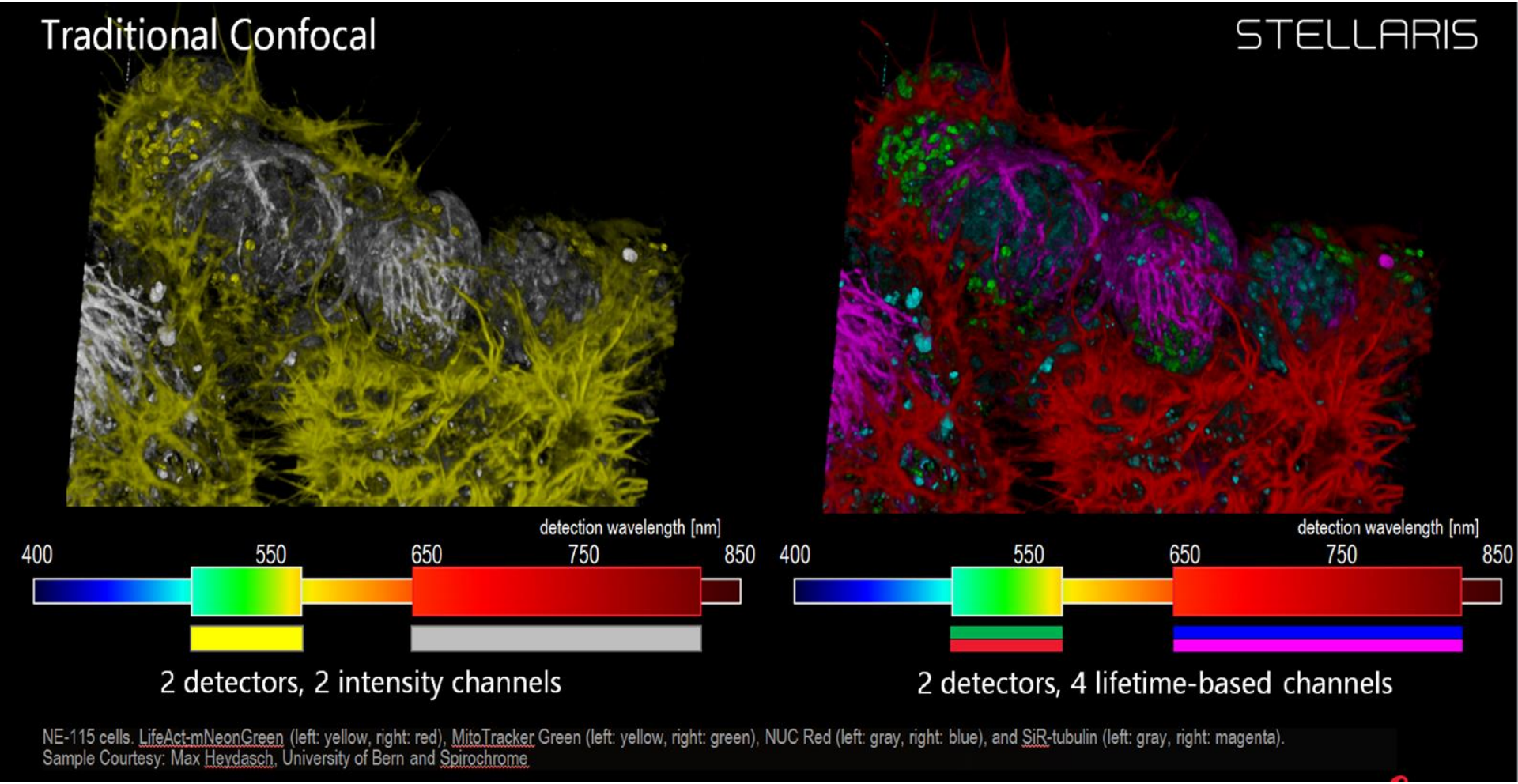
8-通道, 可調分光 AOBS

連續光譜偵測 410 - 850 nm

最適合多色螢光的活細胞影像工作

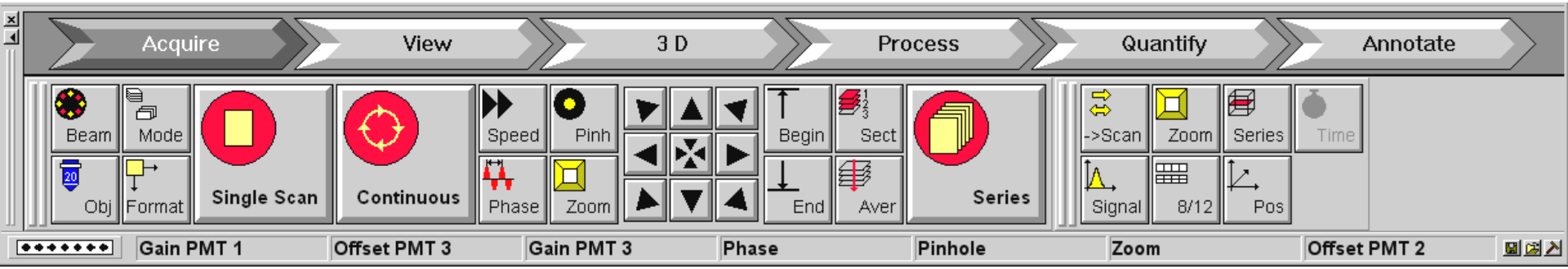
共軛焦影像新技術：時脈頻譜技術

TauSense :TauSeparation



Tau-Seperation 時脈頻譜技術是用來讀取螢光染劑的螢光衰減周期的時間訊號，依據每一種染劑的螢光衰減周期的時間，把訊號抽離分開，所以，可以取得清晰的多種訊號的螢光影像.

Mode : Scanning and Image Capture

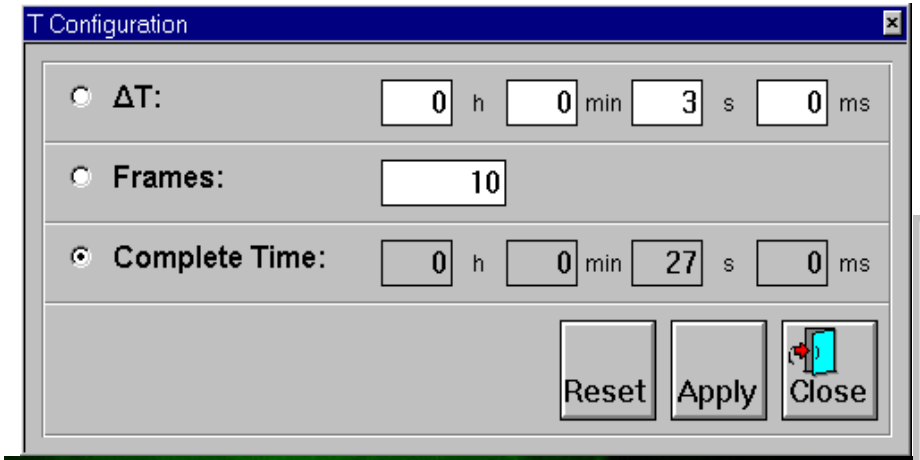


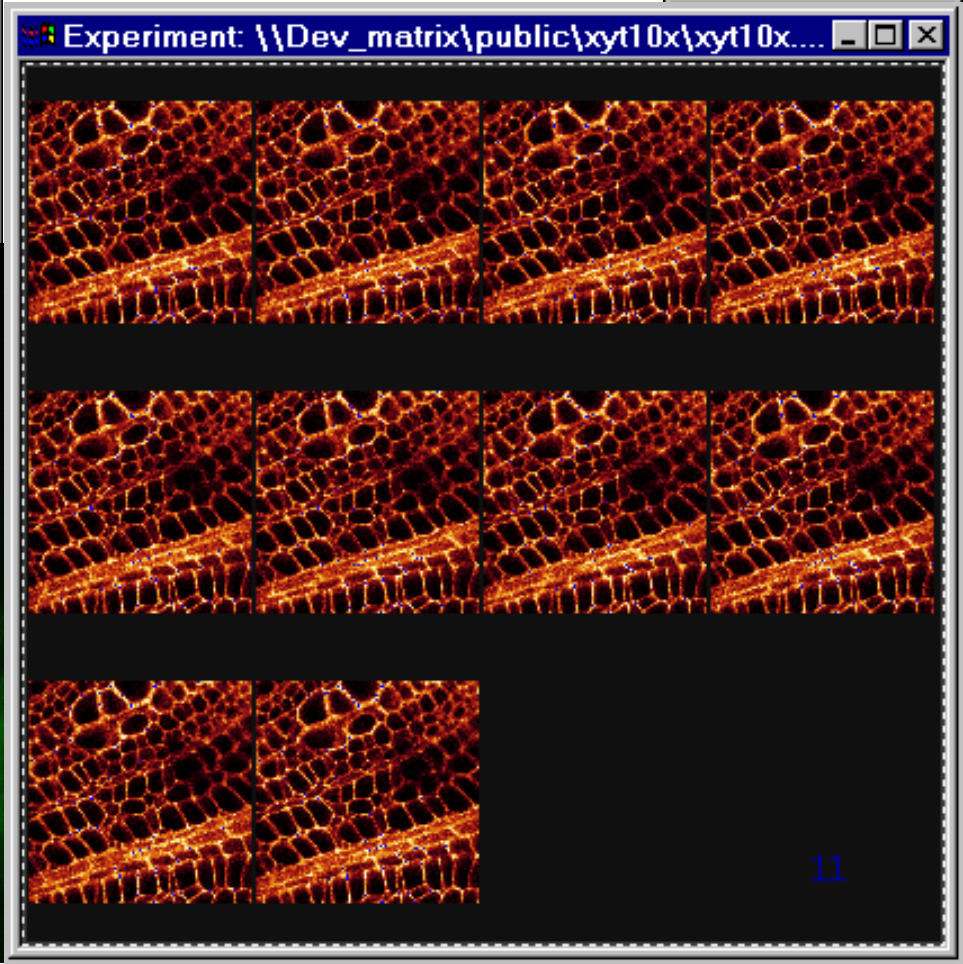
Mode	Functions
xyz	**An image stack is recorded from xy-sections in z-direction. (3D)**
xzy	An image stack is recorded from xz-sections in y-direction.
xt	A line is recorded several successive times.
xyt	An xy-section is recorded several successive times.
xzt	An xz-section is recorded several successive times.
xyzt	**An image stack is recorded from xy-sections in z-direction several successive times. (Example: drosomoitose)**
xyl	An xy-section is recorded at different wavelengths. (wavelength)
xzl	An xz-section is recorded at different wavelengths.

Frame-Mode xyt Configuration

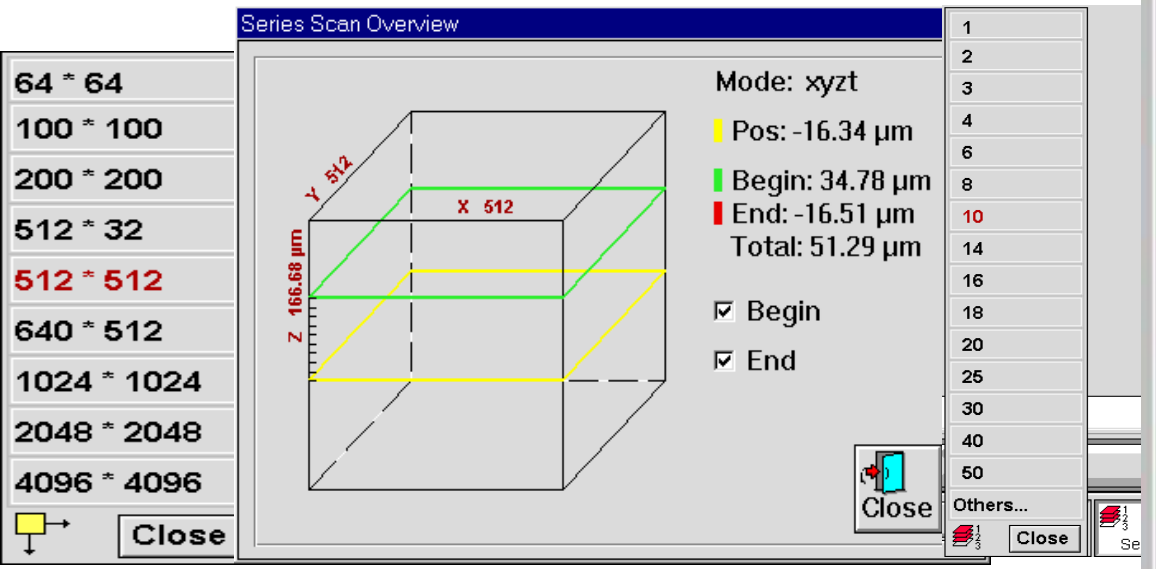
Time-lapse vs. Real Time (movie)

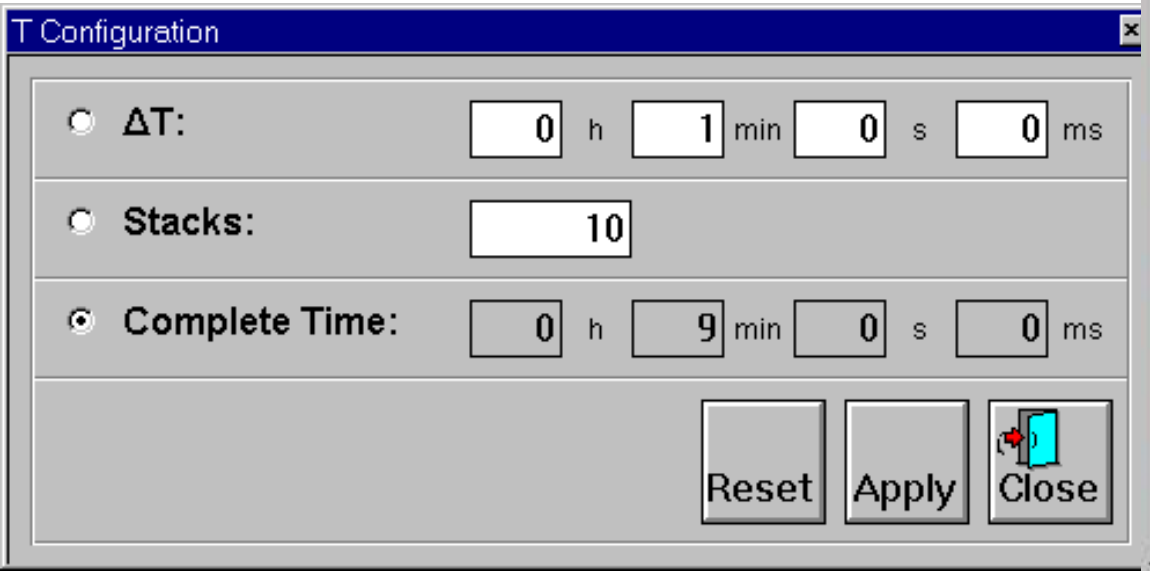
64 * 64
100 * 100
200 * 200
512 * 32
512 * 512
640 * 512
1024 * 1024





Stack-Mode xyzt Configuration





光電倍增器 Photomultiplier (PMT)

主要運用在分光譜後的共軛焦顯微鏡上共軛焦顯微鏡所使用的感測器是光電倍增器(PMT)所提供的感測器精密度達 0.1 nA, 俱有冷卻設計,可除去暗電流 (Dark current), 提供超高解析。

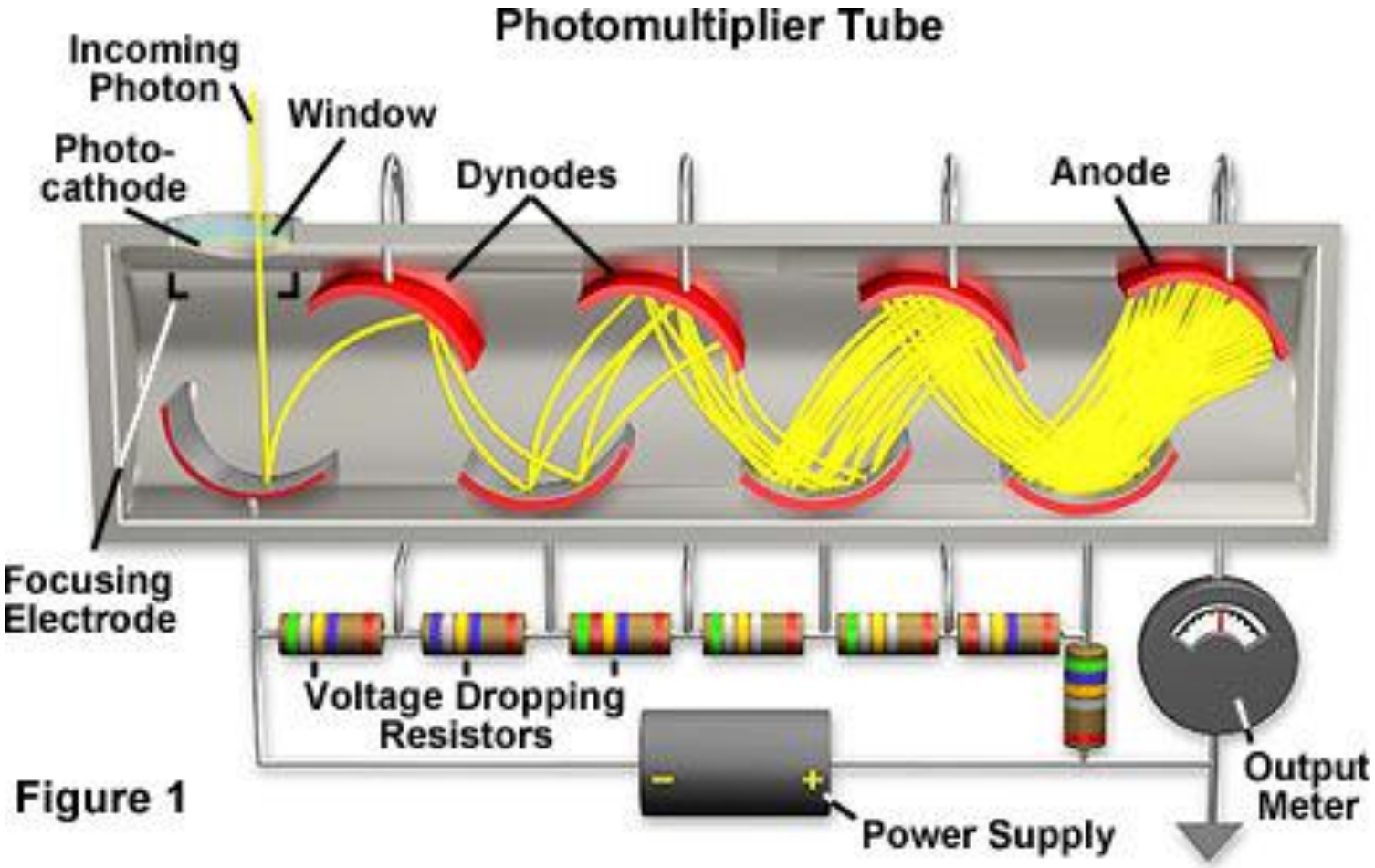
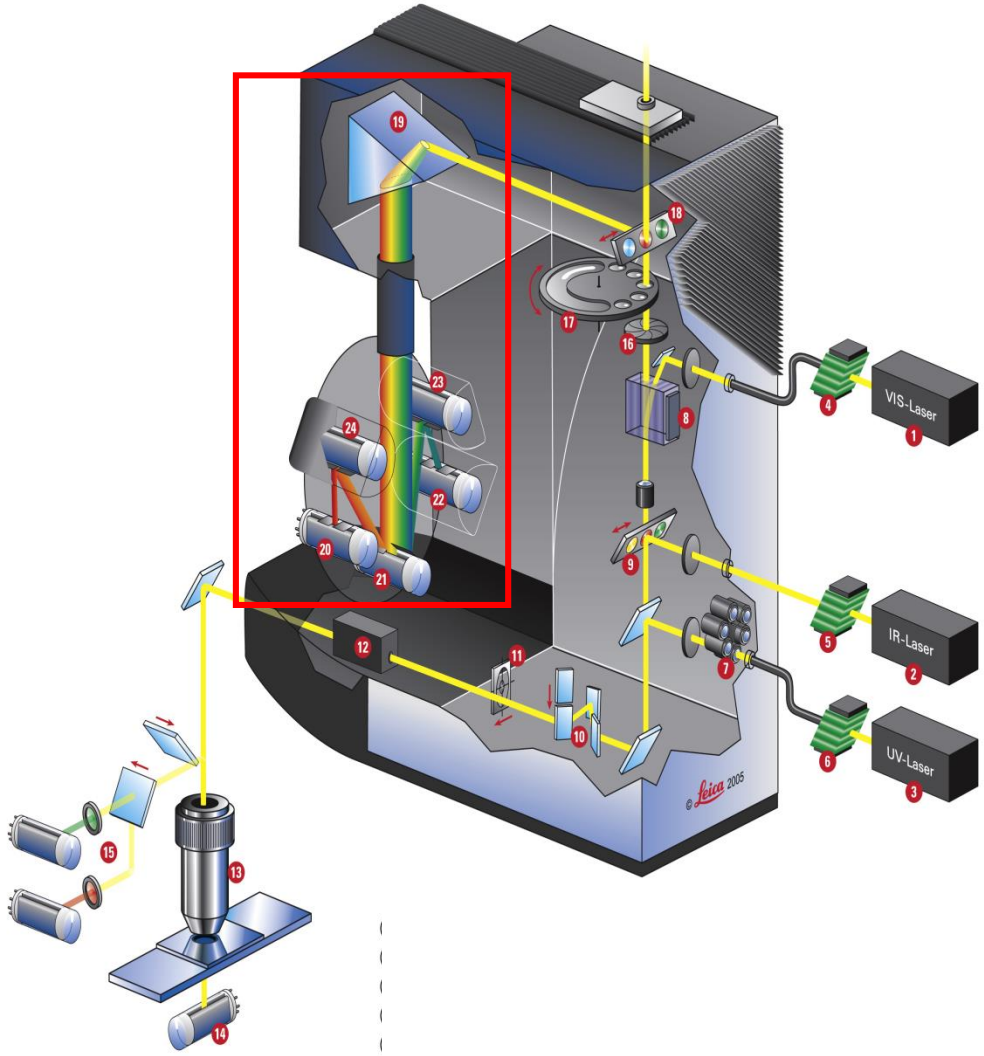


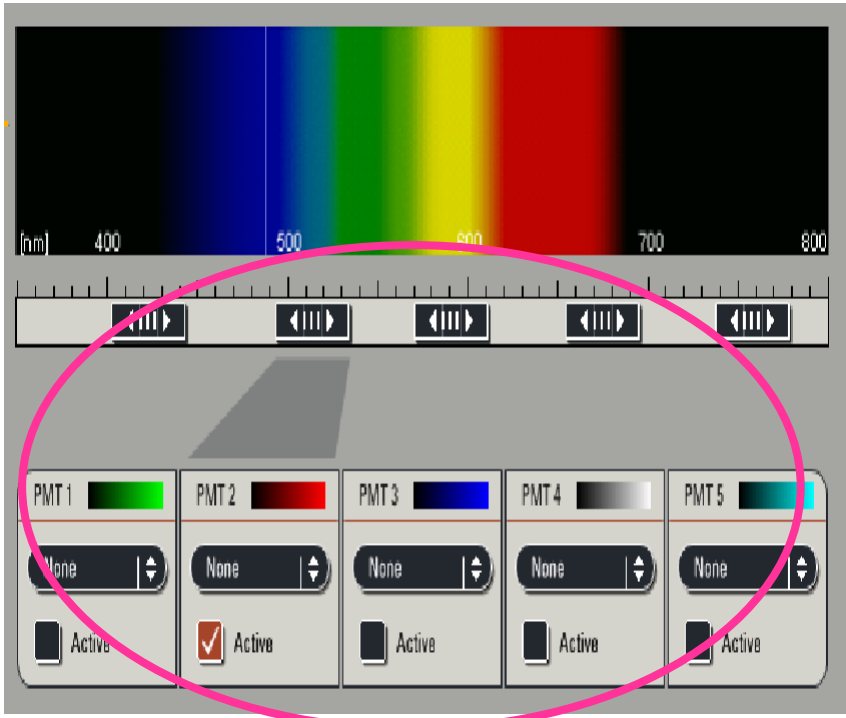
Figure 1

Spectral Base Detector

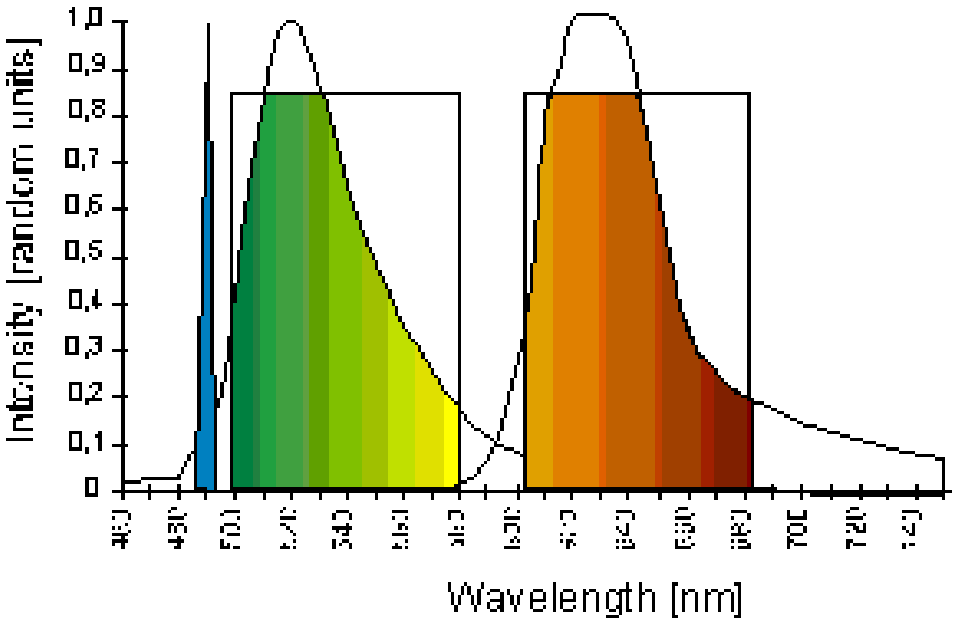
- Software Controller -

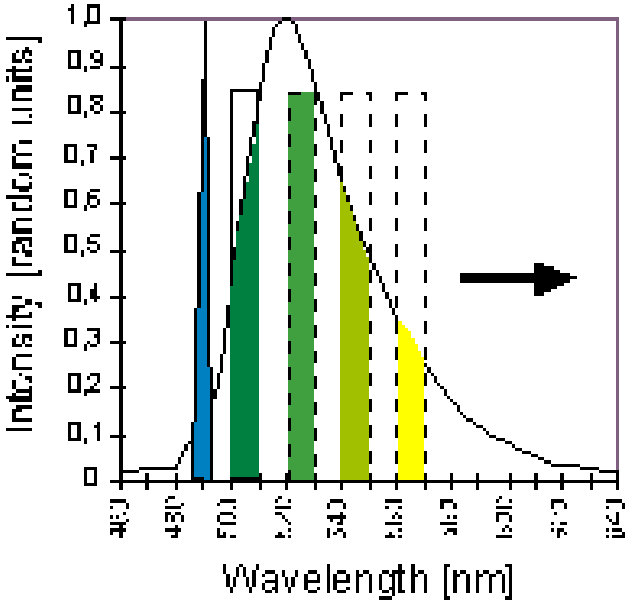


Spectral Based Detector



TCS SP/SP2: Prism Spectrophotometer Benefits



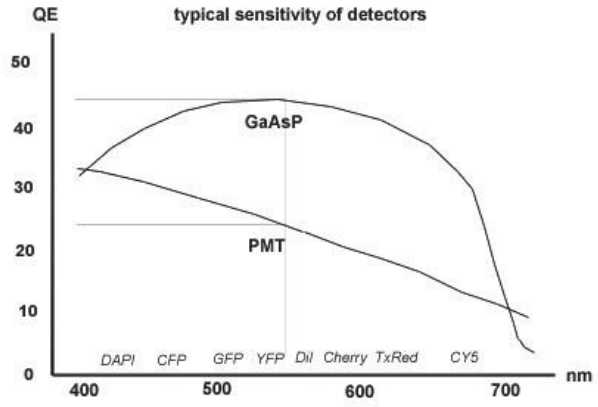


- Maximize efficiency
- Maximize flexibility
- Minimize crosstalk

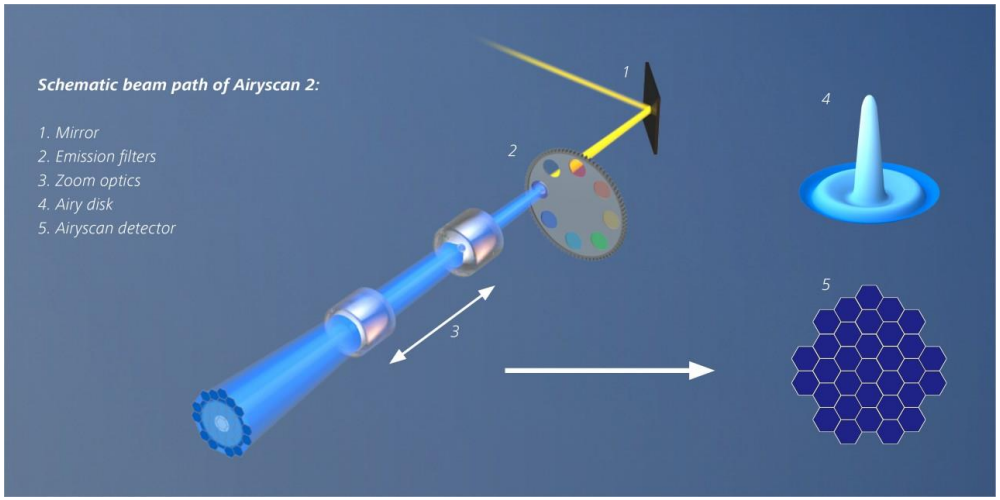
- Analyze the spectrum

ZEISS LSM 900 with Airyscan 2

Compact Confocal for Multiplex Imaging

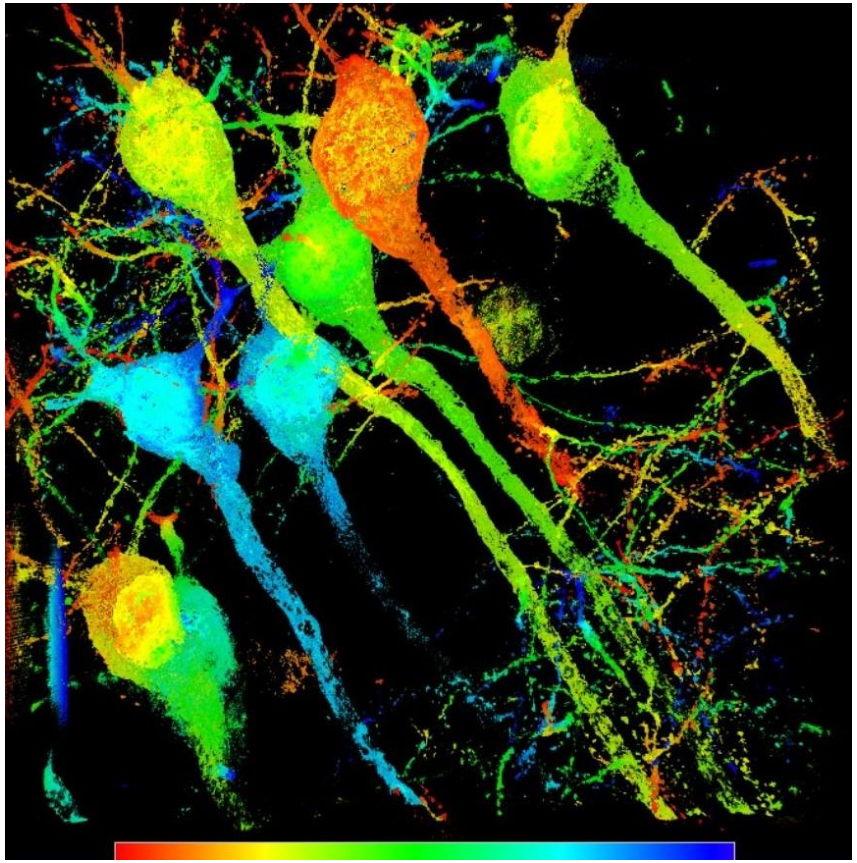


螢光感測器可以選擇傳統光電倍增管(PMT)或是磷酸砷化鎵(GaAsP)感測器

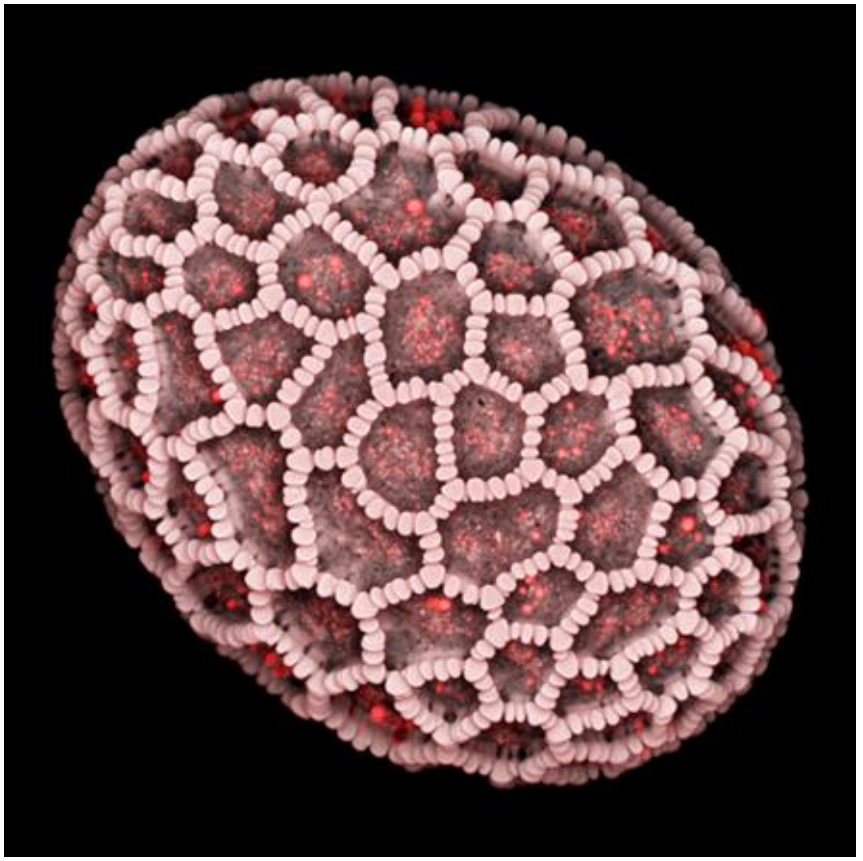


LSM900 同時支援*Zeiss* 最新技術 *Airyscan 2* XYZ解析度同步提升2倍的超高解析技術

ZEISS LSM 900 with Airyscan 2
Compact Confocal for Multiplex Imaging



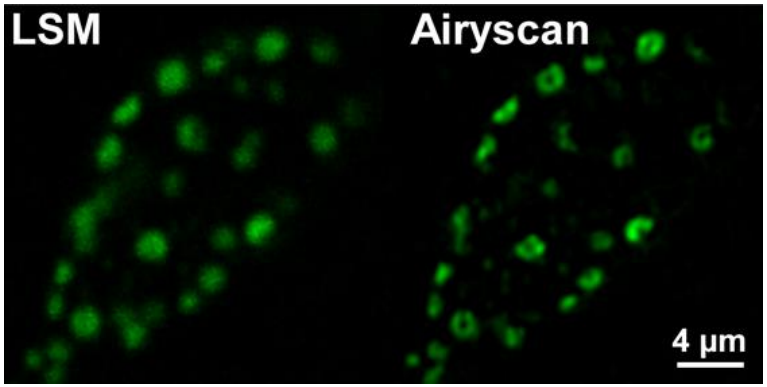
LSM 900 Neurons DepthCoded 3D, Fluorescence



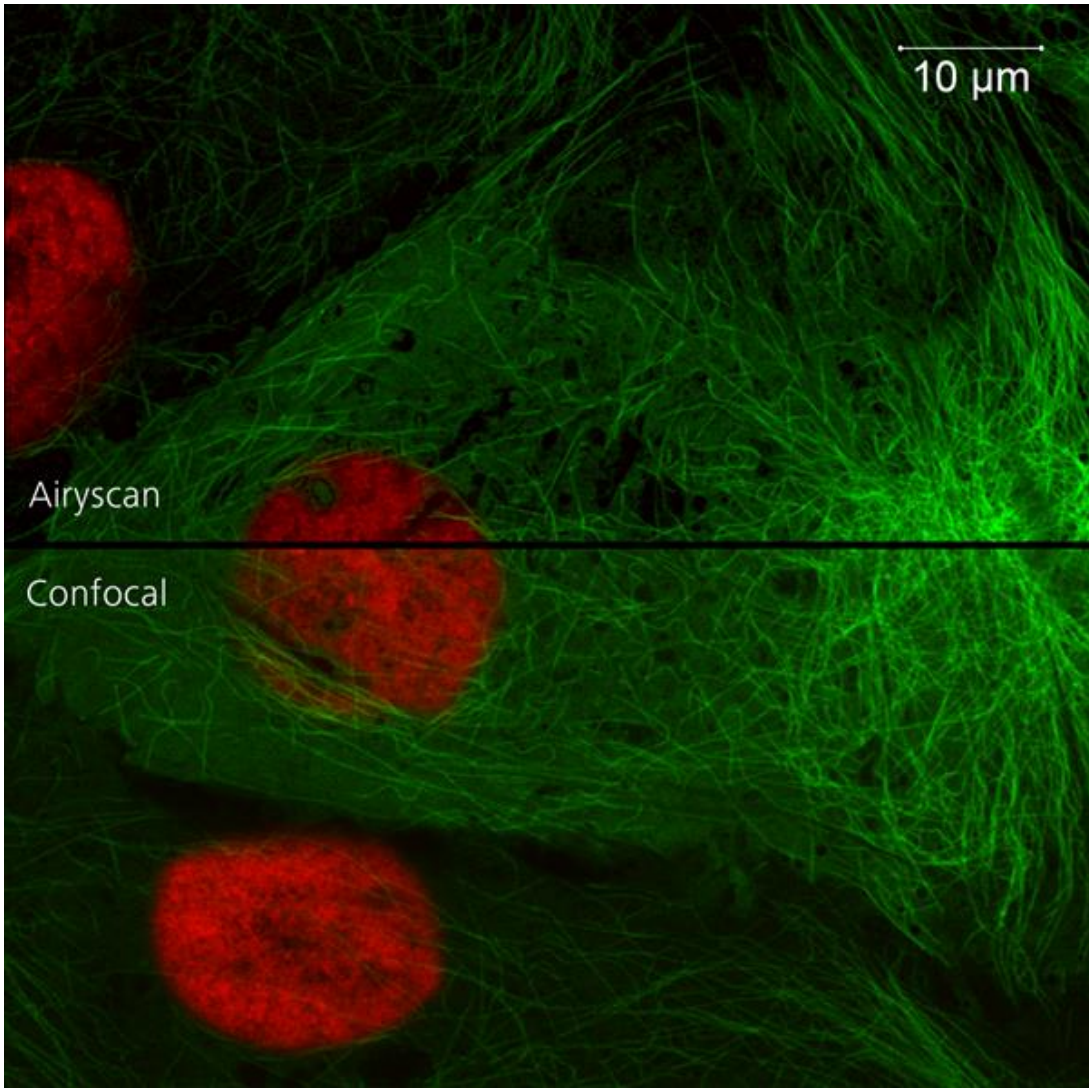
The micrograph shows a Lilium auratum pollen grain, acquired *with Airyscan* 2 in Multiplex mode. Image courtesy of Jan Michels, Zoological Institute, Kiel University

ZEISS LSM 900 with Airyscan 2

Compact Confocal for Multiplex Imaging



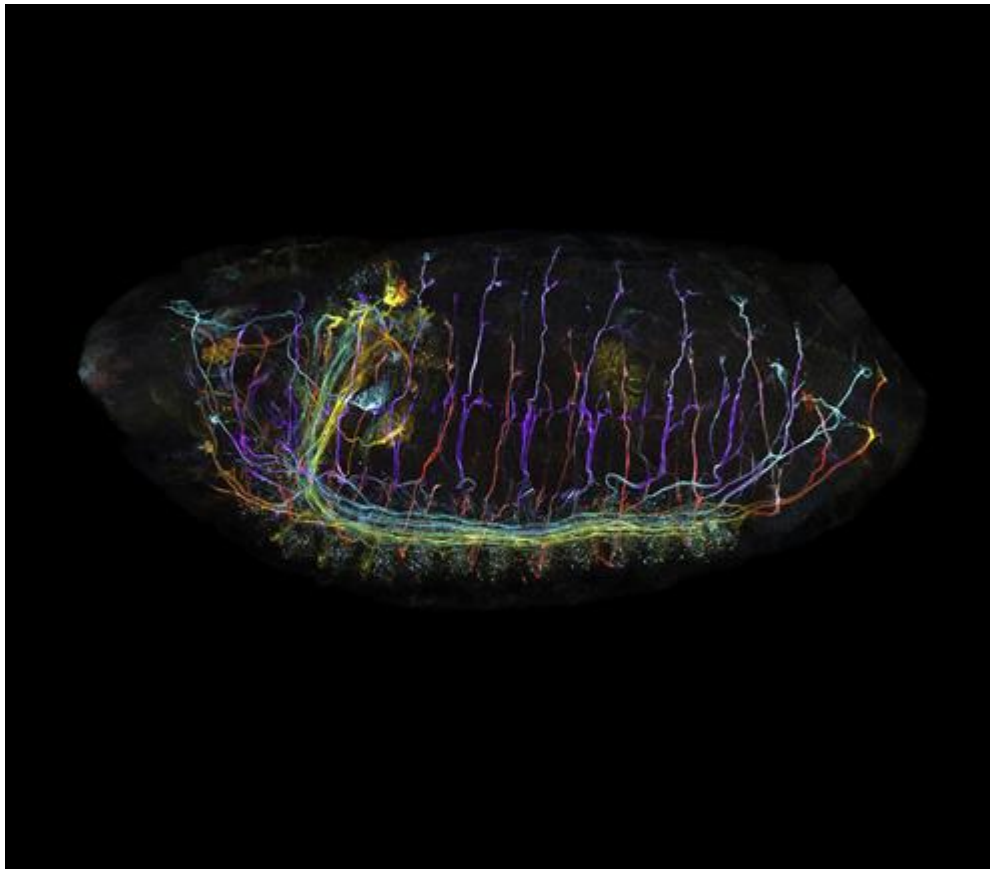
Drosophila brain, neuromuscular junction stained for Bruchpilot (BRP), comparison between confocal *LSM* and *Airyscan*.



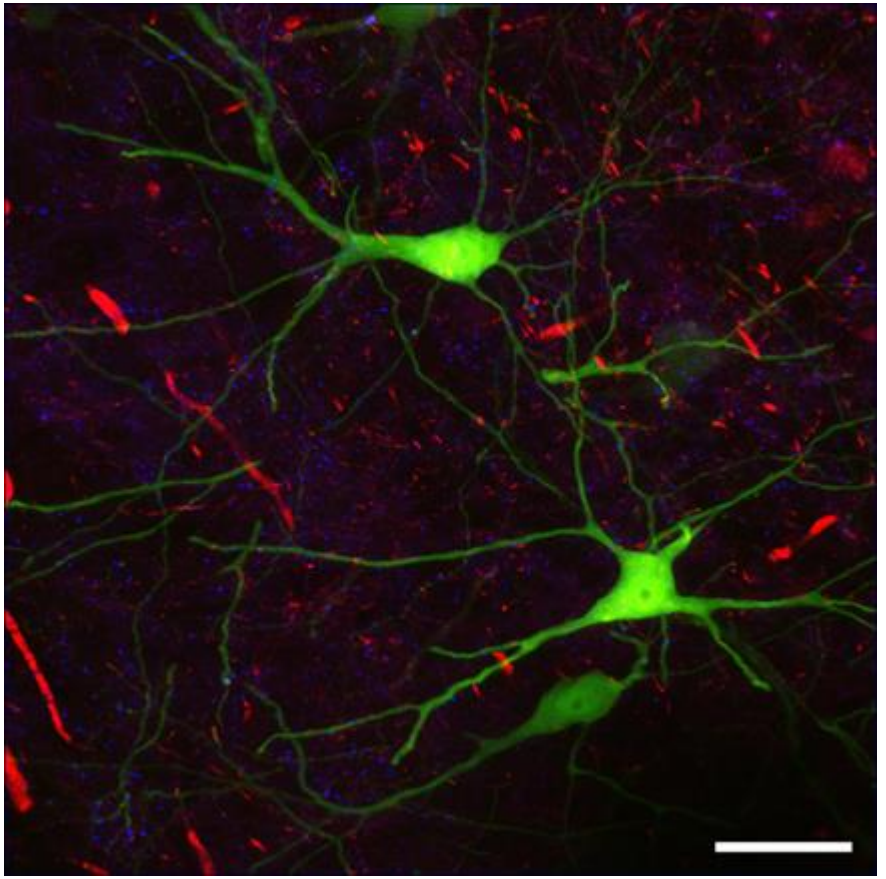
Living Pig Kidney Epithelial cells (LLC-PK1), green: Tubulin-eGFP, red: h2b-mCherry; Imaged *with ZEISS LSM 800 with Airyscan*, Plan-Apochromat 63x/1.4 Oil,

ZEISS LSM 900 with Airyscan 2

Compact Confocal for Multiplex Imaging



Drosophila ZEN Connect 1-01 *Airyscan* Processing-01-Stitching-02-Color-coded Projection-04-2

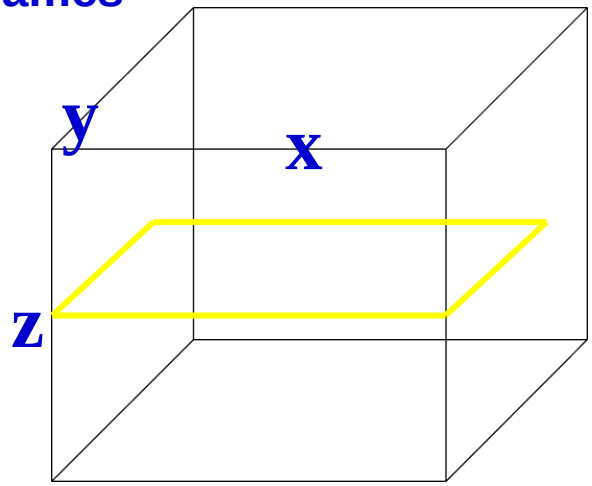


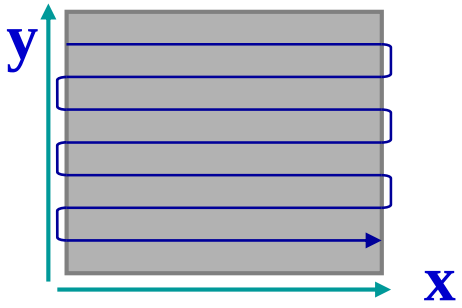
Mouse brain slice; EGFP-Thy1 (green): nerve cells (subset), Calretin-Cy3 (red): Calretinin-expressing neurons,GAD65-Cy5 (blue): GABAergic synapses. Scale bar 50 µm.[19]

Application

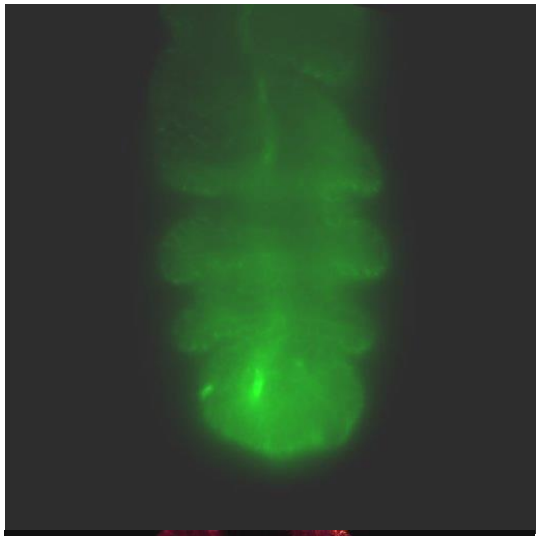
xy

Acquisition of a single frames

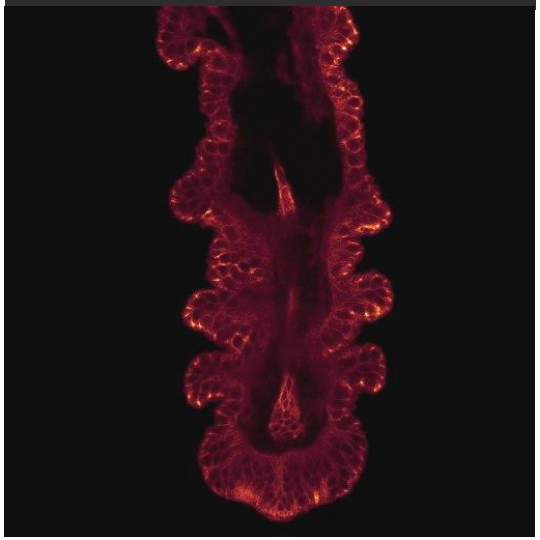




Drosophila leg, FITC

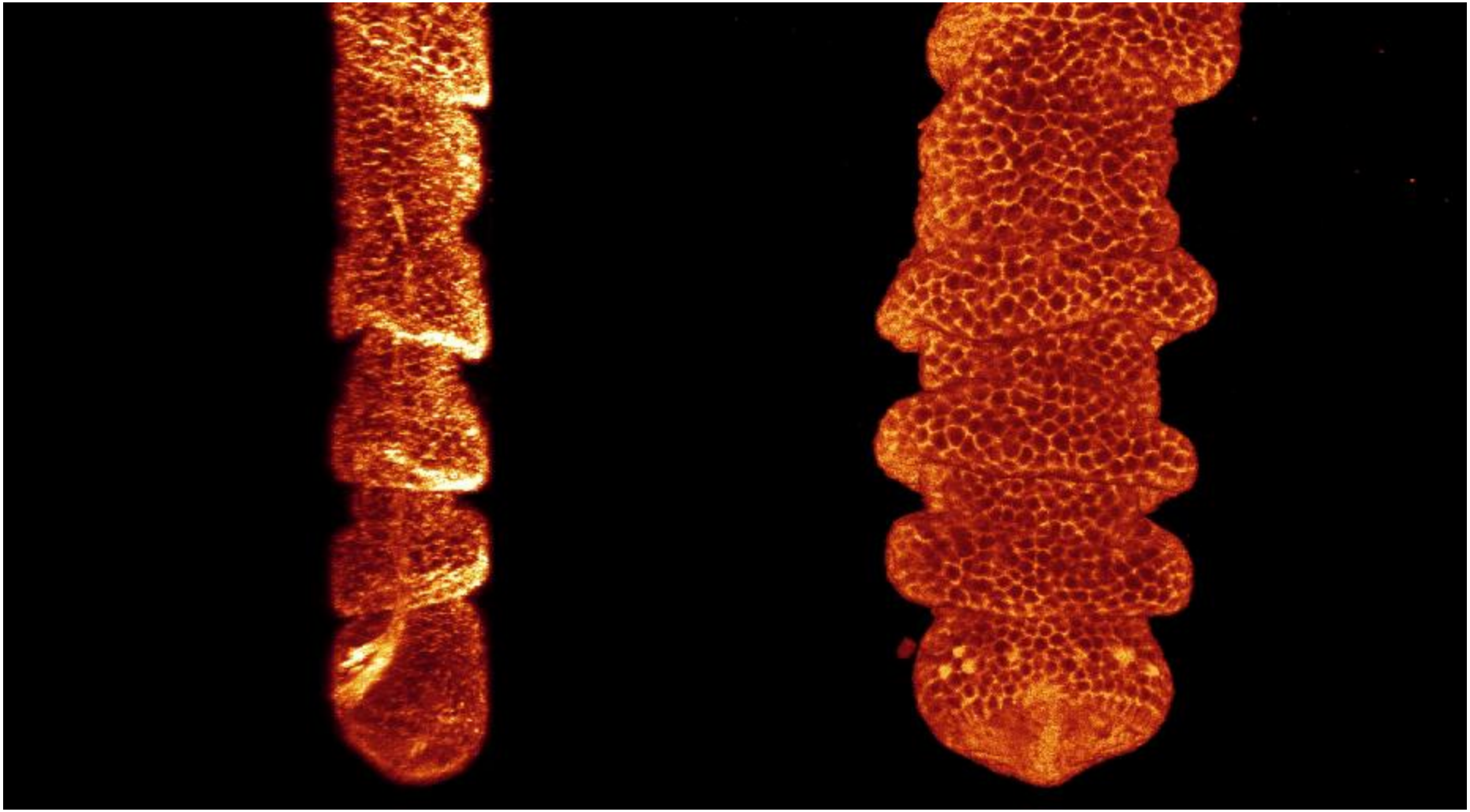


Non-confocal



Confocal 3D-structure

xyz projections: different algorithms

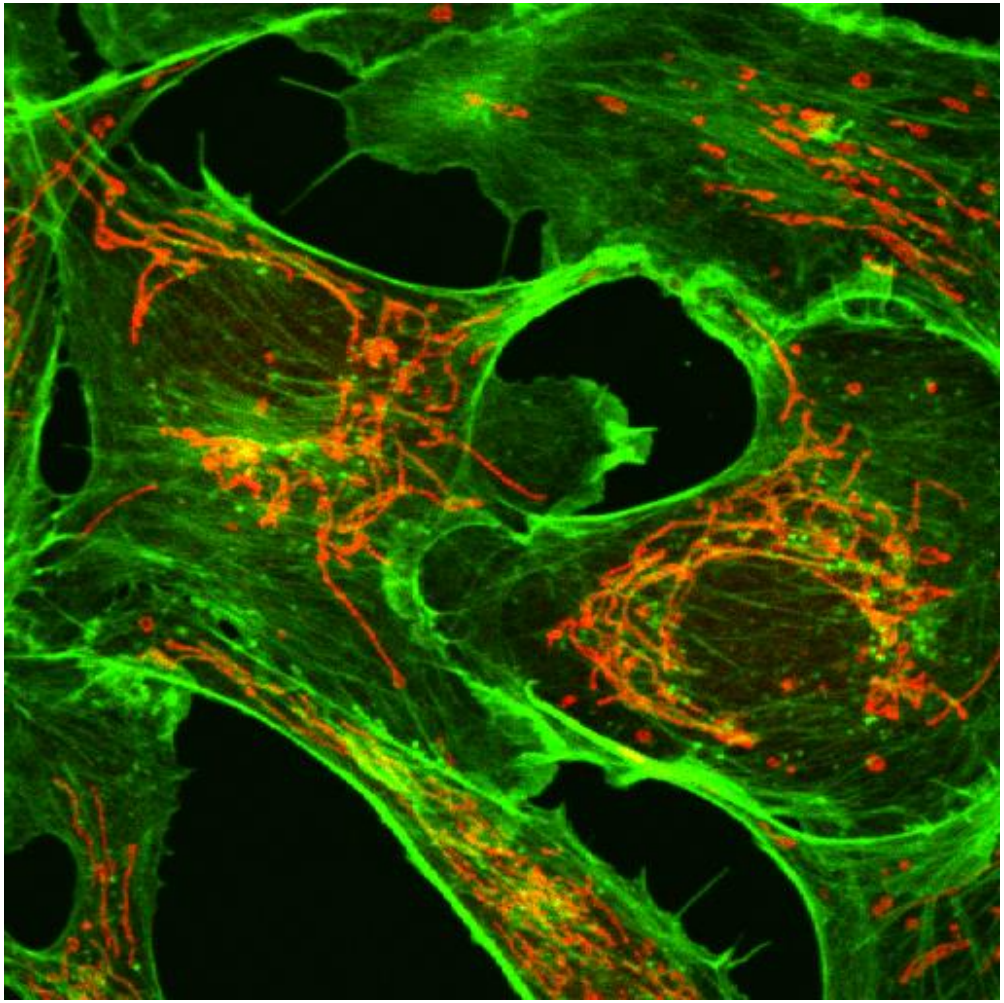


Drosophila leg, FITC, projection

Surface rendering

xy scanning

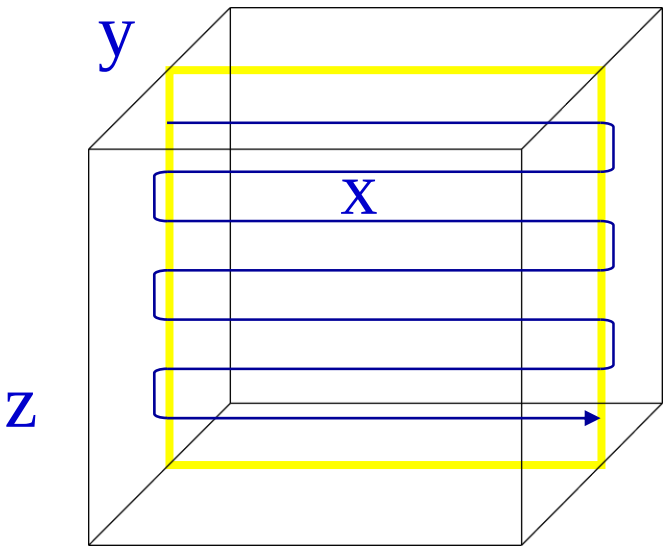
- Sample overview
- Colocalization studies
- Resolution-enhanced, high contrast images



Endothelial cells
FITC (Actin)
Mito-Tracker

xz

Beam is scanned in x-direction
Sample is moved in z (z-stage)



Z resolution depends on axial resolution of objective, generally 2x less than in xy
xy: 180 nm, z: 360 nm

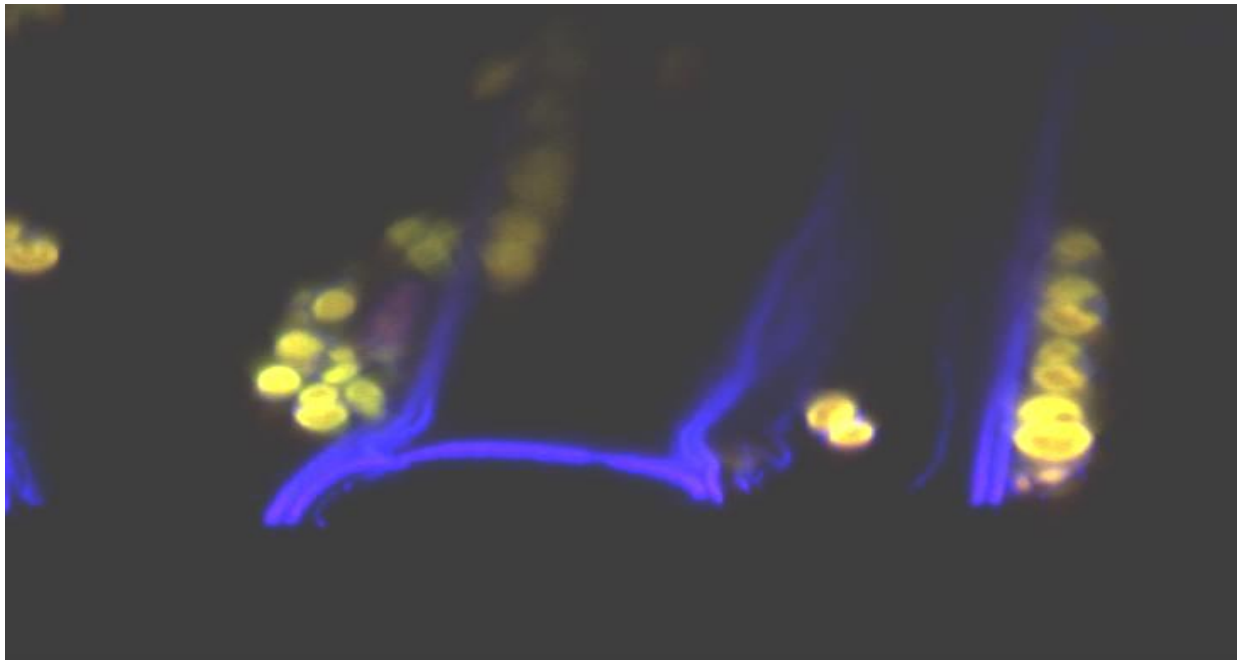
- Orientation of sample
- Spatial relations between structures in z
- Polarized cells

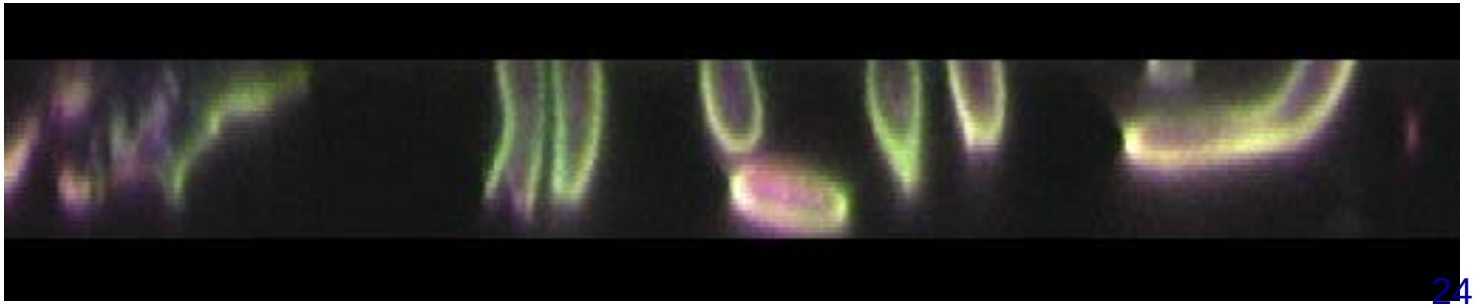
xz scanning

- Up to 20 frames per second with the Leica TCS SP2!

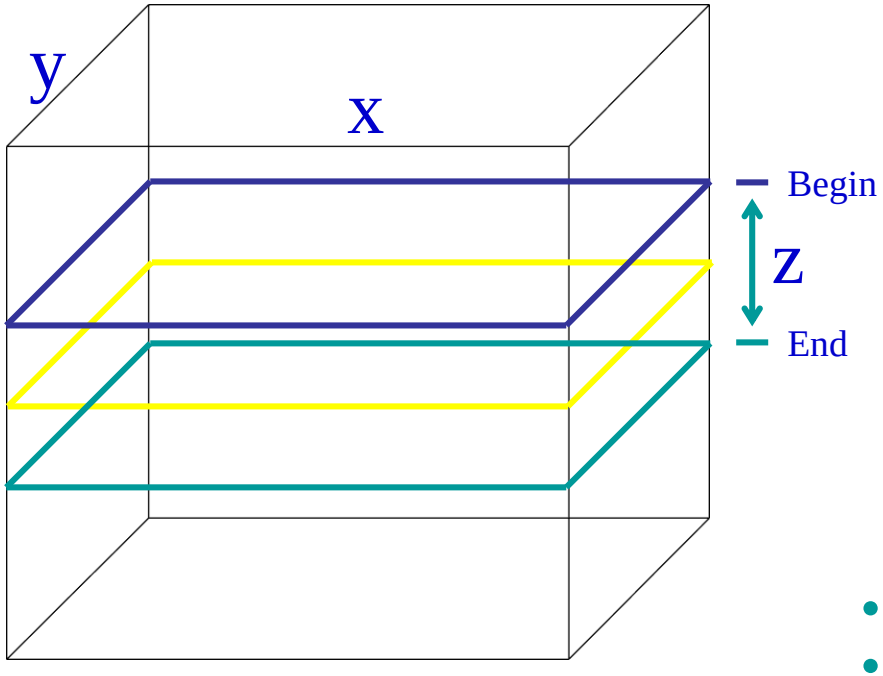
Convallaria

Starch grain
Cell wall





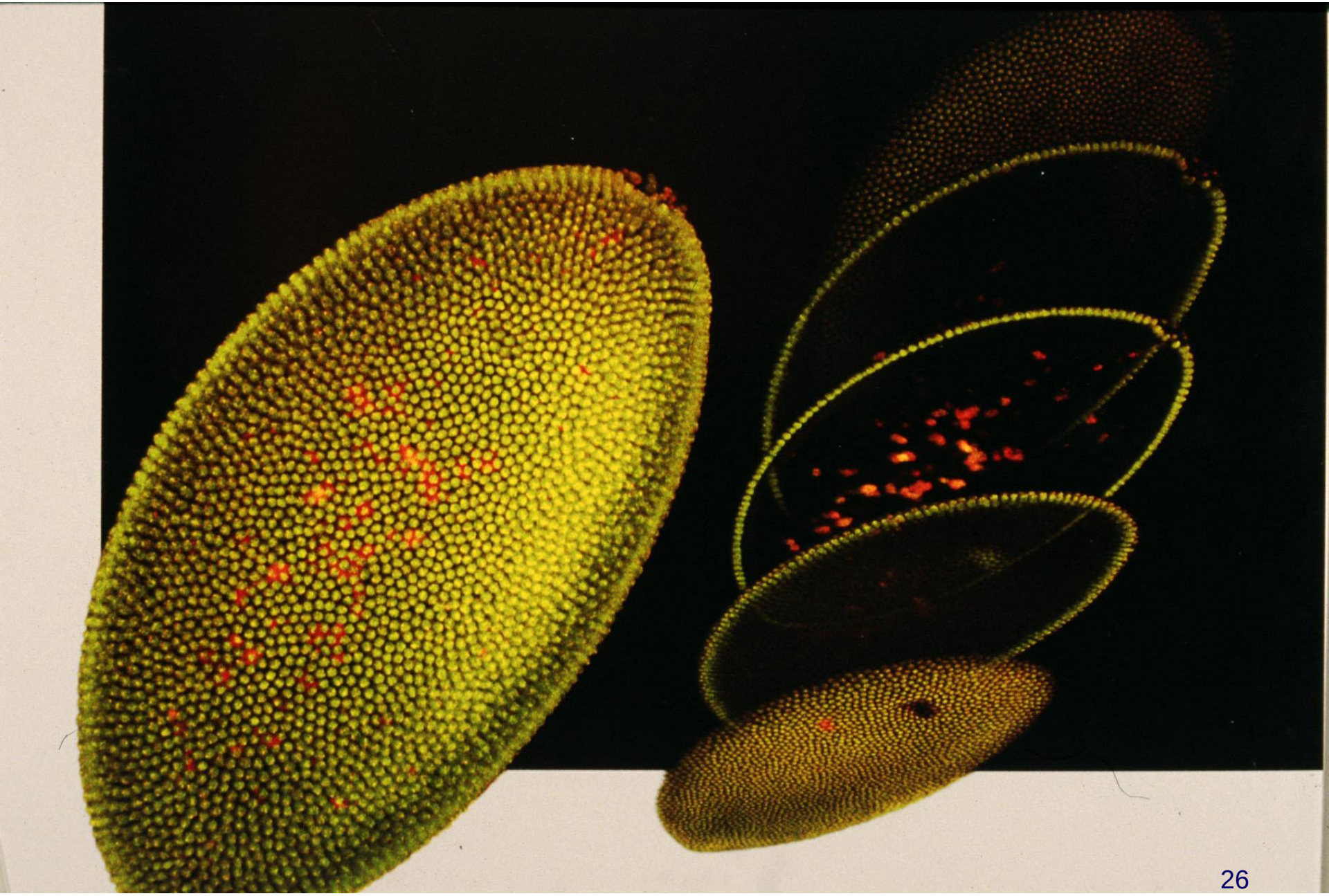
xyz



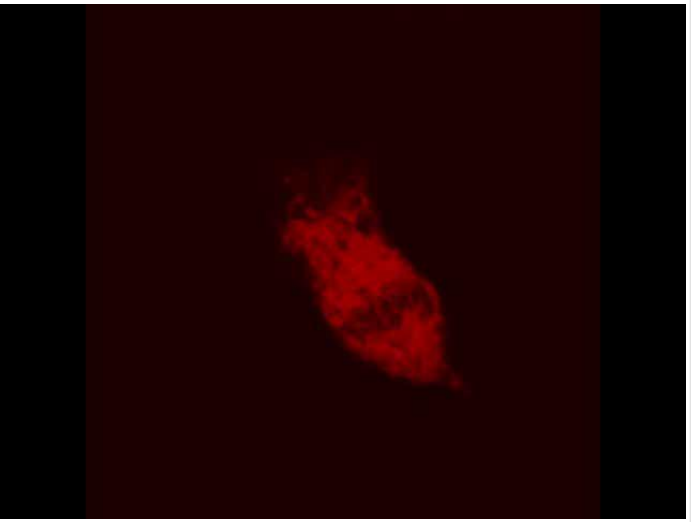
Beam is scanned in x and y direction and sample is moved in z via galvo stage or electronic focus of microscope

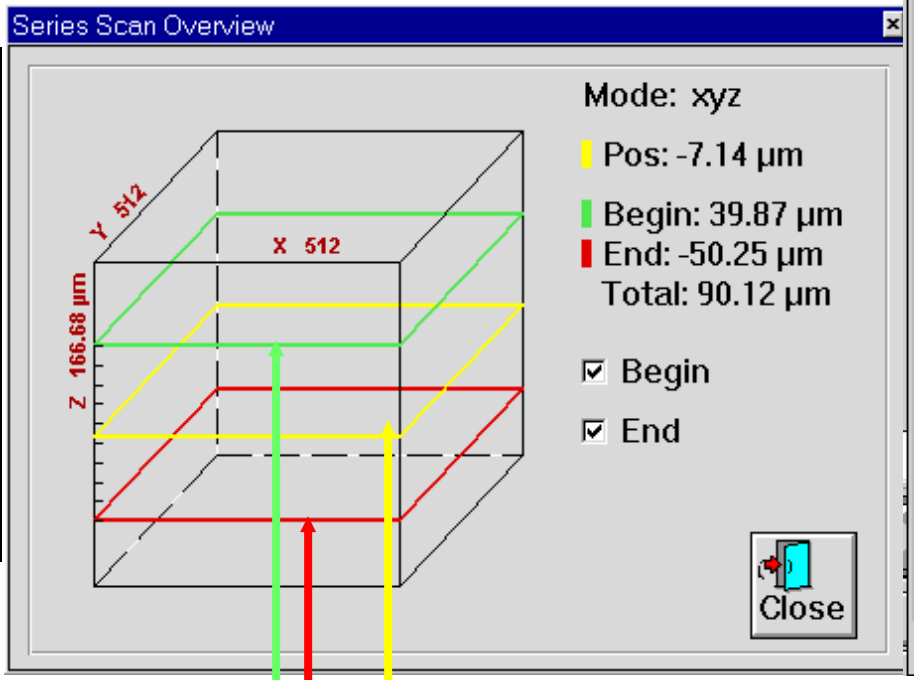
- Developmental Biology
- Neuroscience

- Optical sectioning,
- 3D stacks
- 3D projections
- 3D Animations
- Structural information from large focal depth – just depending on the stack size!

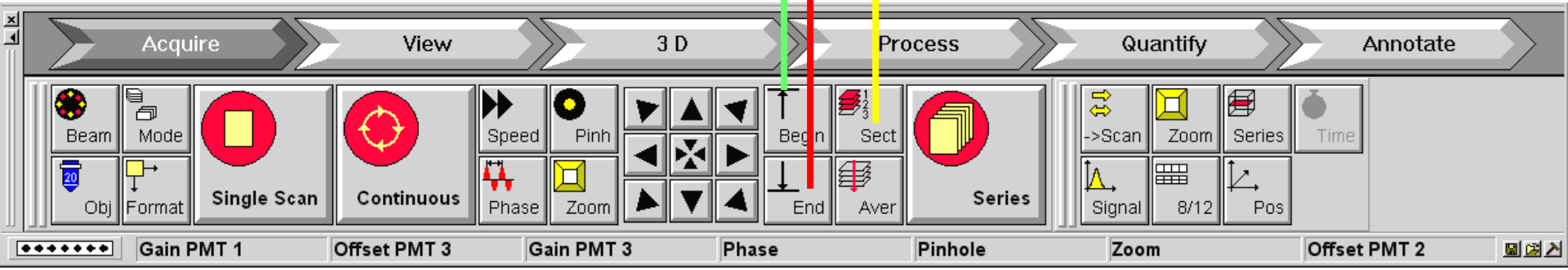


3D (xyz) series
Continuous scanning



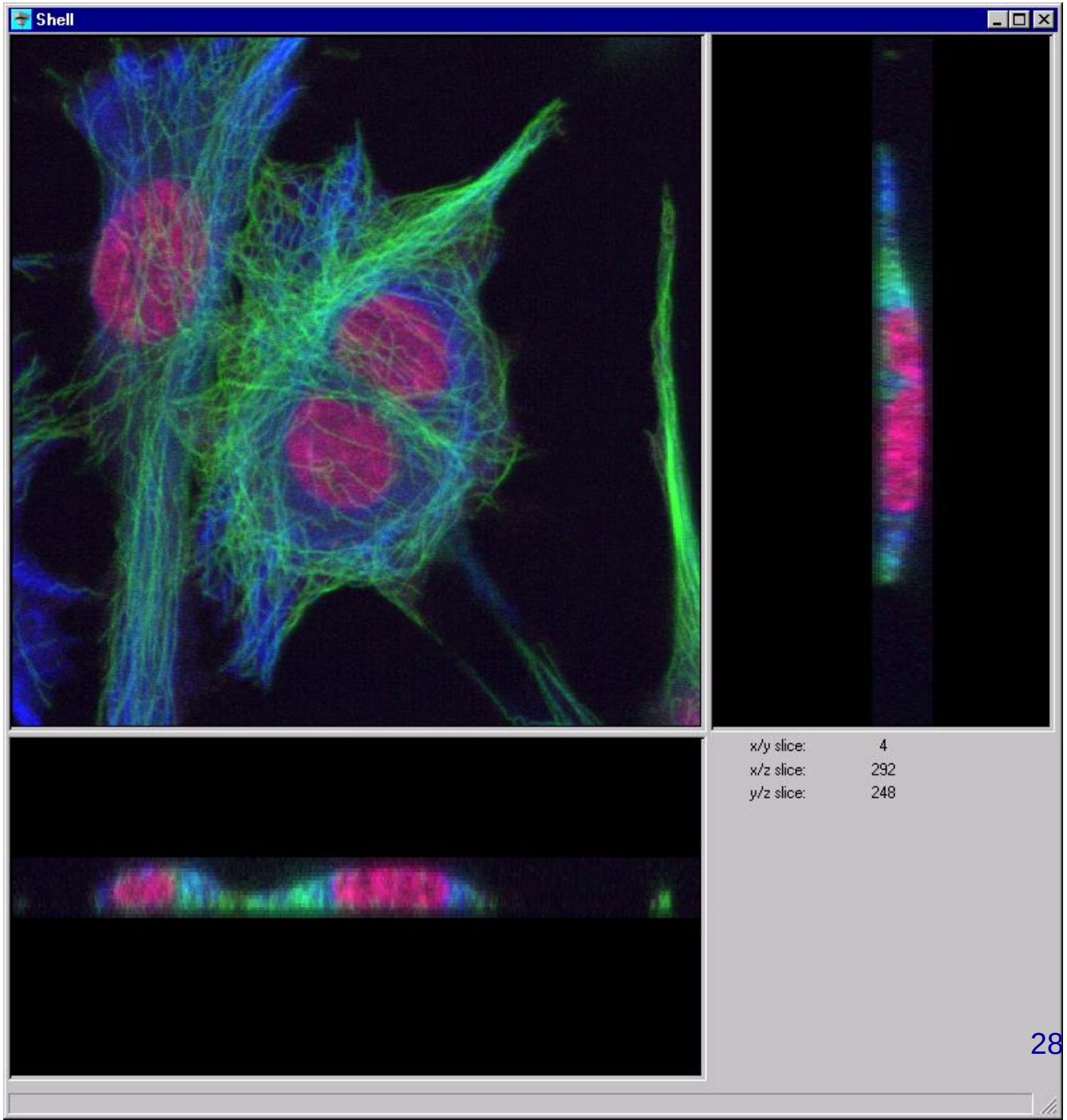


Number of optic sections

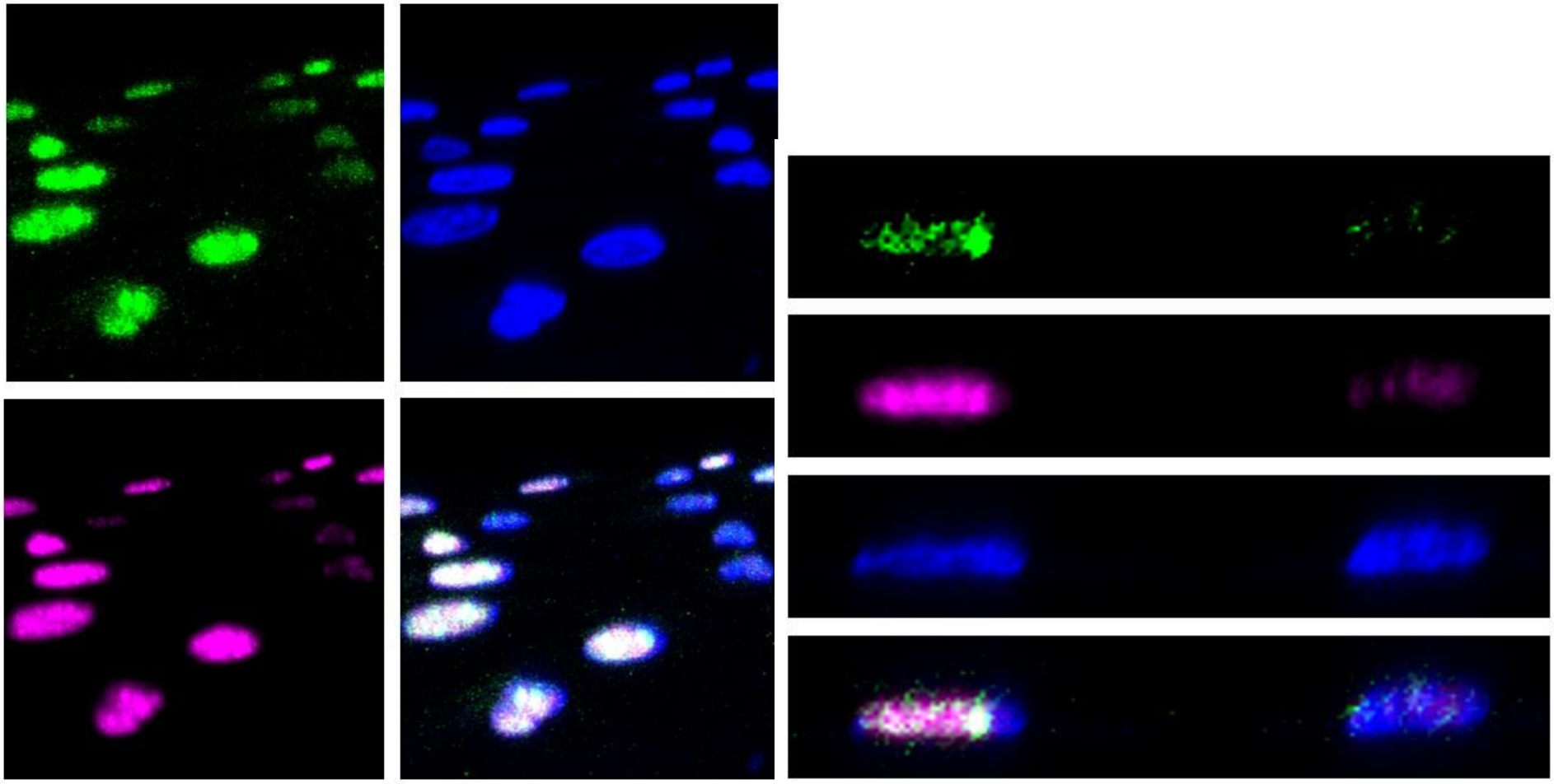


Application

3D-Section

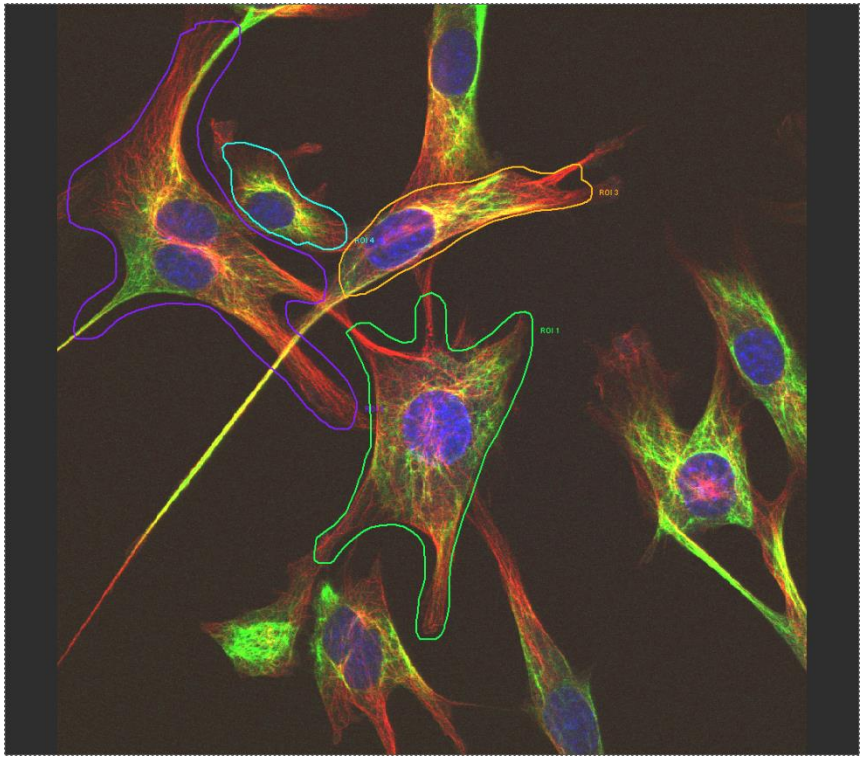


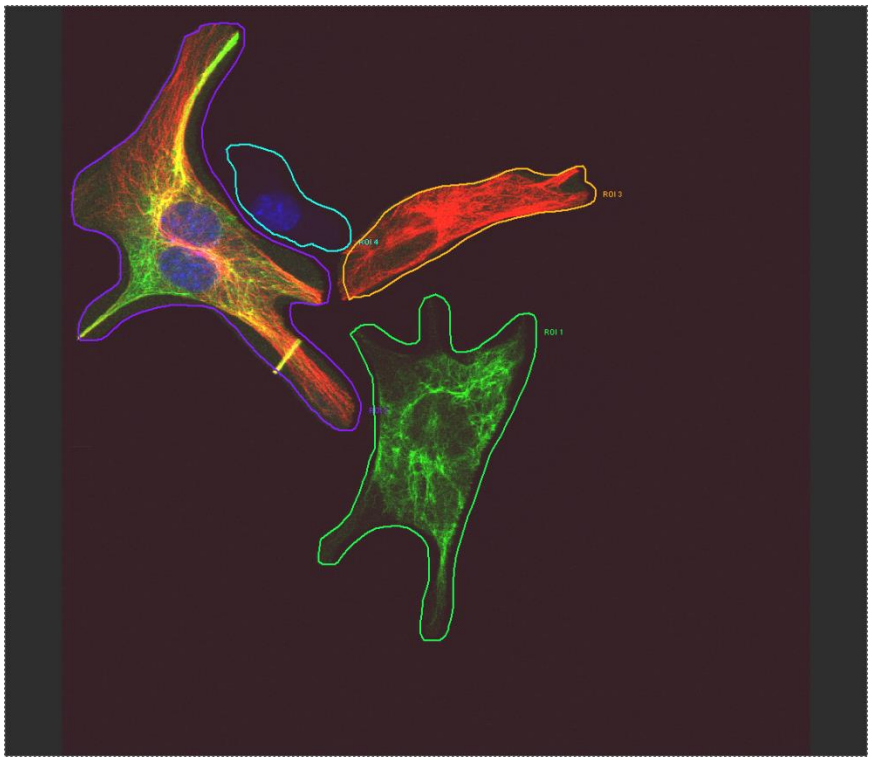
XYZ position precision photolabeling



Two-photon excitation microscopy provides x-y-z 3D axes precision photolabeling of targeted cellular or subcellular structure with a resolution up to 250nm.The figure shows xyz precision labeling of nuclei.

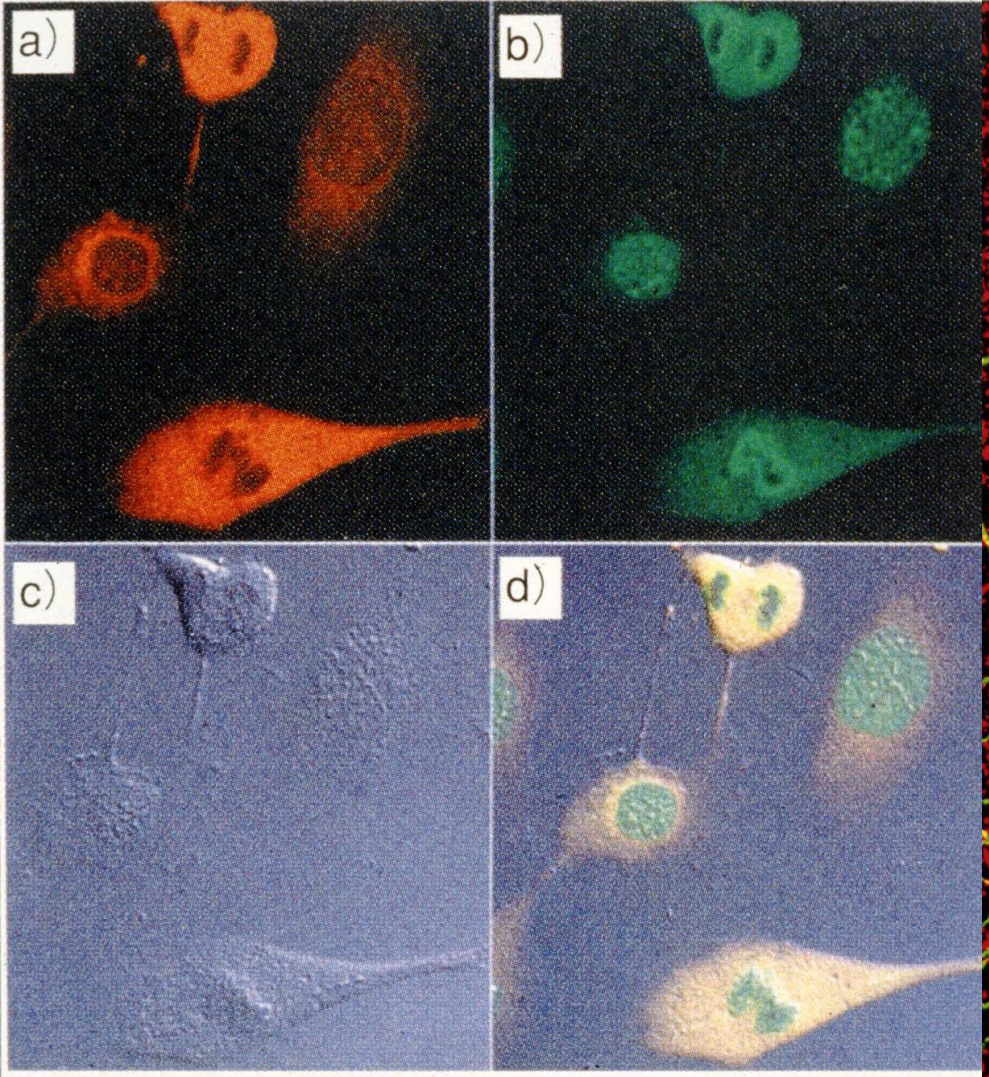
xy scanning, special:
ROI (region of interest)



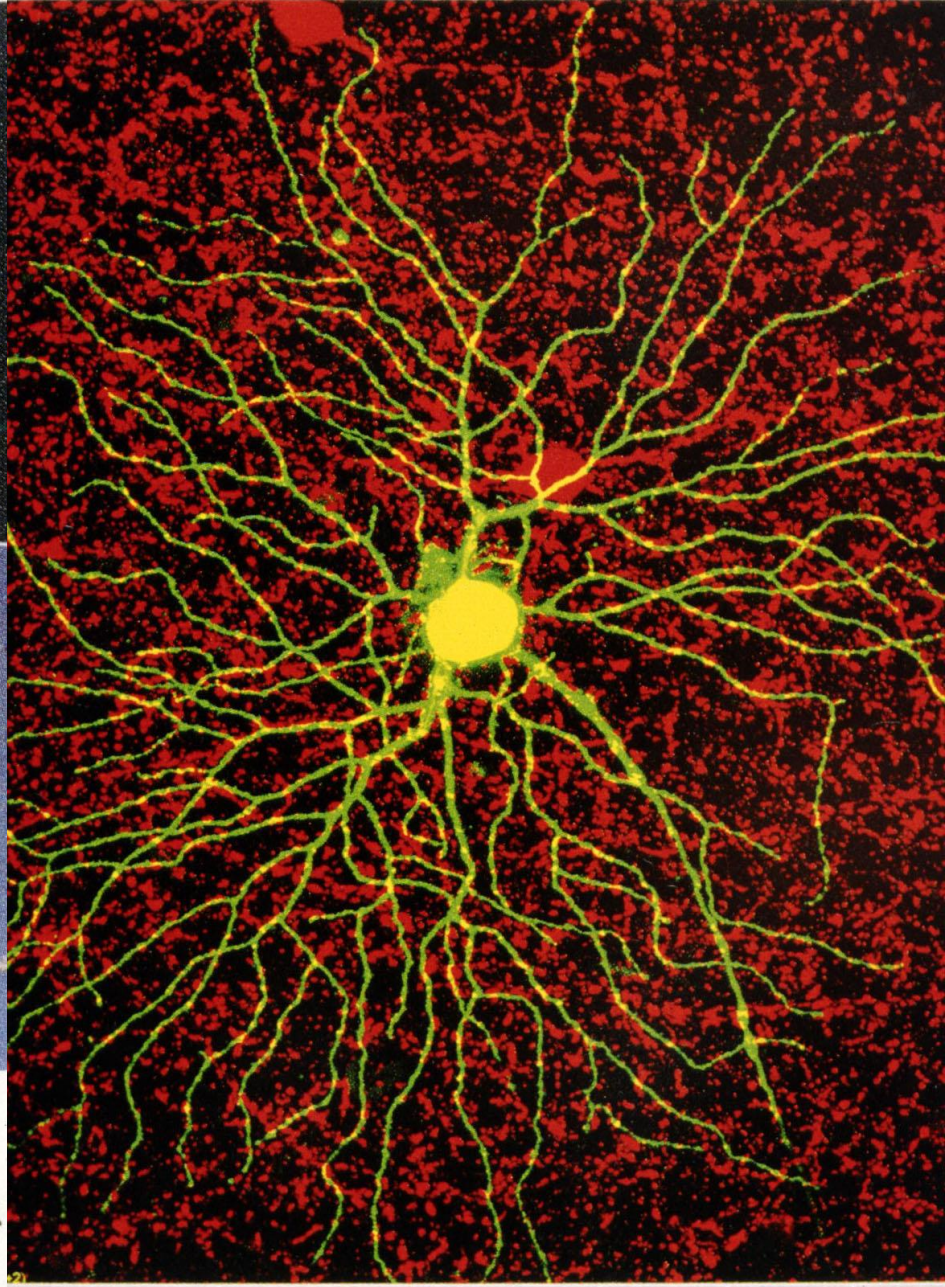


- **Freely configurable laser lines and intensities for ROI´s and surrounding area**
- **FRAP**
- **Uncaging**

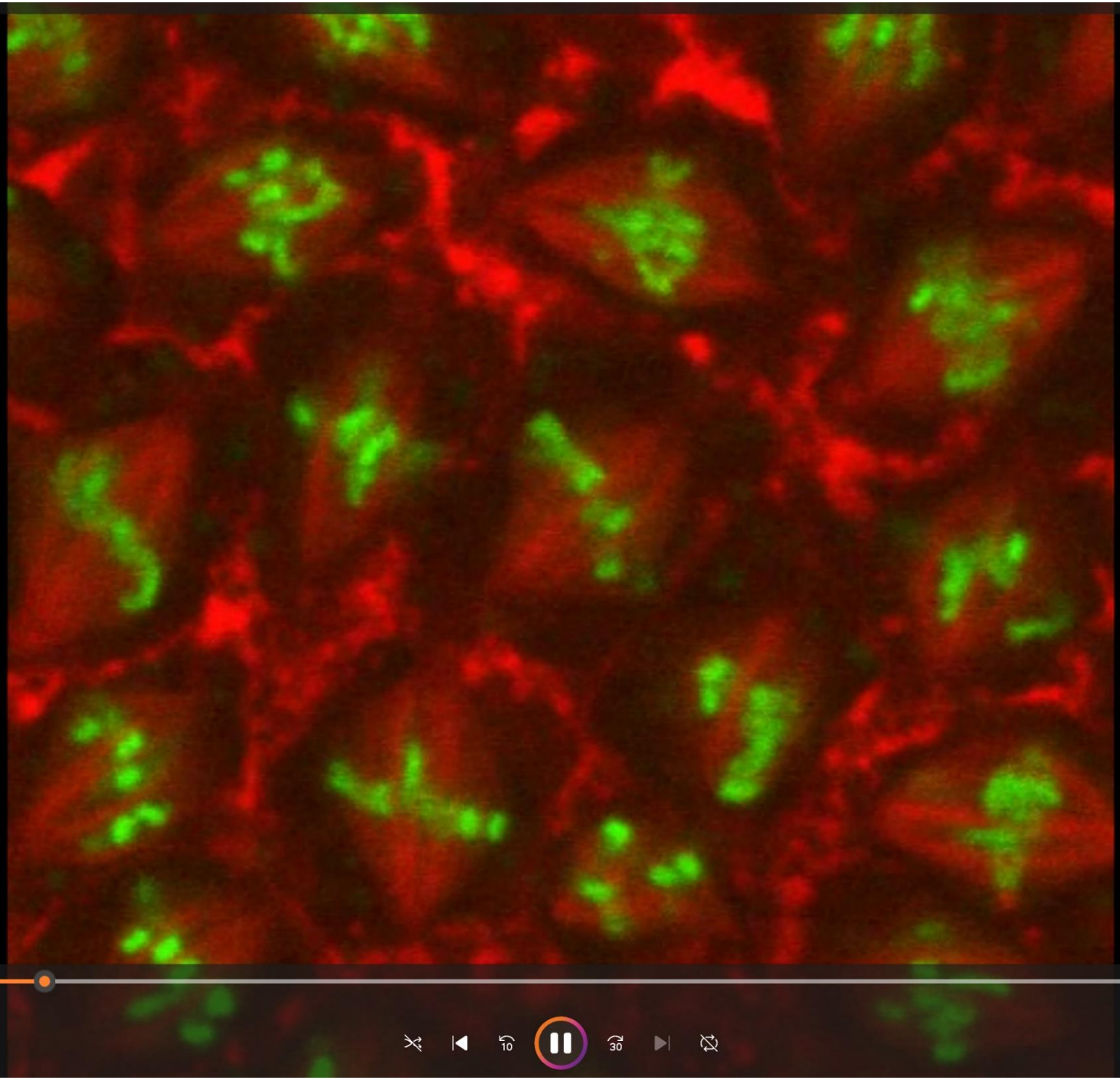
Fibroblasts
ROI 1 543 Cy3 (Intermediate Filaments)
ROI 2 all lines
ROI 3 488 FITC (Microtubules)
ROI 4 UV DAPI (Nucleus)



12 FITC, Rhodamine, 微分干渉および合成像
a) F-アクチン/Rhodamine染色像, b) クリスン/FITC染色像, c) 微分干渉像, d) 合成像.
ラット由来の筋芽細胞 (L-6) の観察像.
東京大学大学院総合文化研究科広域科学専攻生環境科学系跡見順子先生より提供



細胞とドーパミン作動性アマクリン細胞のレーザー顕微鏡エクス
像. Lucifer Yellowで網膜神経節細胞, Texas Redでドーパミン
を染色.



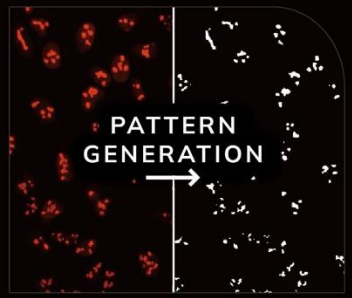
SYNCELL

MICROSCOOP®

PROTEIN-PICKABLE MICROSCOPE

Microscopy-Guided
Subcellular Proteomic Discovery

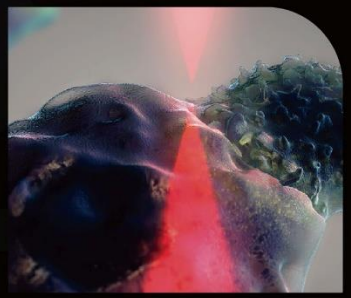
WORKFLOW



STEP 1

REAL-TIME IMAGE ANALYSIS

Photolabeling kit (i.e. Synlight-Pure™ Kit or Synlight-Rich™ Kit) is first added to a cell or tissue sample for a photochemical reaction. After the sample is loaded onto the stage, Microscoop® takes an image (or images of multiple colors) of the sample at one field of view (FOV) at a time. The image or images are analyzed in real time by Microscoop's software Autoscoop™, which executes traditional image processing or AI deep learning to segment the user's region of interest. Pre- or post-processing can be included to enhance segmentation accuracy.



STEP 2

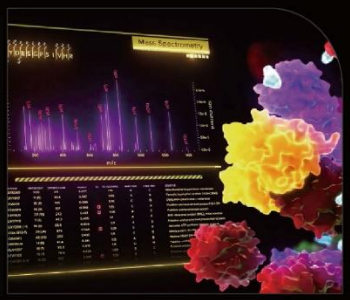
PATTERNED PHOTO-BIOTINYLATION

A femtosecond light source is controlled to illuminate the segmented region of interest one point at a time. This patterned illumination triggers targeted protein photo-biotinlyation in high spatial precision through the reactions of light-sensitive probes of Synlight-Pure Kit or Synlight-Rich Kit. This patterned photolabeling is repeated for thousands of FOVs automatically to assure that enough proteins are biotinlyated for later proteomics analysis using mass spectrometry.

STEP 3

PROTEIN EXTRACTION

Photolabeled cells or tissues are scraped from the slide or chamber. Materials from multiple slides or chambers can be pooled together to increase the total protein contents. The scraped sample is then treated with reagents of protein extraction kit (i.e. Synpull™) Kit to lyse the sample, enrich the proteins by immunoprecipitation, and digest them into peptides for proteomics analysis.



STEP 4

PROTEOMIC IDENTIFICATION

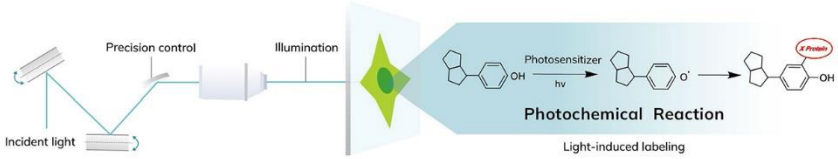
The collected peptides are sent to a mass spectrometer to perform LC-MS/MS analysis. Proteomes of both the photo-labeled and unlabeled (CTL) samples are obtained. By comparing the control and photolabeled proteomes, a location-specific proteome at the region of interest is obtained in high sensitivity, high specificity, and high spatial precision. Validation can be done by colocalization of immunostaining or additional functional assays.

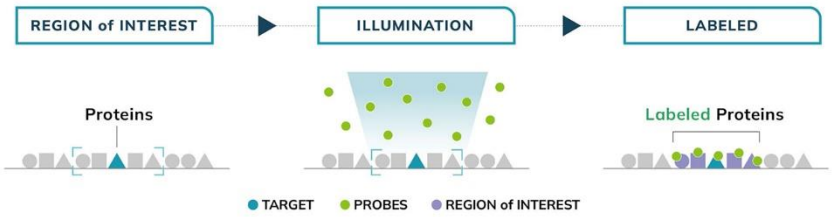
HOW MICROSCOOP® WORKS?

PHOTOCHEMISTRY

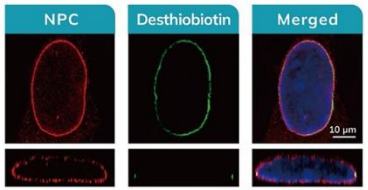
Submicron spatial photo-biotinylation

Photolabeling is achieved by utilizing two-photon illumination to trigger a photochemical reaction with a photocatalyst, which drives redox reactions of molecules that are composed of biotin and a photoactivable amino acid linker to form covalent bonds with, or biotinylate, amino acids within the illuminated focal spot at the submicron labeling resolution. Duration of each illumination spot is in the millisecond or sub-millisecond range to allow fast biotinylation for the entire sample.

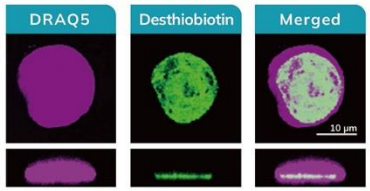




PHOTOLABELING IMAGING

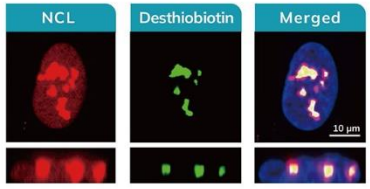


Nuclear pore complex



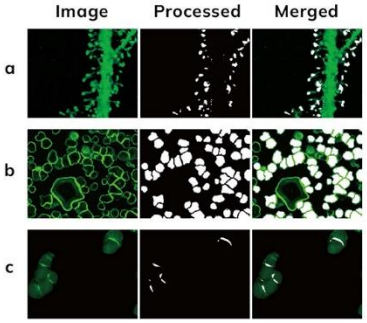
Nucleus

Stress granules



Nucleoli

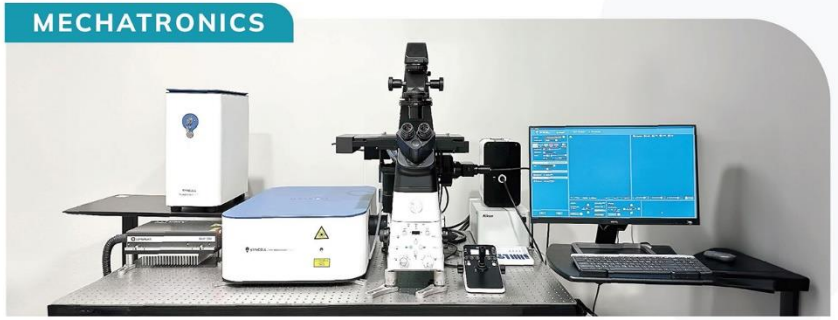
ON-THE-FLY AI



AI-Guided Targeted Photolabeling

When traditional image processing is not precise enough to segment the region of interest, possibly due to the complexity of the images or image quality, one can use deep learning-based image segmentation to achieve proteomic discovery. Hundreds of annotated images are used to train the neural network for a specific biological problem. Microscoop's software Autoscoop™ calls the trained neural network so that the system can recognize the region of interest for each FOV on the fly. It is important to perform traditional image processing (a) or AI (b,c) on the fly to achieve high-speed photolabeling.

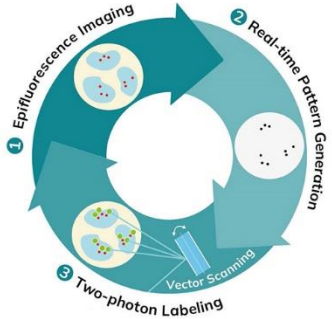
MECHATRONICS



Synchronized Automation

The hardware-firmware-software integrated mechatronic system enables accurate and fast control of scan systems, lasers, microscope, camera, epi-illumination light source, and peripheral devices in real time. The automated process was optimized by synchronizing steps from imaging to intelligent labeling with sub-millisecond temporal precision through this integrated system to allow high-speed, high-resolution spatial photolabeling.

THOUSAND CYCLES OF REPEATS

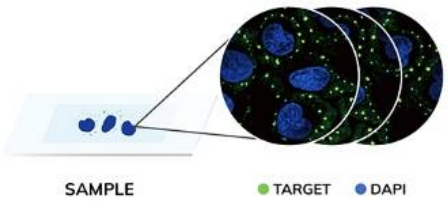


Ultra-Content

Proteins collected from the regions of interest of one FOV are not enough for mass spectrometer's sensitivity to reveal low abundant proteins. To address the protein amplification problem, Microscoop® achieves protein accumulation by performing automated targeted photolabeling at ~10,000 or more FOVs to biotinylate enough proteins for mass spectrometry. The three steps of imaging-pattern generation-photolabeling are repeated for all FOVs. The speed of each step is optimized so that the entire photolabeling process can be finished overnight.

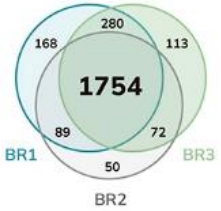
SUBCELLULAR SPATIAL PROTEOMIC DISCOVERY

INPUT

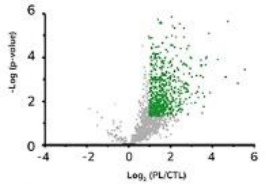


MICROSCOOP®
+
Mass Spectrometry

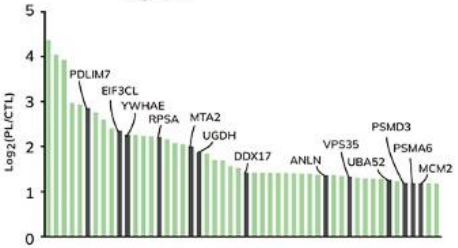
OUTPUT



Venn diagram of the stress granule proteins spatially isolated by Microscoop® and analyzed by mass spectrometry.



Volcano plot of relative protein levels in photolabeled (PL) samples to control (CTL) samples in log_2 scale. Over-represented (enriched) proteins are shown in green.



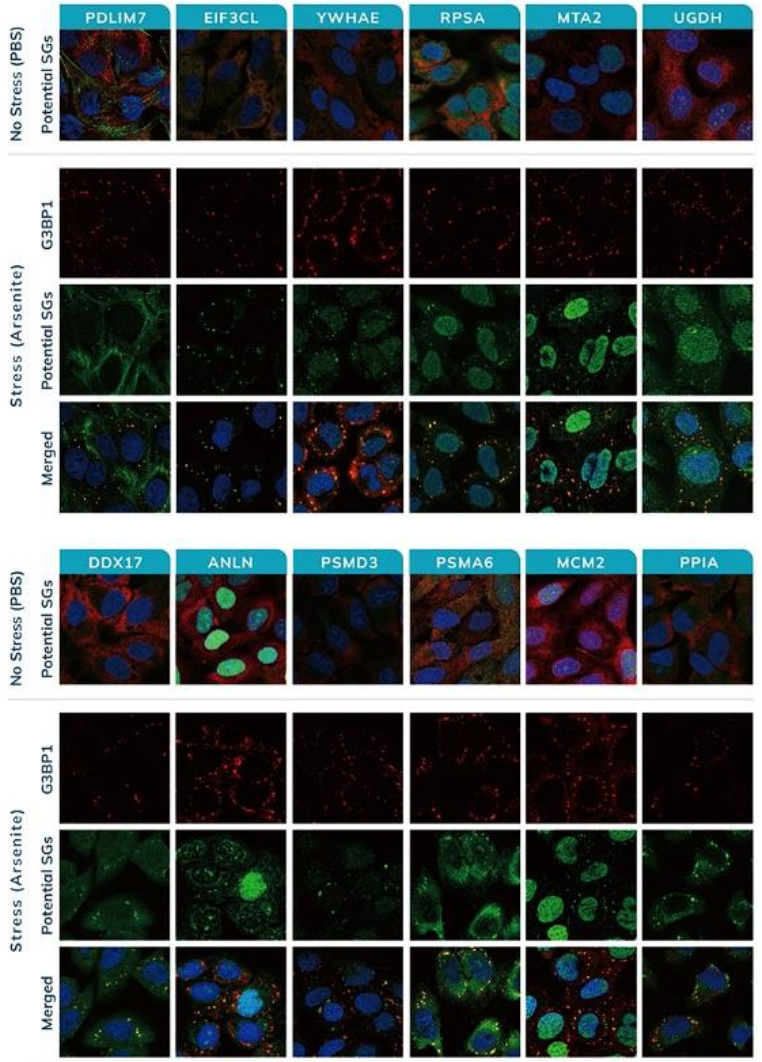
Top 50 spatially enriched proteins by Microscoop® include many known stress granule proteins (green) and others without clear prior annotation as stress granule proteins (gray).

Protein (● Known ● Novel)	Log_2 (PL/CTL)
PALLD	4.37
HNRNPK	4.04
CSDE1	3.92
VCL	2.97
GSPT1	2.92
PDLIM7	2.83
PABPC1	2.74
GARS1	2.60
HNRNPH1	2.39
EIF3CL	2.34
YWHAE	2.25
TARDBP	2.24
FXR1	2.23
EIF3E	2.22
RPSA	2.21
MOV10	2.16
HDLBP	2.07
HNRNPL	2.04
MTA2	1.99
UGDH	1.86
PCBP1	1.83
SYNCRIP	1.69
VCP	1.68
PABPC4	1.56
DPYSL2	1.52
DDX17	1.42
DDX50	1.42
EEF1D	1.42
CPNE3	1.42

A List of Proteins at the Targets

COLOCALIZATION VALIDATION

Proteins without clear prior annotation as stress granule proteins were checked by co-immunostaining with stress granule marker G3BP1 one at a time. The colocalization result shows high specificity of the Microscoop® technology. Novel protein constituents of stress granules were identified in bulk.



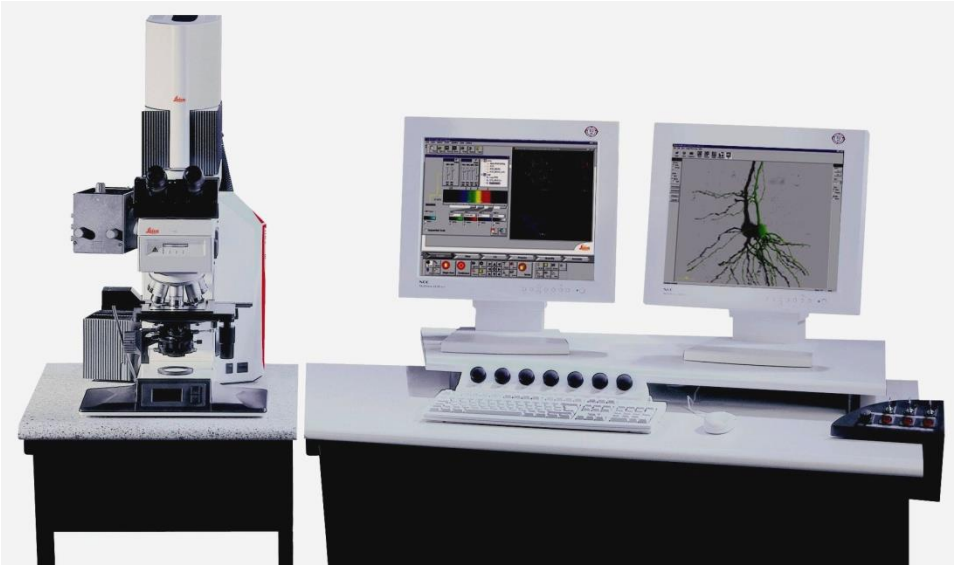
Colocalization validation of novel protein components of stress granules identified by the Microscoop® technology. Confocal micrographs depict stress granule formation in U-2OS cells with or without arsenite stress. Twelve proteins without clear prior annotation as stress granule proteins are highly colocalized with stress granule marker G3BP1. Green: proteins identified by Microscoop®; Red: G3BP1; Blue: DAPI.

Multi-dimensional Live-cell Imaging System

Functions:

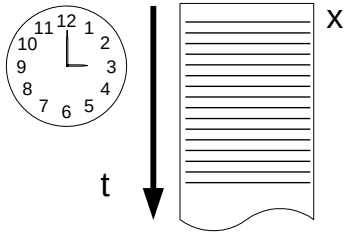
1. Provide non-invasive ways to observe and measure the *in situ* behavior of gene products.

2. Analysis of the dynamics of proteins association/dissociations at cellular structures.

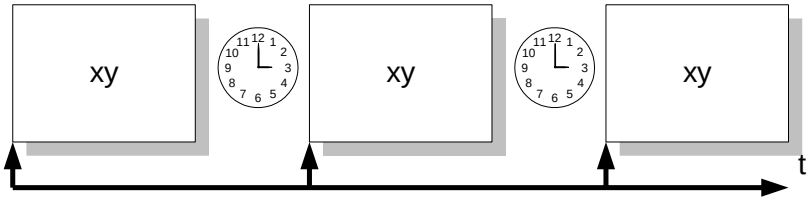


Time-Series

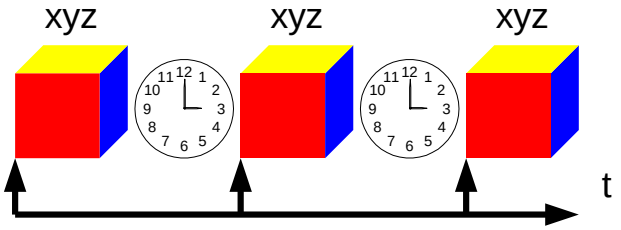
- Line-Mode „xt“



- Frame-Mode „xyt“

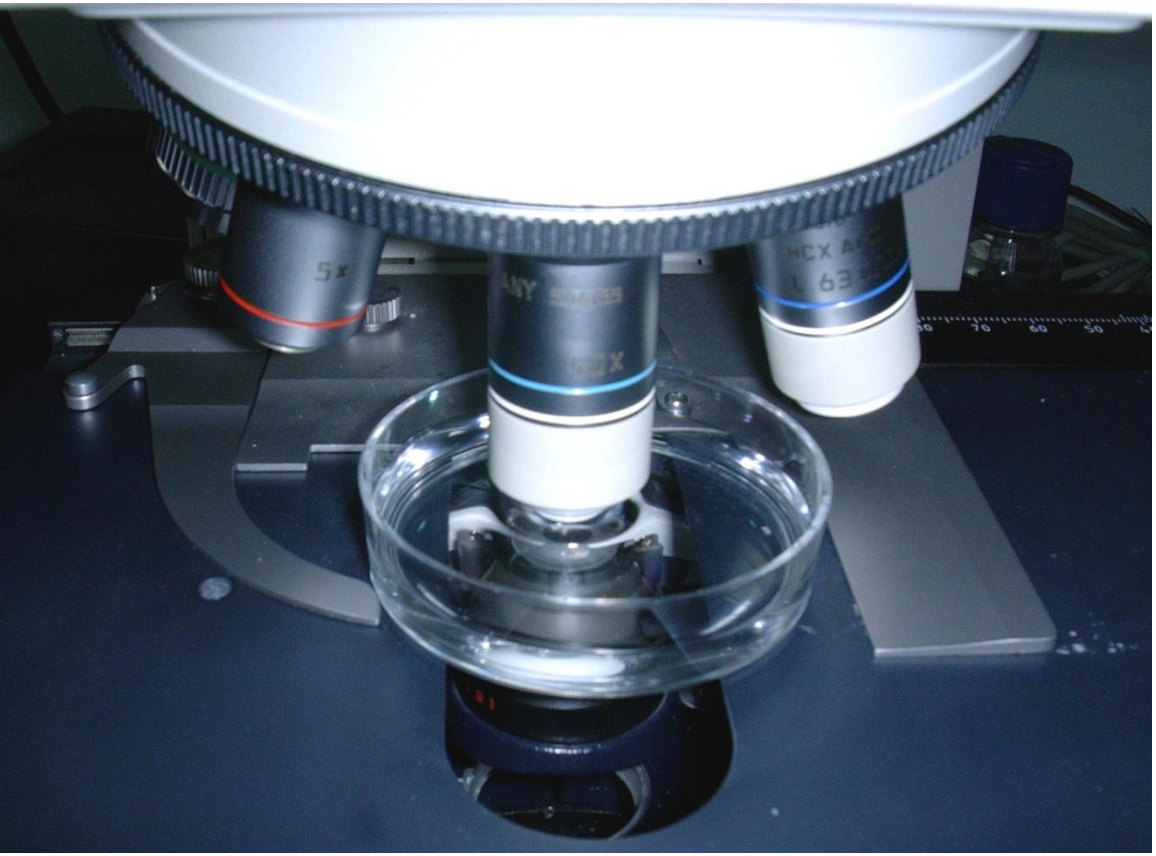


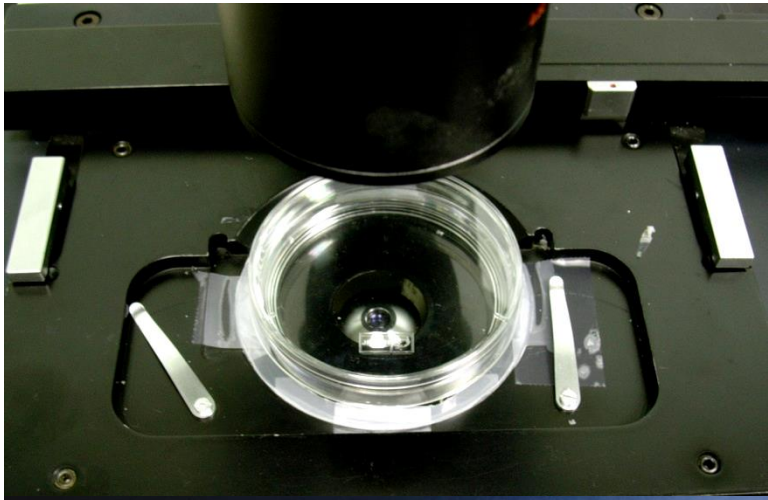
- Stack-Mode „xyzt“



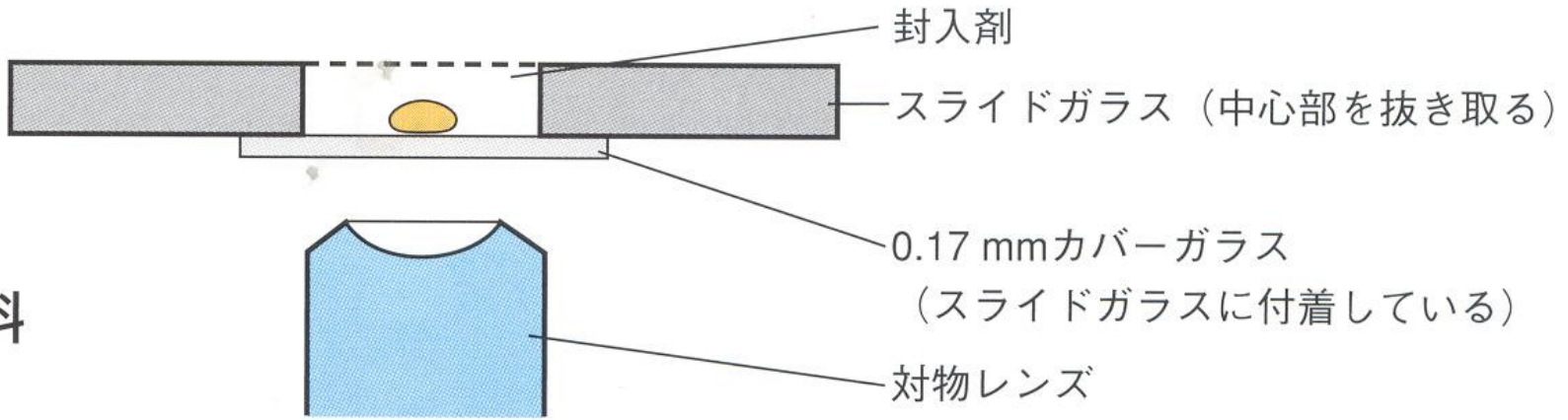
Traditional Live Cell Observation

Up-right microscope with Water Lens or Inverted microscope



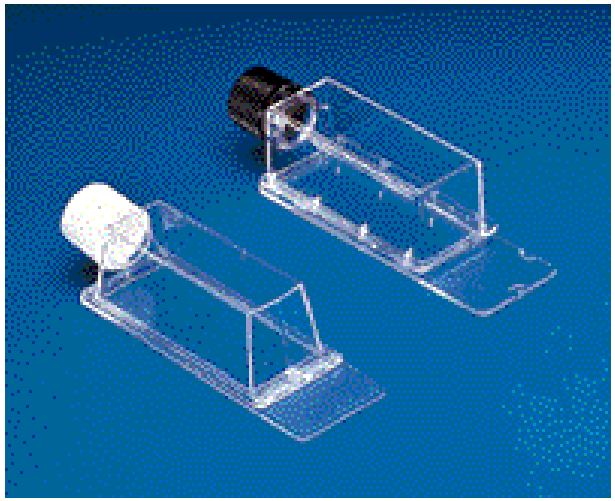


2）倒立型顕微鏡の場合

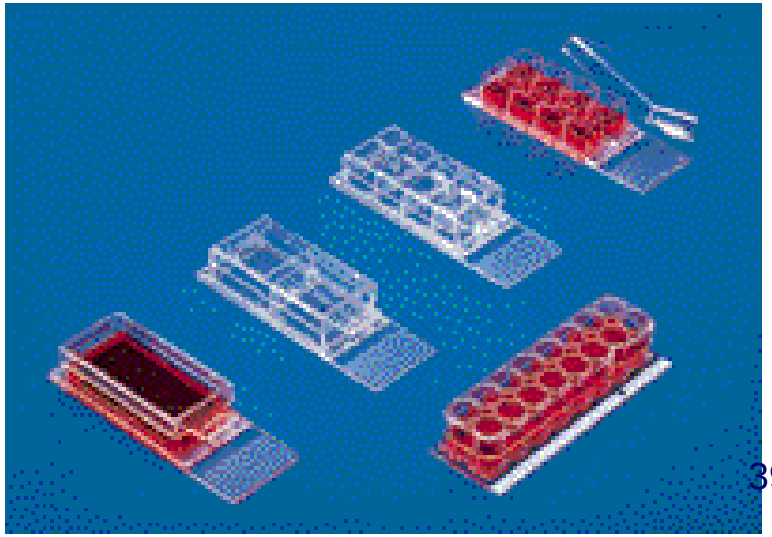


ための試料

NUNC FLASKETTE®
CHAMBER SLIDE/FLASKS



LAB-TEK® II
CHAMBERED COVERGLASS



Computerized Fluorescence Inverted Microscope Leica DM IRE 2 HC

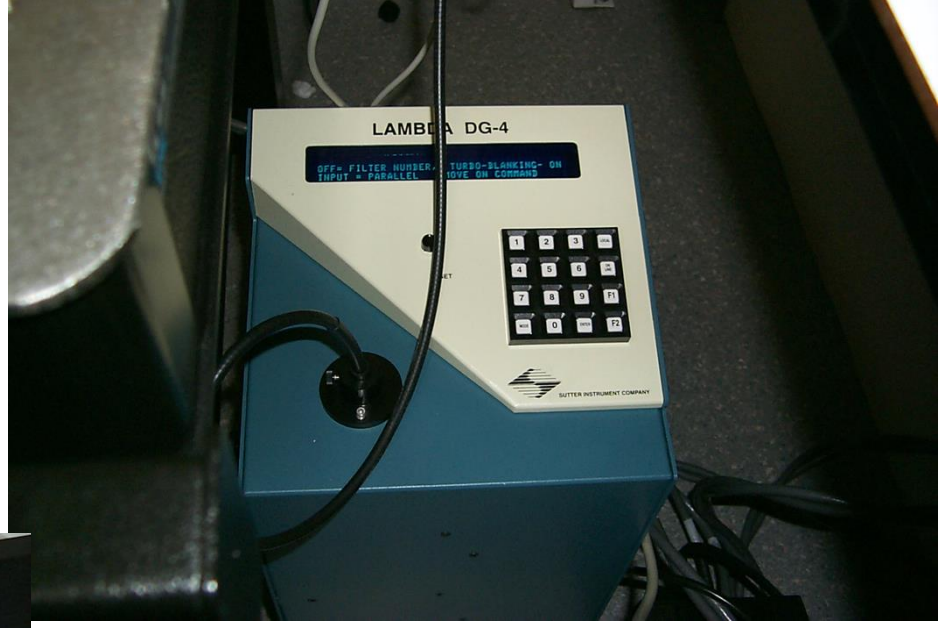
Universal Microscope Controller with Remote Control Knob

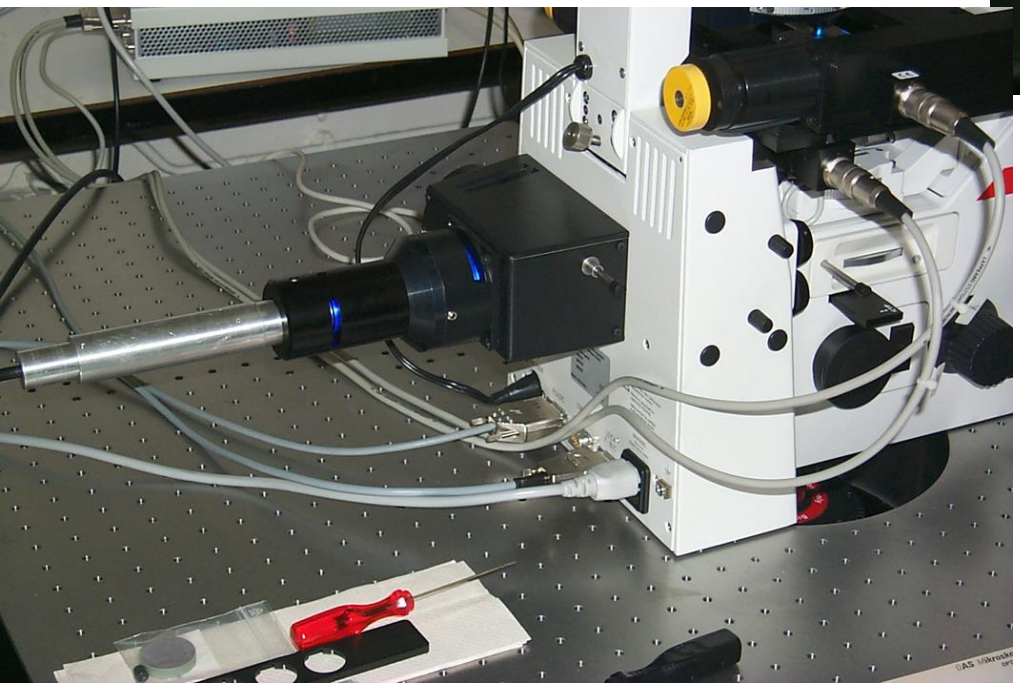
Objectives :
The best axial and laterial resolution
Optimized correction for cell-imaging

HCX PL-APO 10x/0.40 Ph 1
HCX N Plan L20x/0.40 Ph 1, 0-2 mm corr
HCX Pl-Fluotar L40x/0.60 Ph 2, 0-2 mm corr
HCX PL-APO 100x/1.35 OIL Ph 3

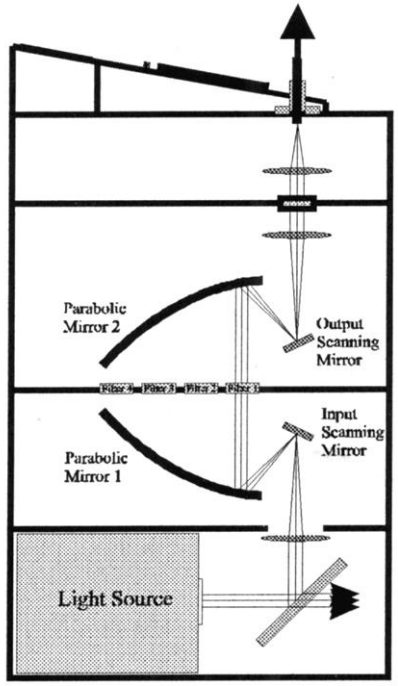
Sutter DG-4 light source:

- Quick wavelength switcher (<2msec)
- Quick shutter and modulator of output energy
- 175 Watt xenon lamp

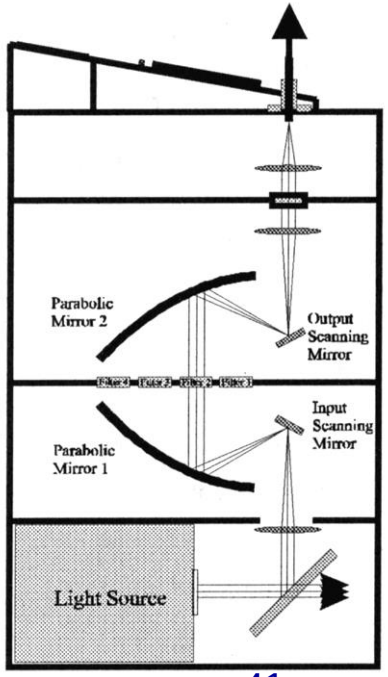




Even and planar illumination: Light source is coupled to the microscope *via* an optical fiber

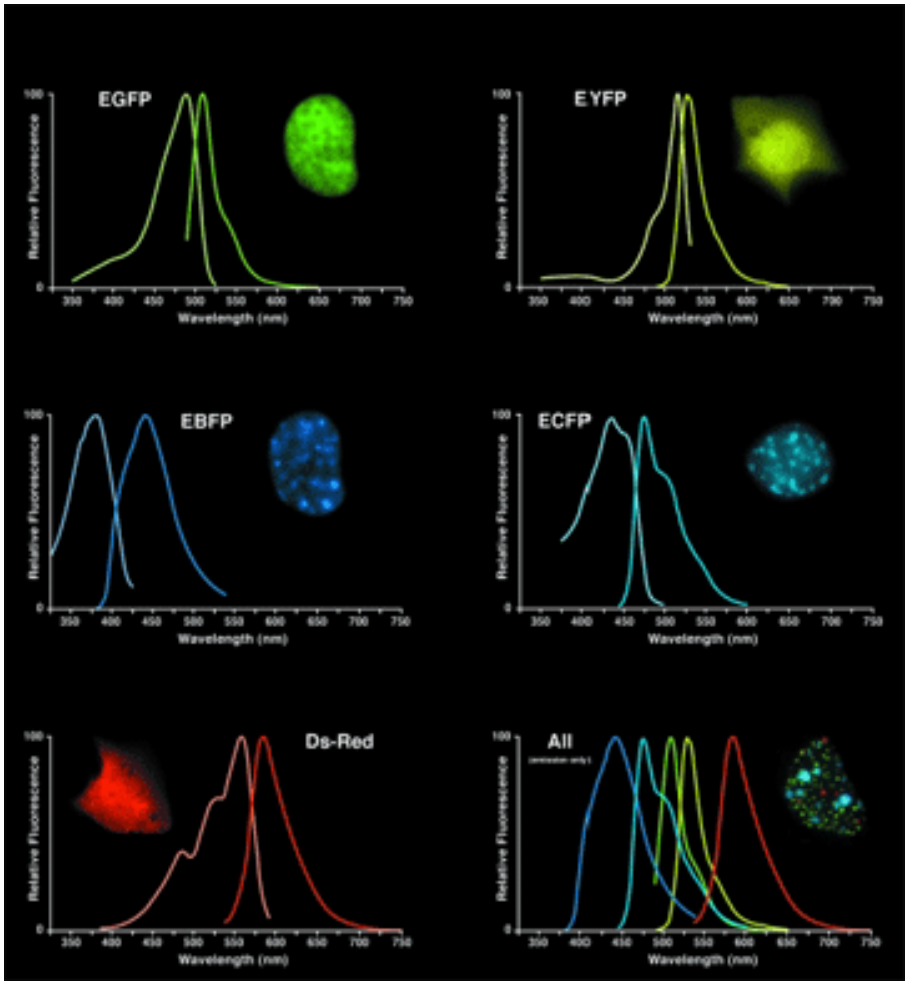


Filter Position 1



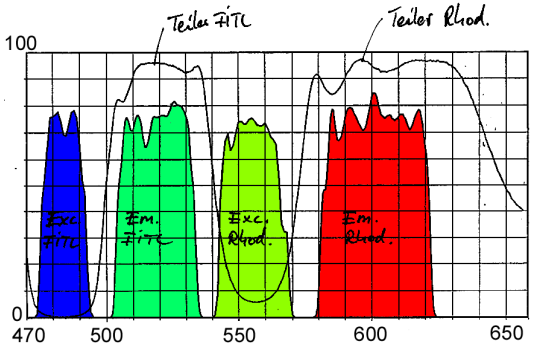
Filter Position 2

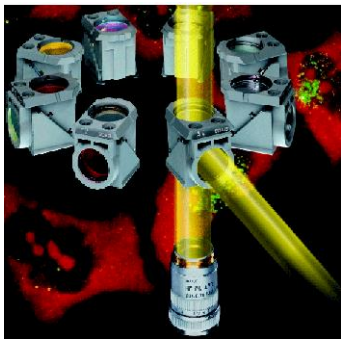
Computerized Fluorescence Inverted Microscope Leica DM IRE 2 HC Fluorescence Filters

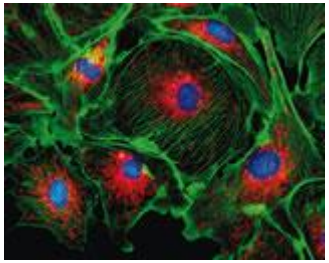


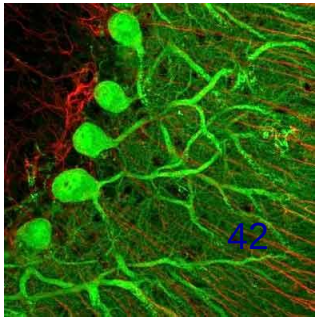
Built-in Four Microscope Filters

- GFP
- CFP
- YFP
- DsRed



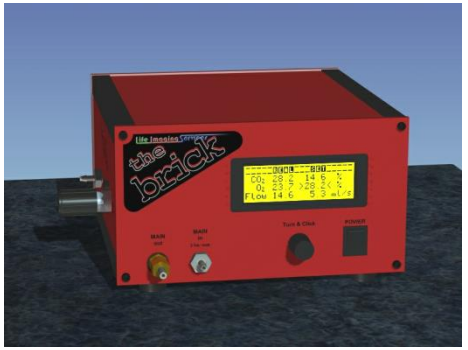






Leica DM IRE2 microscope

enclosed within a computerized CO_2-incubator for indispensable thermal and mechanical stability

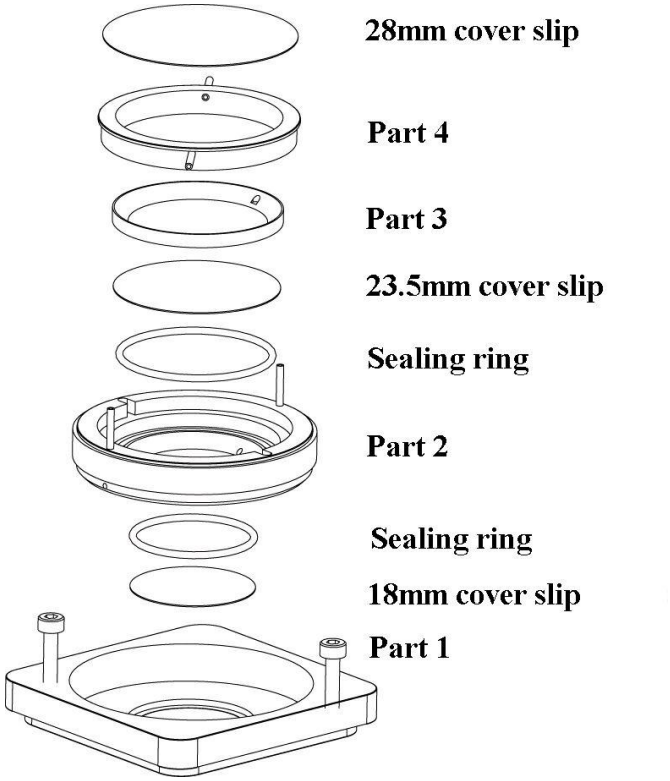


CO_2 controller

Microincubation Imaging-Chamber

Microincubation imaging-chamber: mechanical stability

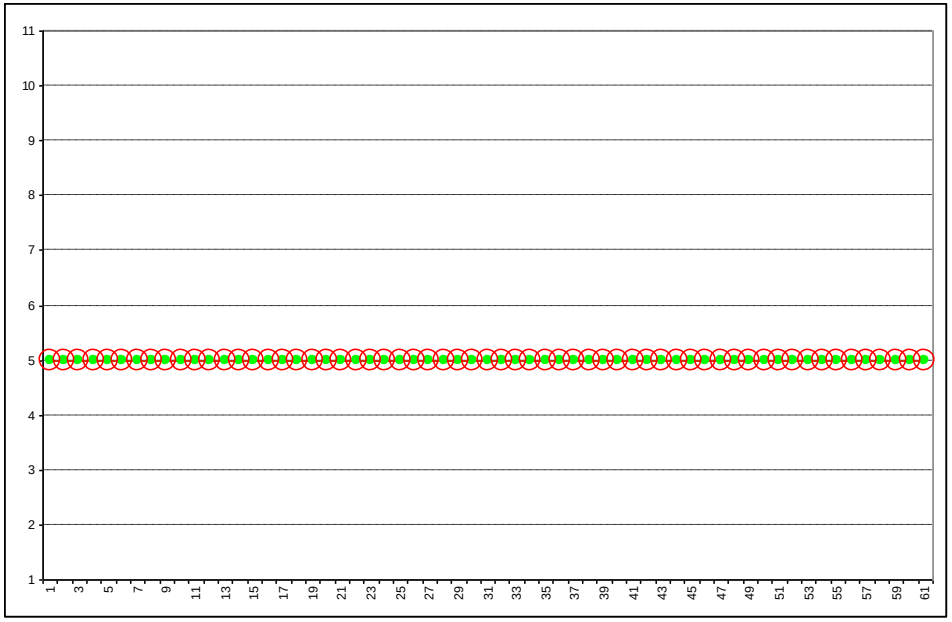
Open / Close / Perfusion

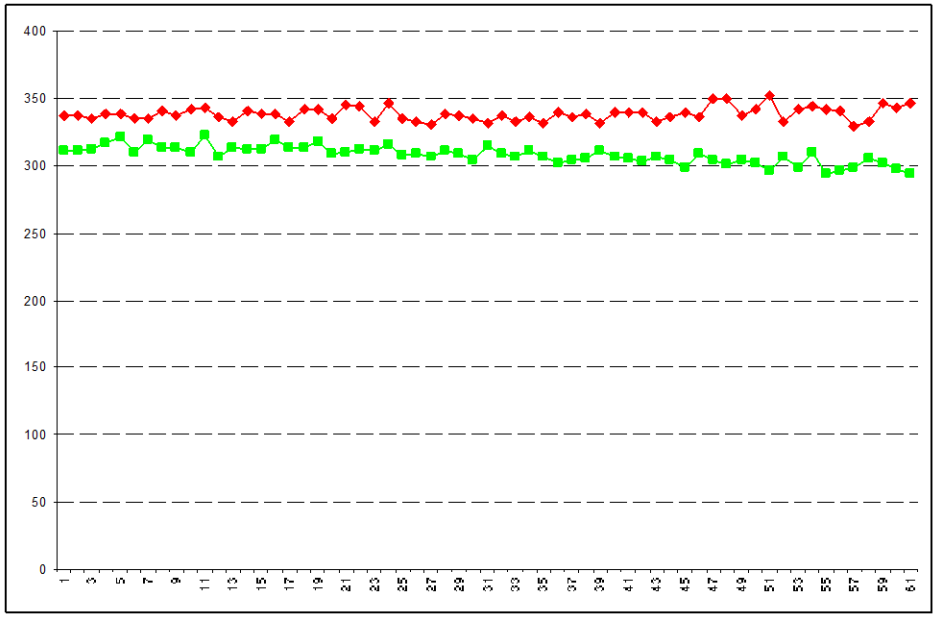


A Stable System on the vibration-free table

Beads Do Not Move during 2-Color 4-D Acquisitions

Measured light intensities at the bead's center are stable

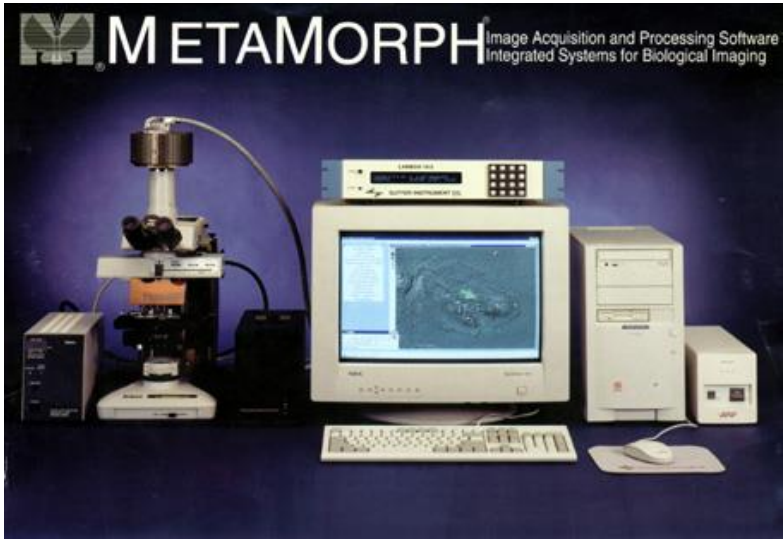


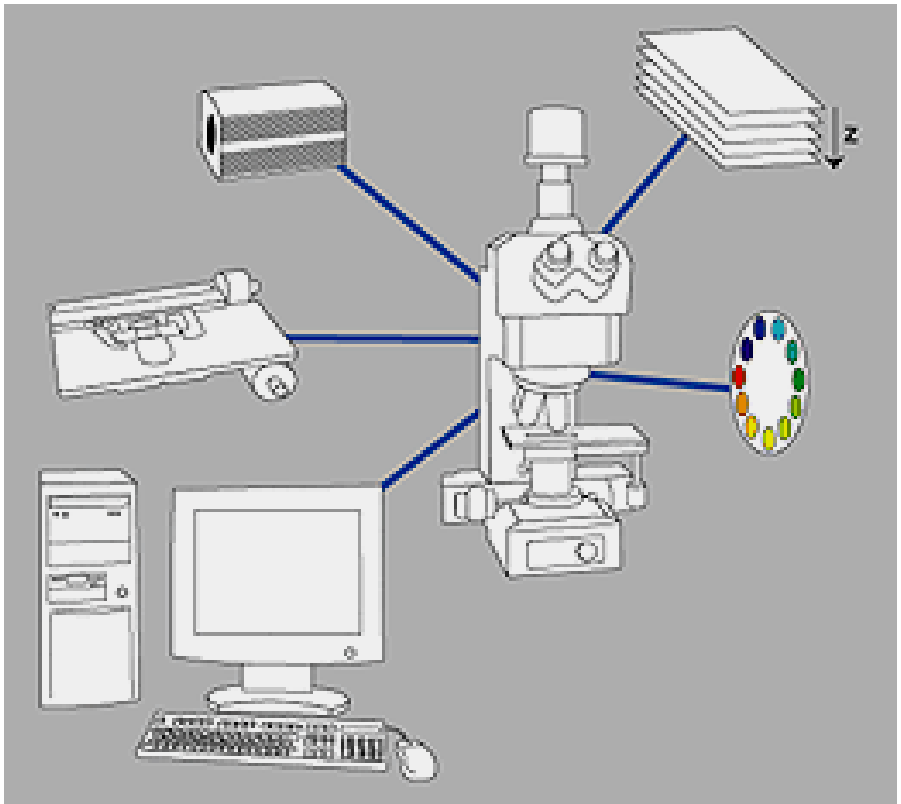


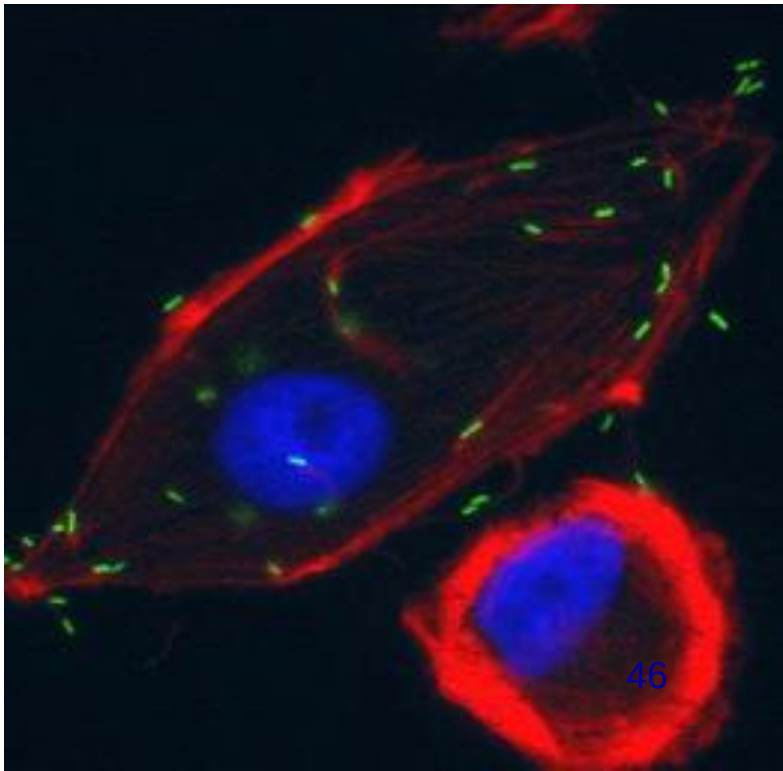
Software: MetaMorph System

integrated imagining system for maximized control

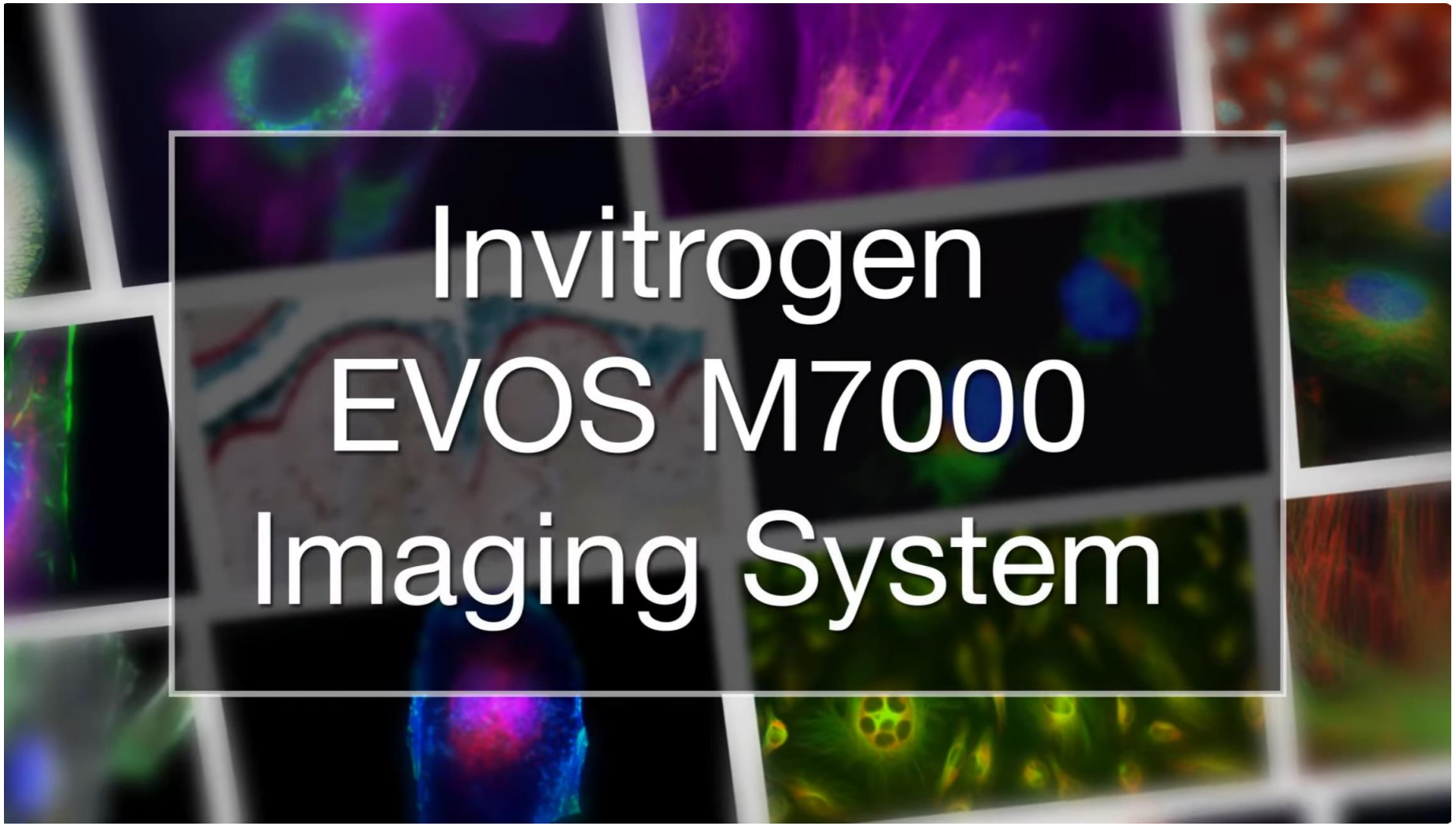
1. Multi-dimensional imaging
2. 3D reconstruction/ deconvolution
3. Time lapse recording
4. Z-series acquisition
5. Morphometry: Cell counting







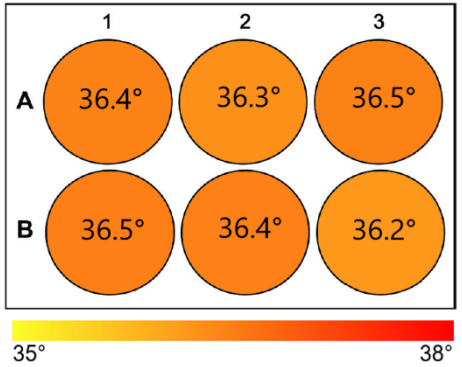
ThermoFisher SCIENTIFIC

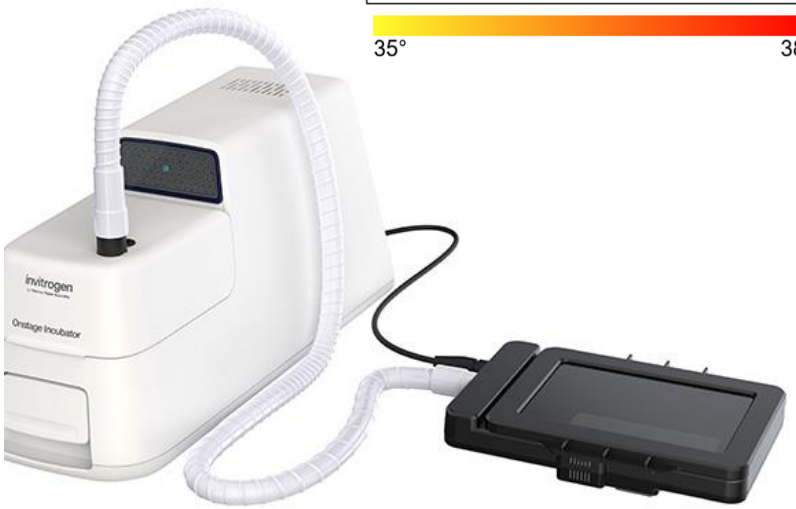


https://www.youtube.com/watch?v=7DBnu3NAinE

EVOS Onstage Incubator (OSI-2) is an accessory for EVOS M5000 and EVOS M7000 Imaging Systems that enables the incubation of cells at user-defined temperature, humidity, and gas (O_2, CO_2 or N_2), for capture of images and recording of time-lapse movies of live-cells under physiological and non-physiological conditions (e.g., hypoxia) over long periods of time.

ThermoFisher SCIENTIFIC





https://www.youtube.com/watch?v=qQe5aRfyVW8

ASTEC

Real-Time Cultured Cell Monitoring System

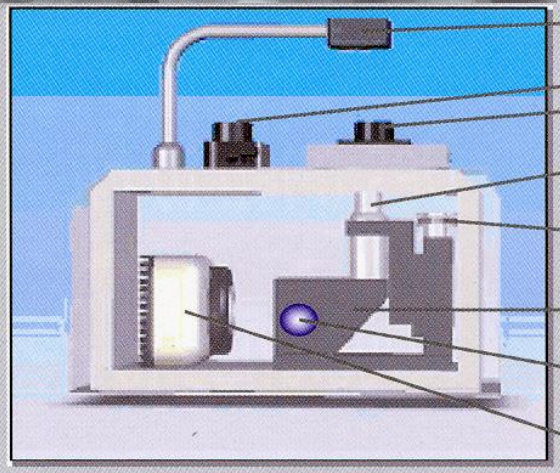
—Supporting the Challenge of Discovery—

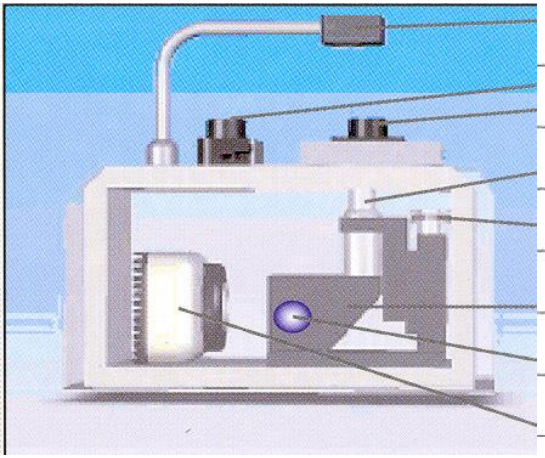
Designed to Fit...

Designed to Resist...

Designed to Discover...

Microscope Now Rests in Incubator!!



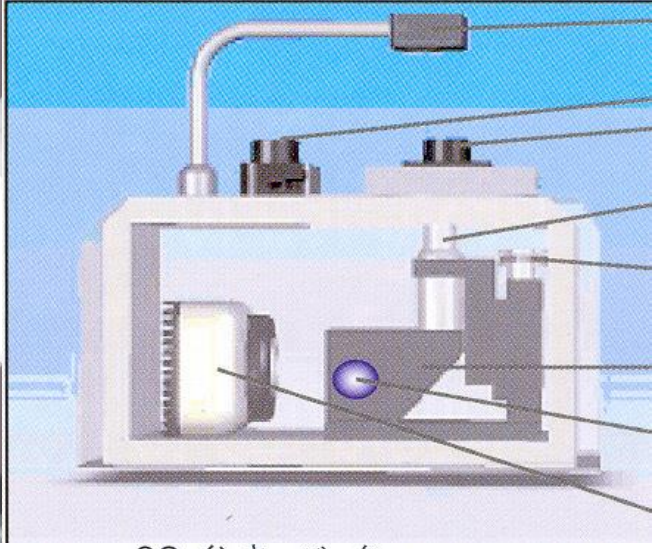


ASTEC
弘優科技代理

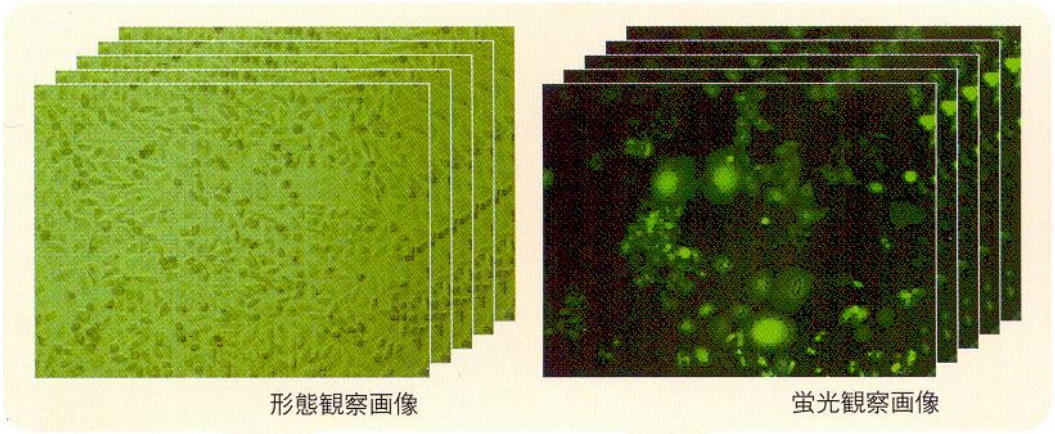
•Real-Time Cultured Cell Monitoring System (MSC Normal light)

	CCM-330F	CCM-500F
Resolutions	3.3 Mega Pixels (2048x1536)	5.0 Mega Pixels (2560x1920)
Camera / Chip Size	Cooled CCD / 1/2 Inch	Cooled CCD / 2/3 Inch
Cooling	Peltier Divice RT-10℃	Peltier Divice RT-10℃
Pixel Size	3.45μm x 3.45μm	3.4μm x 3.4μm
Field of View (Objective X10)	707 x 530 μm	870 x 650 μm
Exposure Time	1.6μs x 17.9min	1.6μs x 17.9min
Capturing Interval	1min - 24h	1min - 24h
Image Format	TIFF / BMP	TIFF / BMP
Objective Lens (Standard)	X 10 / NA0.22	X 10 / NA0.22
Integrated magnification (17" LCD monitor)	X 440	X 360
Light Source (VIS)	White LED	White LED
Light Source (FL)	Blue LED	Blue LED
Excitation Filter	472.5nm Half band width 30nm	472.5nm Half band width 30nm
Fluorescence Filter	520nm Half band width 35nm	520nm Half band width 35nm
Dichroic Mirror	503nm - 730nm	503nm - 730nm
Focus Adjustment	Remote Control from the Controller	Remote Control from the Controller
PC	WindowsXP Professional SP2	WindowsXP Professional SP2
CPU	Intel Pentium4, 3.0GHz 512MB and up	Intel Pentium4, 3.0GHz 512MB and up
Standard Display	SXGA 17" LCD display	SXGA 17" LCD display
Camera Unit Dimensions	W165 x D275 x H165 (8.0kg)	W165 x D275 x H165 (8.0kg)
Controller Dimensions	W220 x D260 x H120 (6.0kg)	W220 x D260 x H120 (6.0kg)

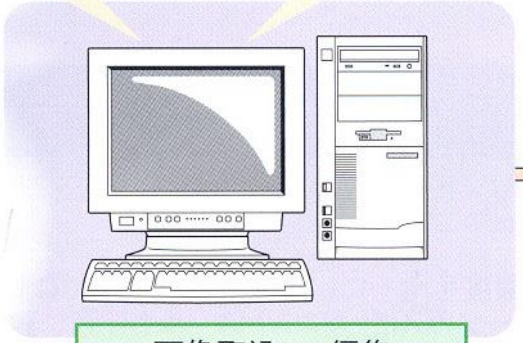

Real-Time
ASTEC
Cultured Cell Monitoring System
—Supporting the Challenge of Discovery—
Designed to Fit...
Designed to Resist...
Designed to Discover...
Microscope Now Rests in Incubator!!



使用装置例


形態観察画像
蛍光観察画像


CO2インキュベーター
(37℃ 95%RH 以上)
カメラユニット
・撮影


・画像取込　・編集


コントロールユニット
・ライトコントロール
・エアーポンプ
・フォーカス

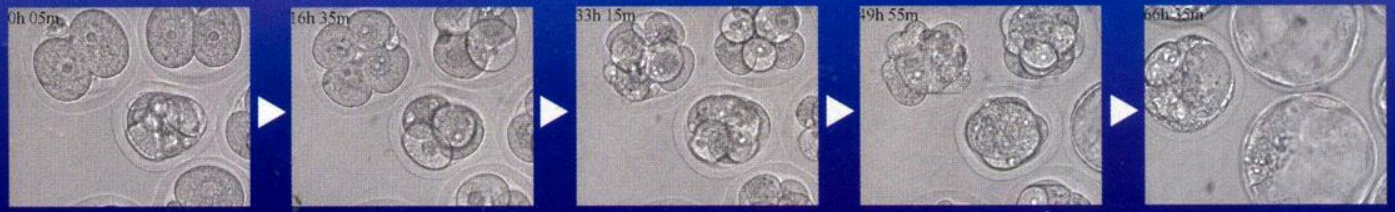
ASTEC CCM-MULTI

■ 機器仕様：インキュベーター部

内　容　量	80L
外 形 寸 法	W735×D510×H760mm
内 形 寸 法	W418×D377×H510mm
棚 板 寸 法	W350×D350×H11mm
加 温 方 式	エアージャケット
温度制御方式	デジタルPID
温 度 範 囲	室温+5℃～50℃
温 度 精 度	±0.3℃
加 湿 方 式	自然蒸発（バット注入）
湿　　度	95±3%RH（5%CO_2時）
CO_2 制御範囲	0～20%
CO_2 精　度	±0.1%
O_2 制御範囲	2～18%（オプション）
O_2 精　度	±0.5%（オプション）
製 品 質 量	78kg
電　　源	AC100VMax7A 50/60Hz（インキュベーター専用）
電　　源	AC100VMax5A 50/60Hz（カメラユニット関連用）

カメラユニット
内部の温度を
管理します。

ラット受精卵（対物20×　5分間隔で撮影）



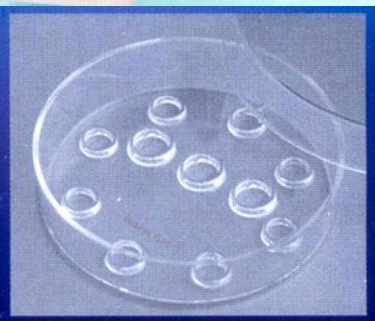
2細胞期から4細胞期。受精卵の中には、極体も確認できる。

殆どの受精卵が4細胞期に移った。

8細胞期。

細胞同士の接着性が変化し、コンパクションが発生する。

胚盤胞となり、次第に透明帯を破るハッチングが確認されるようになる。

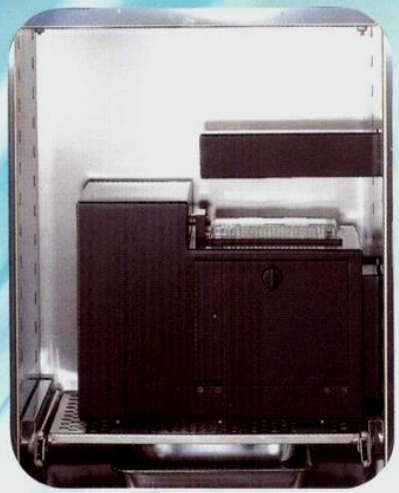


IVFディッシュG-2Sにて
受精卵を観察撮影

ASTEC CCM-MULTI

■ 機器仕様：カメラユニット部

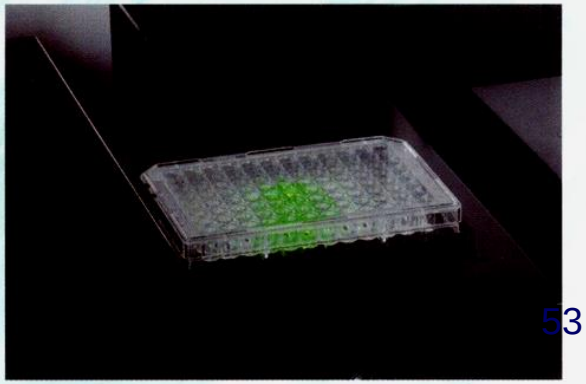
画素数	140万画素（1392×1040）
イメージセンサー	モノクロ冷却CCDカメラ
対応可能対物レンズ	4×（NA0.2）、10×（NA0.22）、20×（NA0.45）
撮影範囲	640μm×480μm（対物10×）
冷却温度	周囲温度-25℃（ペルチェ素子）
タイムラプス時間設定	1min～24hr
形態観察光源	緑色LED
形態観察方式	透過照明（偏斜照明）
蛍光観察光源	青色LED（470nm peak）
蛍光観察方式	同軸落射照明
励起フィルター	透過ピーク:472.5nm（半値幅30nm）
ダイクロイックミラー	透過幅:503nm～730nm
蛍光フィルター	透過ピーク:520nm（半値幅35nm）



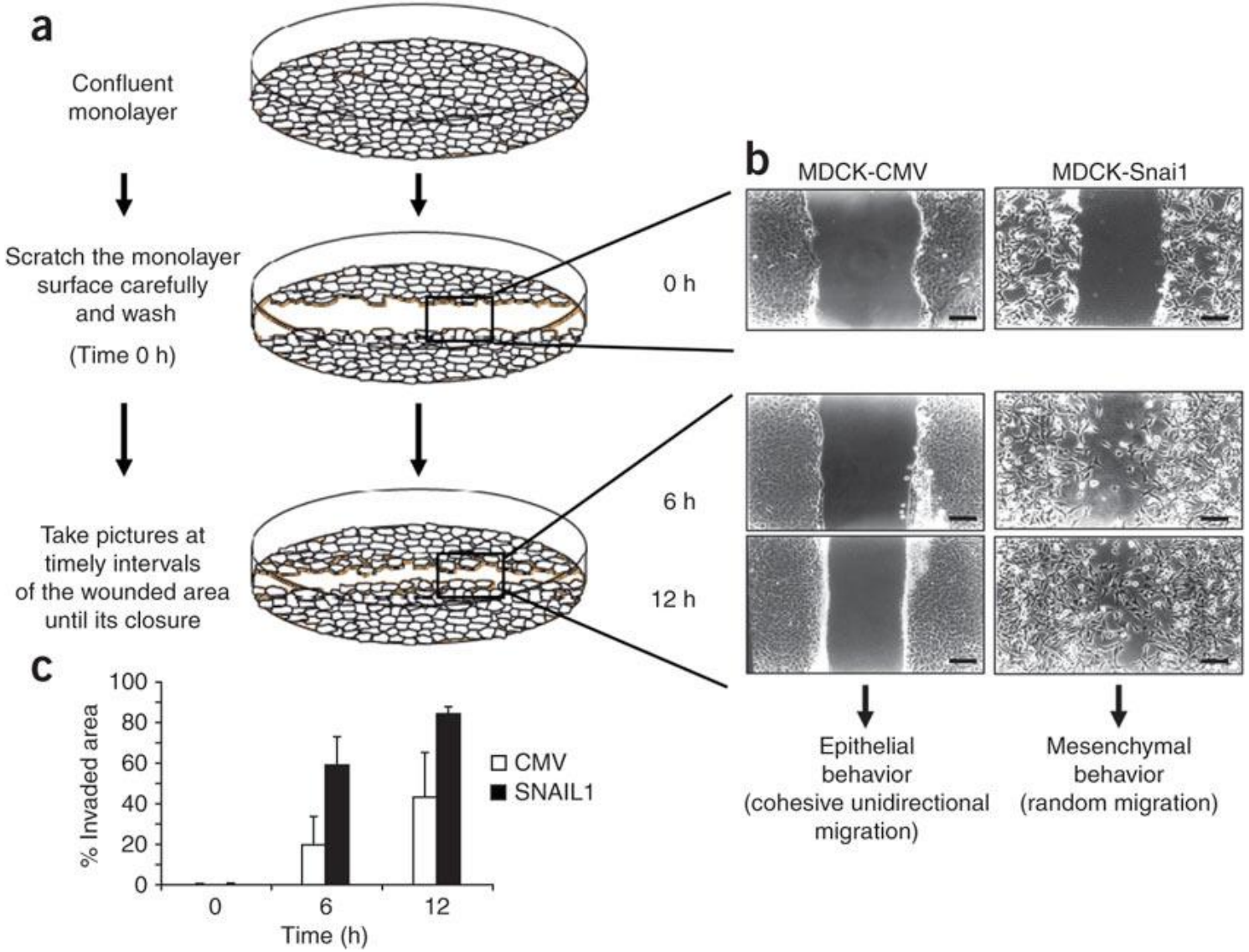
■ 機器仕様：メカニカルステージ部

駆動制御	超高精度ステッピングモータ
分解能（X方向）	0.05μm（ステージ動作）
分解能（Y方向）	0.05μm（ステージ動作）
分解能（Z方向）	0.5μm（対物レンズ動作）
繰り返し誤差	XY方向10μm以内
稼動範囲	100×65mm

※http://www.astec-bio.com　弊社WEBにて
サンプルムービーをご覧になれます。



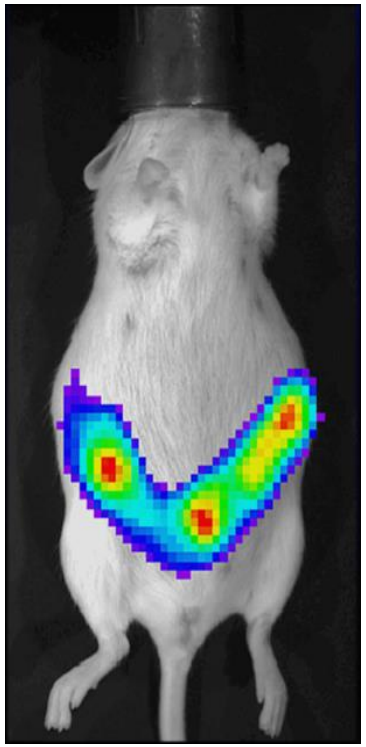
Cell migration assay / Wound healing assay



The morphological and molecular features of the epithelial-to-mesenchymal transition
Moreno-Bueno et al., *Nature Protocols* **4**, 1591 - 1613 (2009)

IVIS™

Biology and User-Driven Technology and Instrumentation Development



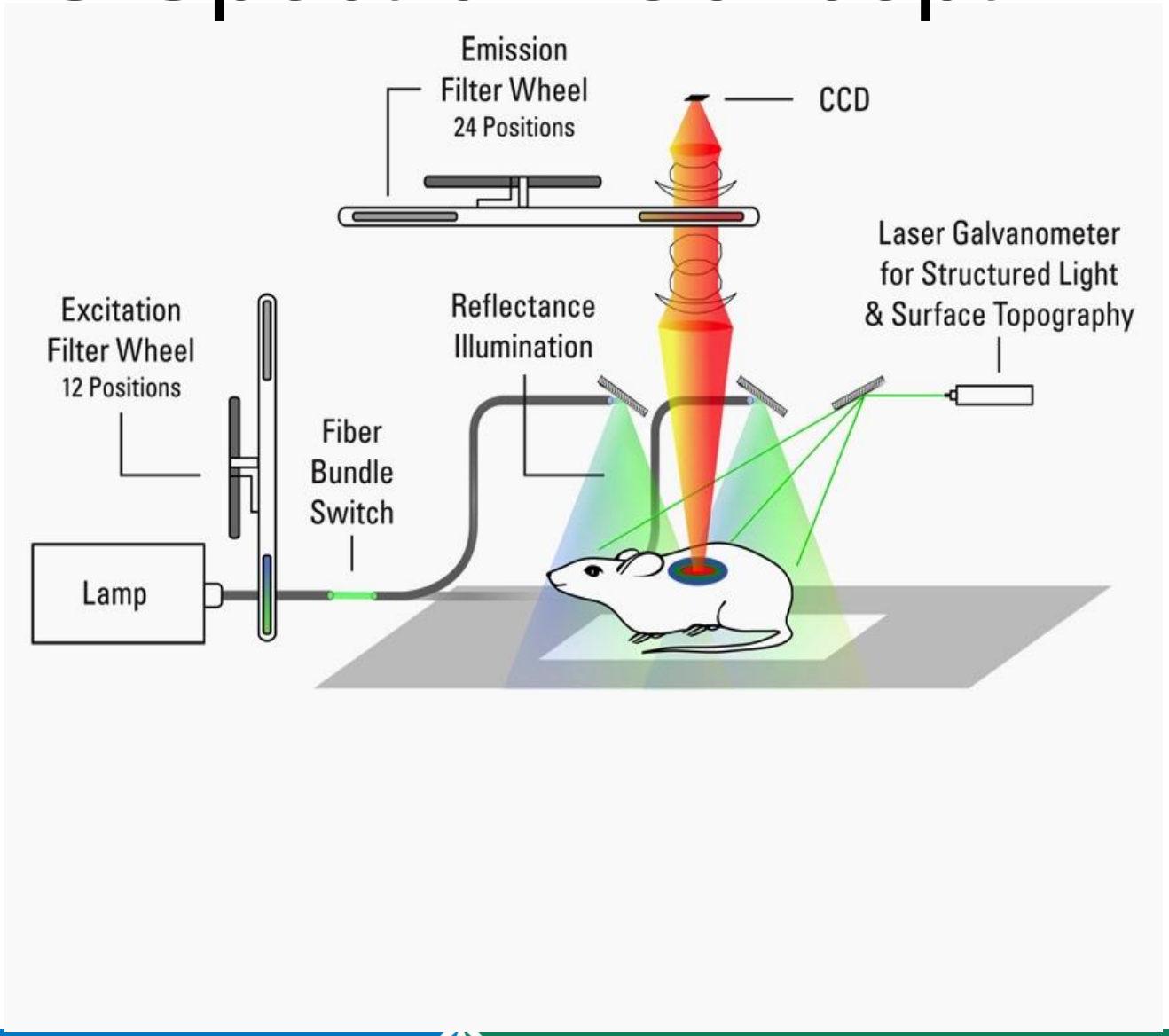
Biology

+

Instrumentation

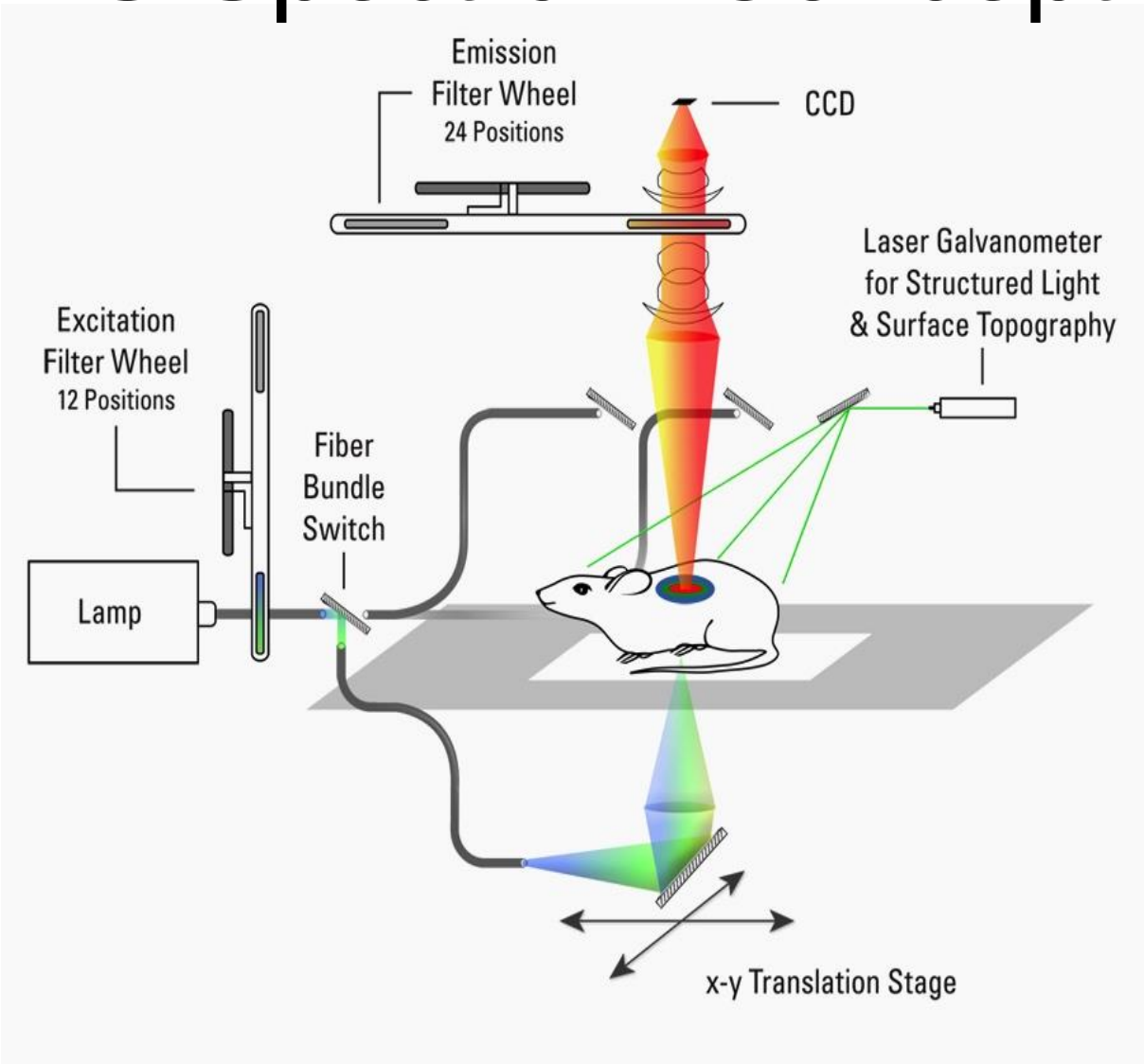
IVIS Spectrum Concept

Reflection-Mode Illumination



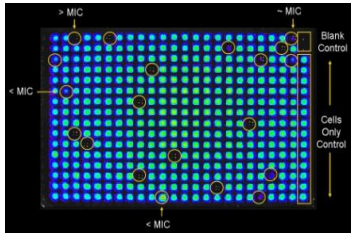
IVIS Spectrum Concept

Transmission-Mode Illumination

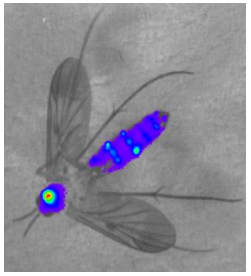


IVIS Spectrum Imaging: Sensitive, quantitative, multi-modal

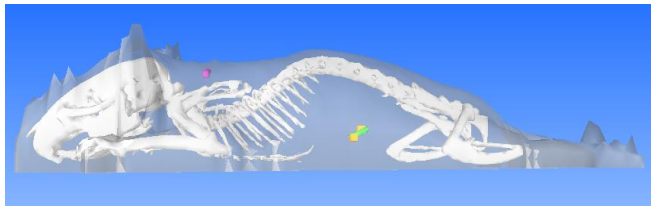
In Vitro



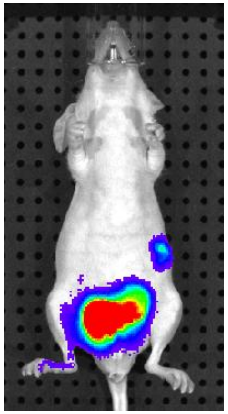
High Resolution



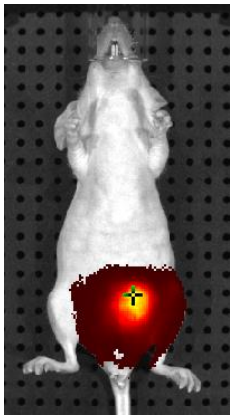
3D tomographic quantification, CT co-registration



Multi-modal

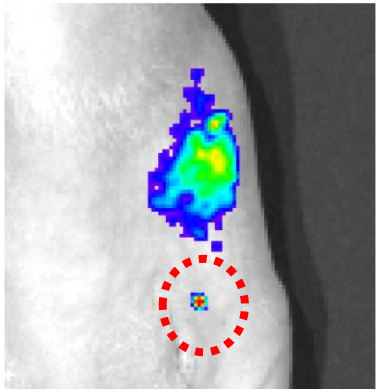


Bioluminescence PC3M-luc

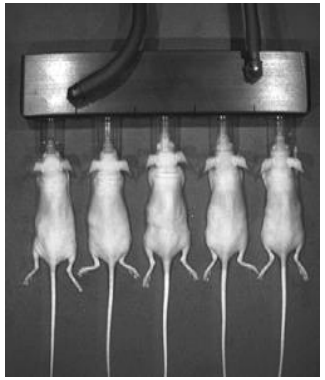


Fluorescent Conjugate – Herceptin®

Fluorescent protein – GFP



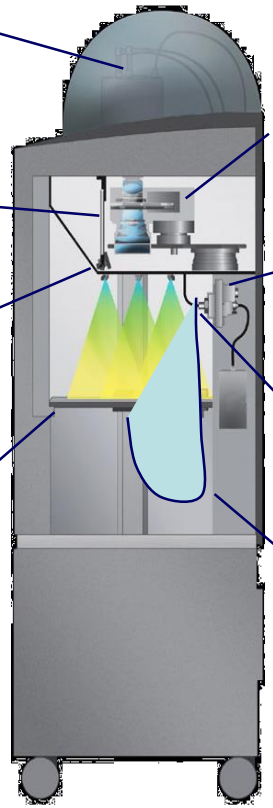
Single cell sensitivity in-vivo
4T1-*luc2*-1A4



High throughput

IVIS Spectrum

CCD, TE-cooled to -90C



Lens assembly

Emission filter wheel

Excitation filter wheel

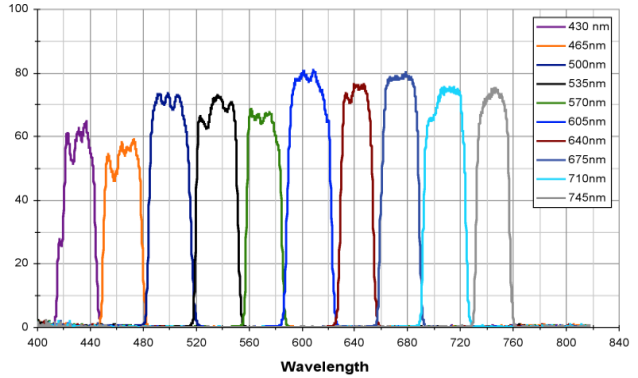
Scanning laser

Optical switch

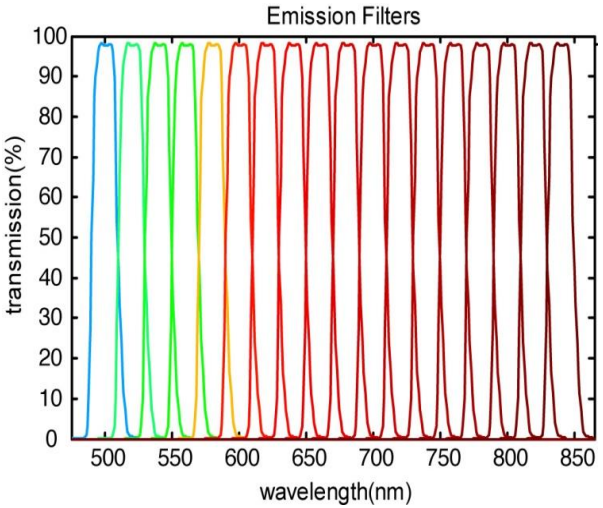
Heated sample stage

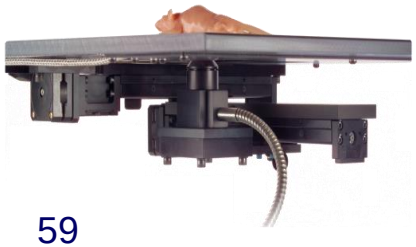
Transillumination Fiber bundle

10 excitation filters (35 nm bandwidth)

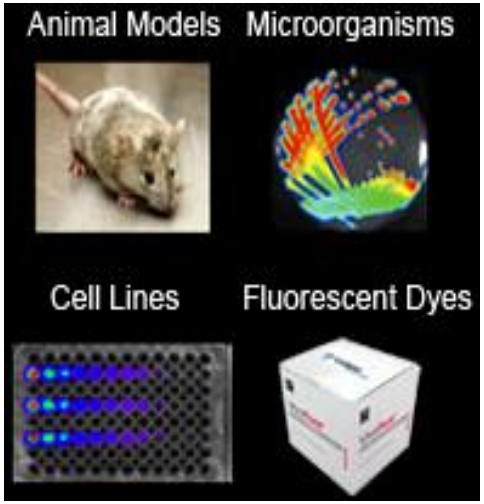


18 emission filters (20 nm bandwidth)





Basic Methodology



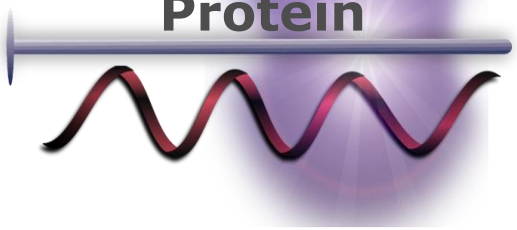
Biological Reporters

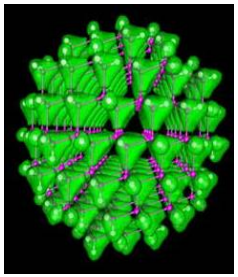
Imaging Hardware

Imaging Software

Reporter Molecules

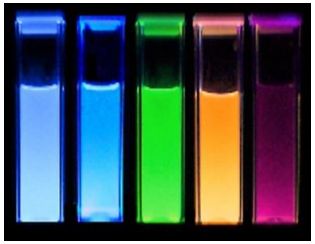
Luciferase, Fluorescent Protein





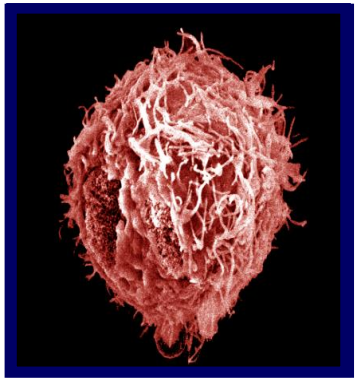
Quantum dots

Fluorescent dyes

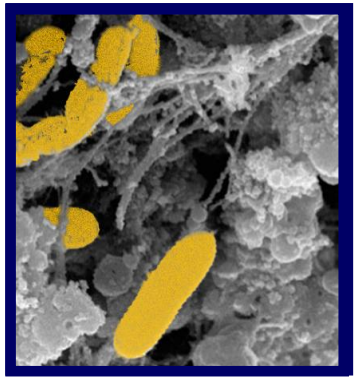


ATP and O_2 required for luciferase

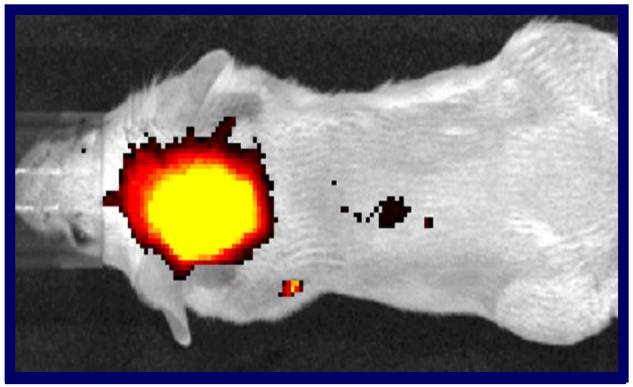
Label Cells



Label Bacteria



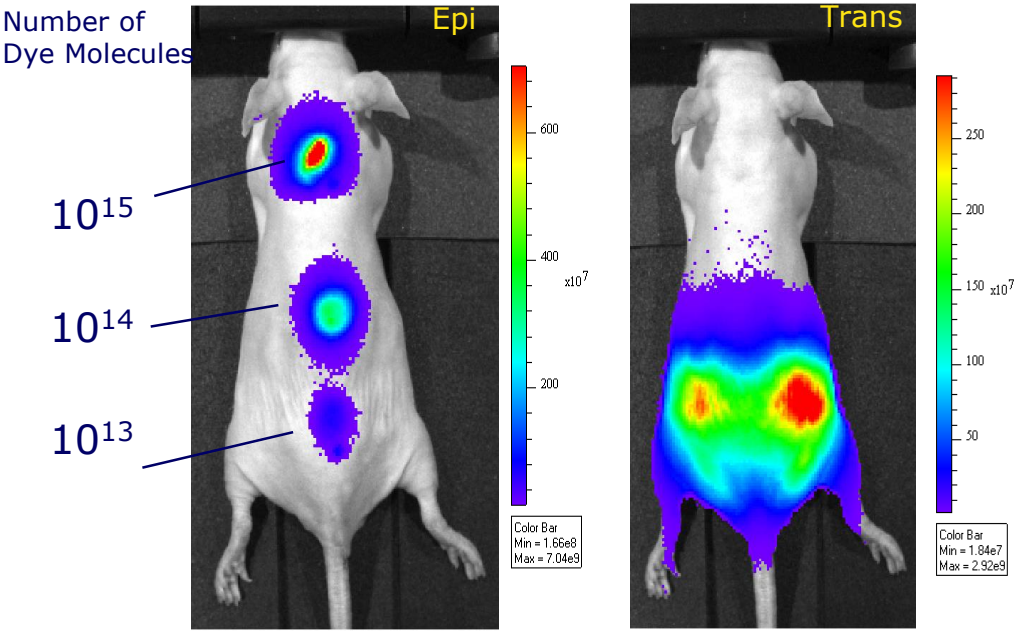
Label Genes



Comparison of Epi and Trans Illumination

Epi-Illumination Reveals Shallow Signals Better Than Trans-Illumination, But Offers Limited Sensitivity For Deep Tissue Fluorescence Imaging

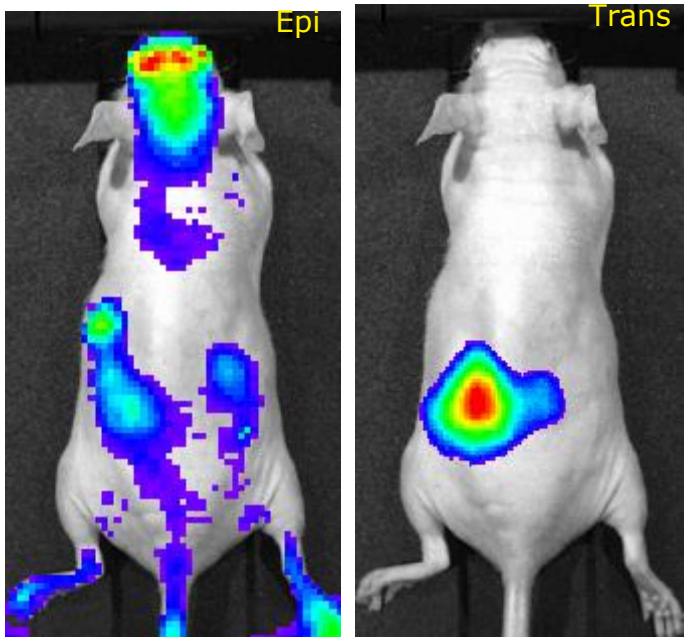
Surface (shallow depth) signal



Different Concentrations of Alexafluor 680 dye
molecules injected subcutaneously

Ex: 640 nm / Em:700 nm

Deep Tissue signal



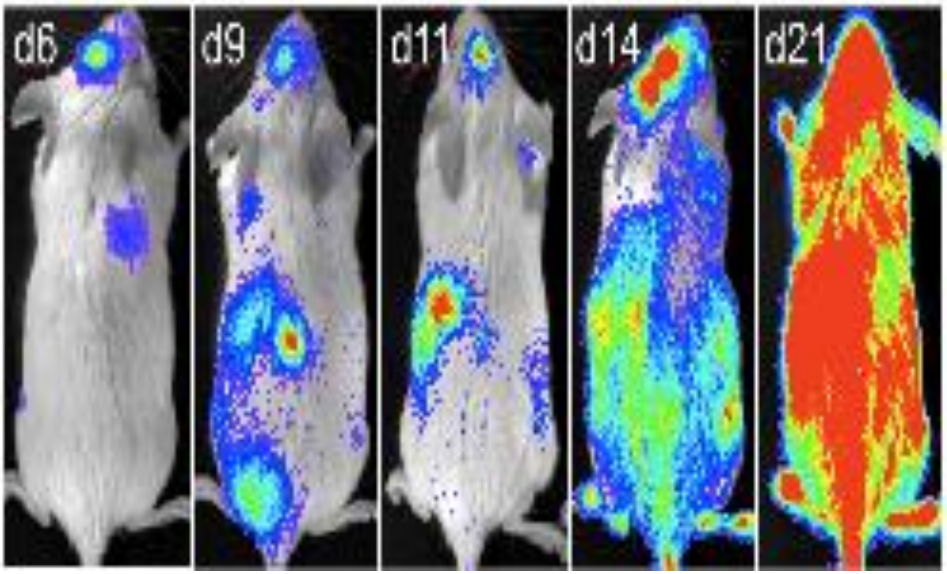
Signal/ bkg=1.10 Signal/ bkg=110

Pillow Containing 1x1015 molecules of Alexafluor 680 Dye implanted medial to left kidney

Ex: 620 nm / Em:700 nm

Cell Transplantation and Trafficking Patterns

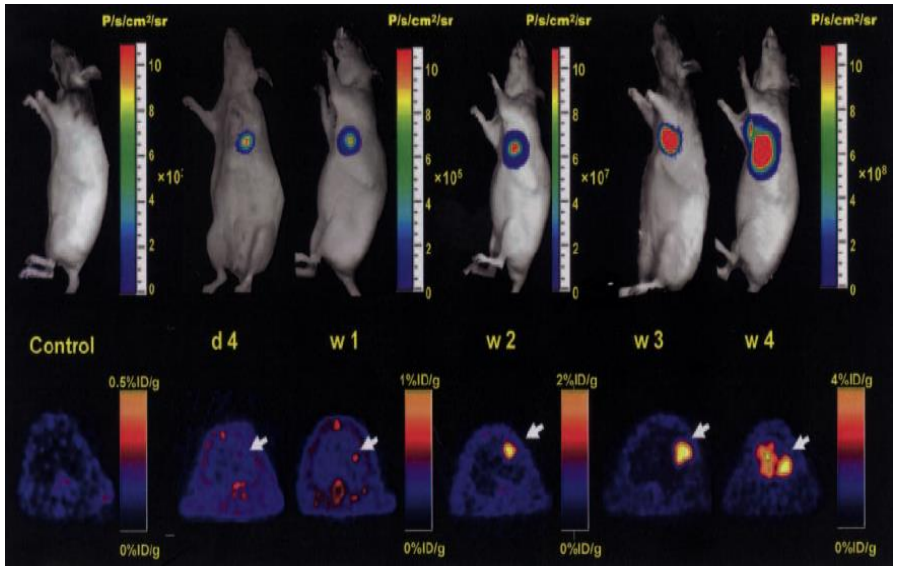
Stem Cell Foci Formation and Hematopoiesis



Transplantation of 250 Luc+ HSC into Lethally Irradiated Hosts

Cao *et al*, *Stem Cells*, 2004

Stem Cell Viability



In Vivo Visualization of ES Cell Survival, Proliferation, and Migration After Cardiac Delivery

Cao *et al*, *Circulation*, 2006